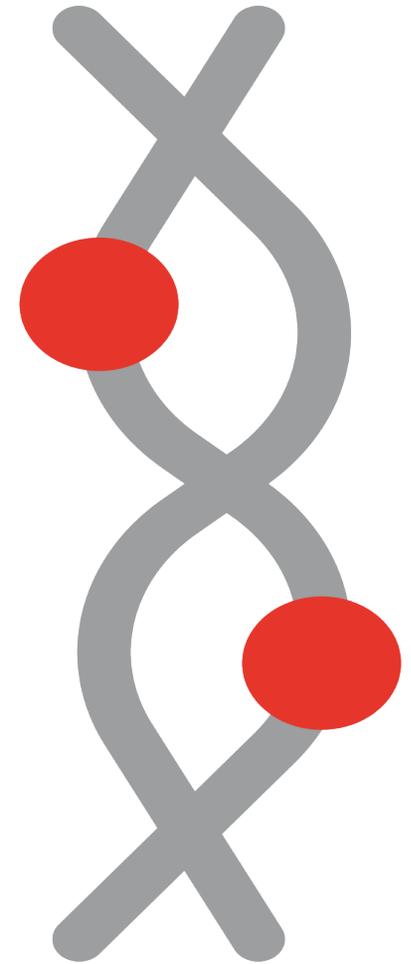


Gene Comprehensive Nutrigenomic Report

Accession Number: #####
Specimen Collected: ##/##/####
Specimen Received: ##/##/####
Report Generated: October 17, 2024
Specimen Type: Buccal Swab
Provider: #####
Patient Name: #####
Patient DOB: ##/##/####
Patient Gender: Female



Do not make any decisions about your health solely based on the information contained in this report.
Always consult with a licensed and experienced health practitioner when you receive this report.

– 32 – Female

(-/-) Normal Risk (-/+) Medium Risk (+/+) High Risk

rsID	Gene	Genetic Result	This column contains the targeted ingredients to support healthy gene activity of the specific genes listed.	Lifestyle Recommendations	Laboratory Recommendations
INFLAMMATORY					
rs2250656	C3	T/C (+/-)	Anti-Inflammatory Therapy: Curcumin, Omega-3 Fatty Acids, Resveratrol, Quercetin, Low Dose Naltrexone (LDN), CBD Oil	Consider Anti-inflammatory Diet and Lifestyle	<p>General Inflammatory Markers: Serum High Sensitivity C-Reactive Protein, Serum Iron and Ferritin, Erythrocyte Sedimentation Rate, Serum Complement C3, Serum Interleukin 6</p> <p>Lymphocyte Profile AND/OR Antibody Testing</p> <p>Additional Options: Adrenal Stress Profile, Sex Hormone Panel, Full Thyroid Panel, Food Allergy Panel, Comprehensive Micronutrient Testing, Microbial Titer (Candida, Epstein-Barr Virus, etc.), Toxic Metal Testing, Environmental Allergy Testing</p>
rs2569190	CD14	A/G (+/-)			
rs2069812	IL5	A/G (+/-)			
rs1800925	IL13	C/C (-/-)			
rs10181656	STAT4	C/G (+/-)			
rs1800795	IL6	C/G (+/-)			
rs1800629	TNF	G/G (-/-)			
rs231775	CTLA4	A/A (-/-)			
rs4795067	NOS2	A/G (+/-)	Inducible Nitric Oxide Synthase (iNOS) Activity, Anti-Infectives, Beta Glucans	Increased iNOS Activity May Promote Higher Levels of Inflammation	

– 32 – Female

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rsID	Gene	Genetic Result	This column contains the targeted ingredients to support healthy gene activity of the specific genes listed.	Lifestyle Recommendations	Laboratory Recommendations
EXTERNAL INFLAMMATORY					
Histamine Sensitivity					
rs10156191	AOC1	C/C (-/-)	Poor Ability to Break Down External Histamine		
Gluten Sensitivity					
rs2187668	HLA DQA1	C/T (+/-)	High Risk of Gluten and Casein Sensitivity, Broad Spectrum Enzyme	Consider Gluten and Dairy Avoidance	Food Allergy Panel if GI Inflammation Is Present
rs7454108	HLA DQB1	T/T (-/-)			
Microbiome Stability					
rs492602	FUT2	A/G (+/-)	Prebiotics and Probiotics Needed	Consider Consumption of Prebiotic and Probiotic Foods	
Vitamin D					
rs2228570	VDR	A/A (+/+)	Vitamin D, Vitamin K		Consider Checking Vitamin D Levels OR Comprehensive Micronutrient Testing
AUTOPHAGY					
rs510432	ATG5	T/T (-/-)	Curcumin, Lithium Orotate, D-Chiro-Inositol, Catechins, Resveratrol, Caffeine, 12 Hour Fasting	May Have Reduced Blood Sugar Control	Routine Blood Sugar, Insulin, and HbA1c
rs26538	ATG12	C/T (+/-)		Intermittent Fasting (12-15 Hours)	
rs10210302	ATG16L1	C/T (+/-)		Exercise Regularly	

– 32 – Female

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rsID	Gene	Genetic Result	This column contains the targeted ingredients to support healthy gene activity of the specific genes listed.	Lifestyle Recommendations	Laboratory Recommendations
MITOCHONDRIA					
rs1467568	SIRT1	A/G (+/-)	Pterostilbene, Resveratrol, Quercetin, NAD+, Coenzyme Q10, Pyrroloquinoline Quinone (PQQ), L-Carnitine, Ornithine, Magnesium, Calcium	Exercise Regularly Caloric Restriction	Organic Acid Testing
rs8192678	PPARGC1A	C/C (-/-)			
rs1937	TFAM	G/G (+/+)			
CoQ10					
rs1800566	NQO1	G/G (-/-)	Coenzyme Q10, Pyrroloquinoline Quinone (PQQ), Riboflavin		
Oxidative Stress					
rs6721961	NFE2L2	G/G (-/-)	Pterostilbene, Green Tea (Epigallocatechin Gallate), Turmeric, Sulforaphane, Endurance Exercise	Consume Antioxidant Rich Diet	
rs4880	SOD2	A/G (+/-)			
Antioxidants					
rs33972313	SLC23A1	C/C (-/-)	High Dose Vitamin C		
rs6994076	TTPA	A/A (-/-)	Vitamin E		

– 32 – Female

(-/-) Normal Risk (-/+) Medium Risk (+/+) High Risk

rsID	Gene	Genetic Result	This column contains the targeted ingredients to support healthy gene activity of the specific genes listed.	Lifestyle Recommendations	Laboratory Recommendations
METHYLATION					
Folate Metabolism					
rs1051266	SLC19A1	T/C (+/-)	Methyltetrahydrofolate (B9), Riboflavin (B2), Niacinamide (B3)		Complete Blood Count Serum and RBC Folate
rs2071010	FOLR1	G/G (-/-)			
rs70991108	DHFR	DEL/DEL (-/-)			
rs1076991	MTHFD1	C/T (+/-)			
rs1801133	MTHFR C677T	G/A (+/-)			
rs1801131	MTHFR A1298C	T/T (-/-)			
Vitamin B12 Metabolism					
rs526934	TCN1	A/A (-/-)	Methylcobalamin, Adenosylcobalamin		Serum Vitamin B12
rs1801222	CUBN	A/G (+/-)			
rs1801198	TCN2	C/C (-/-)			
rs1801394	MTRR	A/A (-/-)		Methylcobalamin (B12)	

– 32 – Female

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rsID	Gene	Genetic Result	This column contains the targeted ingredients to support healthy gene activity of the specific genes listed.	Lifestyle Recommendations	Laboratory Recommendations
HOMOCYSTEINE METABOLISM					
Remethylation					
rs1805087	MTR	A/A (+/+)	Methyltetrahydrofolate, Methylcobalamin, Methionine	Avoid Smoking and Heavy Alcohol Consumption	Plasma Methylation Profile OR Plasma Homocysteine Serum Vitamin B12
rs3733890	BHMT	A/G (+/-)	Choline, Trimethylglycine (Betaine)		Plasma Methylation Profile OR Plasma Homocysteine
Catabolism					
rs234706	CBS	G/A (+/-)	Methyltetrahydrofolate, Methylcobalamin, Pyridoxal 5'-Phosphate (B6), Choline, Trimethylglycine, Serine, N-Acetyl Cysteine	Avoid Smoking and Heavy Alcohol Consumption Consider Anti-Inflammatory Diet and Lifestyle	Plasma Methylation Profile OR Plasma Homocysteine
rs1021737	CTH	G/T (+/-)	N-Acetyl Cysteine, Glutathione, Pyridoxal 5'-Phosphate	Avoid Smoking and Heavy Alcohol Consumption Avoid Herbicides and Pesticides	

– 32 – Female

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rsID	Gene	Genetic Result	This column contains the targeted ingredients to support healthy gene activity of the specific genes listed.	Lifestyle Recommendations	Laboratory Recommendations
DETOXIFICATION					
rs8190955	GSR	G/G (-/-)	Riboflavin, Reduced Glutathione		
rs17883901	GCLC	G/G (-/-)	Glutathione		
rs1695	GSTP1	A/G (+/-)	N-Acetyl Cysteine (NAC), Glutathione	Avoid Herbicides and Pesticides Consider Pre-Anesthesia Glutathione	Whole Blood Glutathione
rs1801280	NAT2	C/C (+/+)	Silymarin, Alpha Lipoic Acid (ALA), P-5-P, Catechins	Avoid Industrial Carcinogens	

– 32 – Female

(-/-) Normal Risk (-/+) Medium Risk (+/+) High Risk

rsID	Gene	Genetic Result	This column contains the targeted ingredients to support healthy gene activity of the specific genes listed.	Lifestyle Recommendations	Laboratory Recommendations
NEUROTRANSMITTER					
rs4680	COMT	A/G (+/-)	Riboflavin (B2), Taurine, Choline, Trimethylglycine (TMG), Dimethylglycine (DMG), Methionine, SAME, Inositol, L-Methionine	High Risk of Depression/Anxiety	Neurotransmitter Metabolite Testing Consider PGx Testing
rs6323	MAOA	T/T (-/-)	Riboflavin (B2), Taurine, Choline, Trimethylglycine (TMG), Dimethylglycine (DMG), Methionine, SAME, Inositol, L-Methionine		
rs1799836	MAOB	T/C (+/-)			
rs3828275	GAD1	C/C (-/-)	Prescription Amantadine, Ketamine, Glycine, N-Acetyl-Cysteine (NAC), Zinc, Magnesium, Oxaloacetate, Elderberry, L-Theanine, Melatonin		
rs769407	GAD1	C/G (+/-)			
rs2274924	TRPM6	T/T (-/-)	Magnesium		

– 32 – Female

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rsID	Gene	Genetic Result	This column contains the targeted ingredients to support healthy gene activity of the specific genes listed.	Lifestyle Recommendations	Laboratory Recommendations
HEALTH PRECAUTIONS					
Thyroid Health					
rs4704397	PDE8B	A/G (+/-)	Iodine, Selenium, Increased Risk of Hypothyroidism		Thyroid Panel Urinary Iodine OR Comprehensive Micronutrient/Mineral Analysis
Estrogen Conversion					
rs4646	CYP19A1	C/C (+/+)	High Activity of Aromatase, Higher Risk of Excess Estrogen Production	Testosterone Therapy May Produce High Levels of Estrogen	Sex Hormones and Metabolites Panel including Progesterone, Testosterone, and Estrogen
Hypertension Risk					
rs4343	ACE	A/G (+/-)	Increased Risk of Salt Retention and Hypertension	Increased Risk of Hypertension and Preeclampsia Salt Restriction, Especially after Age 40	
Caffeine Sensitivity					
rs762551	CYP1A2	A/A (-/-)	Caffeine Metabolism: Slow Metabolizer (CC genotype), Intermediate Metabolizer (CA genotype), Rapid Metabolizer (AA genotype)		
Clot Risk					
rs6025	F5	C/C (-/-)	Increased Risk of Blood Clots	Be Cautious with (or Avoid) the Use of Estrogen and Vitamin K Supplementation as It May Lead to Increased Risk of Blood Clots	Prothrombin Time
rs3211719	F10	G/G (+/+)			

Summary for Optimal Wellness Genetic Insights Panel

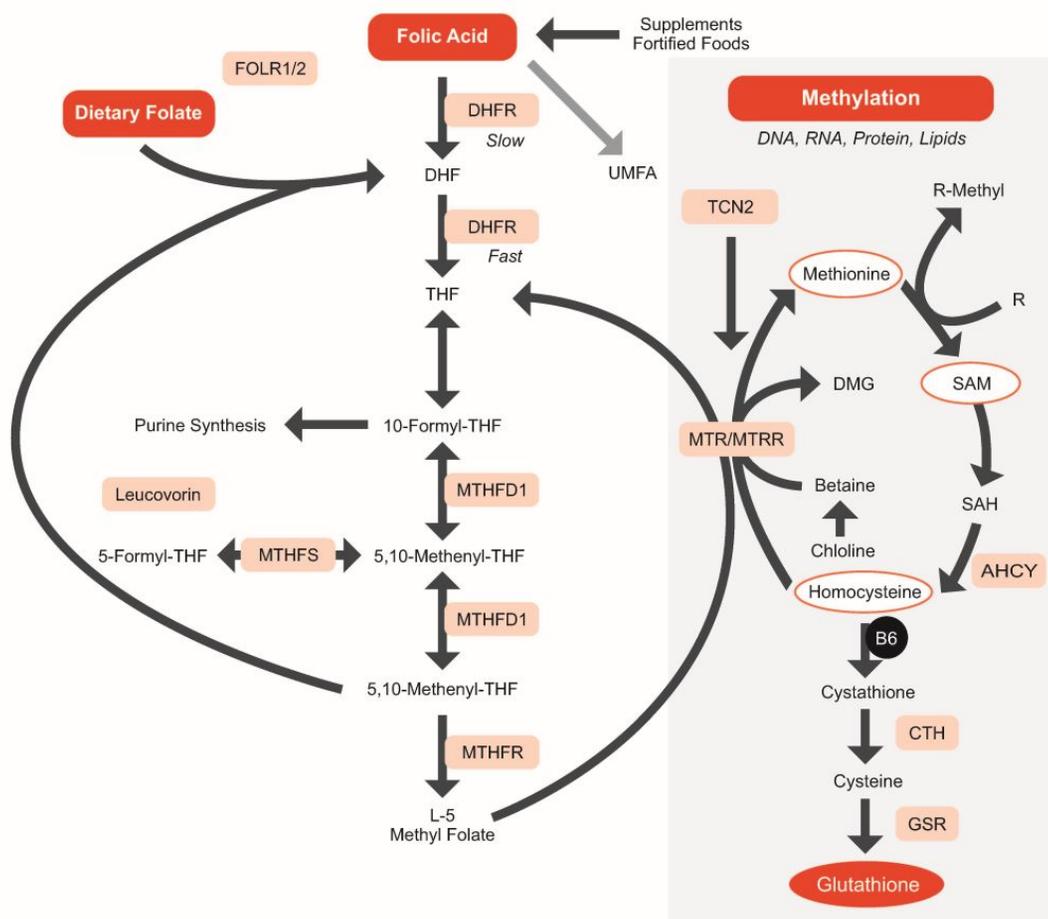
Lifestyle Recommendations

- Consider Anti-inflammatory Diet and Lifestyle
- Increased iNOS Activity May Promote Higher Levels of Inflammation
- Consider Gluten and Dairy Avoidance
- Consider Consumption of Prebiotic and Probiotic Foods
- May Have Reduced Blood Sugar Control
- Intermittent Fasting (12-15 Hours)
- Exercise Regularly
- Caloric Restriction
- Consume Antioxidant Rich Diet
- Avoid Smoking and Heavy Alcohol Consumption
- Consider Anti-Inflammatory Diet and Lifestyle
- Avoid Herbicides and Pesticides
- Consider Pre-Anesthesia Glutathione
- Avoid Industrial Carcinogens
- High Risk of Depression/Anxiety
- Testosterone Therapy May Produce High Levels of Estrogen
- Increased Risk of Hypertension and Preeclampsia
- Salt Restriction, Especially after Age 40
- Be Cautious with (or Avoid) the Use of Estrogen and Vitamin K Supplementation as It May Lead to Increased Risk of Blood Clots

Laboratory Recommendations

- General Inflammatory Markers: Serum High Sensitivity C-Reactive Protein, Serum Iron and Ferritin, Erythrocyte Sedimentation Rate, Serum Complement C3, Serum Interleukin 6
- Lymphocyte Profile AND/OR Antibody Testing
 - Additional Options: Adrenal Stress Profile, Sex Hormone Panel, Full Thyroid Panel, Food Allergy Panel, Comprehensive Micronutrient Testing, Microbial Titer (Candida, Epstein-Barr Virus, etc.), Toxic Metal Testing, Environmental Allergy Testing
- Food Allergy Panel if GI Inflammation Is Present
- Consider Checking Vitamin D Levels OR Comprehensive Micronutrient Testing
- Routine Blood Sugar, Insulin, and HbA1c
- Organic Acid Testing
- Complete Blood Count
- Serum and RBC Folate
- Serum Vitamin B12
- Plasma Methylation Profile OR Plasma Homocysteine
- Whole Blood Glutathione
- Neurotransmitter Metabolite Testing
- Consider PGx Testing
- Thyroid Panel
- Urinary Iodine OR Comprehensive Micronutrient/Mineral Analysis
- Sex Hormones and Metabolites Panel including Progesterone, Testosterone, and Estrogen
- Prothrombin Time

METHYLATION



Methylation

- Involves the addition of a methyl group (CH₃)
- Regulates gene expression and repression
- Reduces or removes toxins that eliminate essential nutrients
- Provides nutrients needed for processes such as detoxification, immune regulation, gut health

Methionine

- Used in protein formation and stabilization
- Elevated levels are associated with risk for coronary heart disease, stroke & neurological diseases

Glutathione

- Important for chemical detoxification & proper mitochondrial functioning
- Genes relevant for production include: AHCY, CTH, CGTP1, GSTM1, GSTM3, GSR, MTRR & MTR

5-Methyl Folate

- Important for dopamine and serotonin formation, detoxification and mitochondrial strength
- Genes relevant for production include: DHFR, FOLR1/2, MTHFD1, MTHFR, MTHFS

Homocysteine

- Elevated levels are associated with risk for coronary heart disease, stroke, neurological diseases
- Variants in the methylation pathway can be associated with increased/decreased levels

FOLATE

FOOD SOURCES



Eggs



Citrus Fruits



Leafy Greens



Legumes



Broccoli



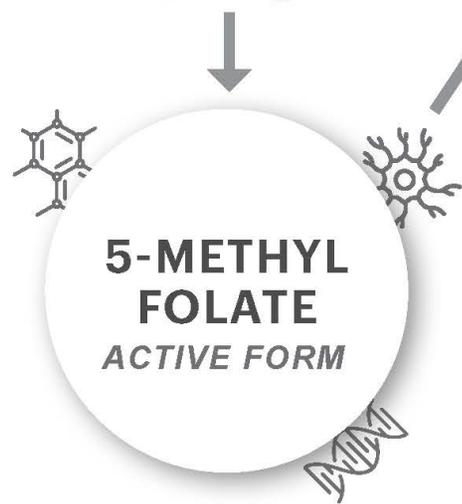
Brussel Sprouts



Breads, Cereal,
Pastas, Rice



**ABSORBABLE
FOLATE**
NATURAL B9



**5-METHYL
FOLATE**
ACTIVE FORM

FUNCTIONS (OR BENEFITS AS YOU AGE)

- Maintains structure & function of proteins
- Maintains structure & function of DNA
- Facilitates DNA replication, neurotransmitter production & detoxification

DEFICIENCY CAUSES

- Neural tube defects
- Cardiovascular disease
- Memory problems
- Depression
- Insomnia
- Irritability

VITAMIN B12

FOOD SOURCES



Eggs



Seafood – clams, trout, salmon, tuna



Meats – liver, beef, ham, chicken



Low-fat milk products



Swiss cheese



Low-fat yogurt



Nori/Seaweed



Fortified Foods



Nutritional Yeast



**ABSORBABLE
VITAMIN B12**



FUNCTIONS (OR BENEFITS AS YOU AGE)

- Formation and maintenance of red blood cells (RBCs)
- Facilitates DNA synthesis
- Regulates homocysteine levels (decreases)
- Facilitates neurological functioning

DEFICIENCY & PATHWAY ALTERATIONS

- Increased production of homocysteine
- Decreased breakdown of homocysteine
- Circadian Rhythm Problems
- Cancers
- Memory-related disorders
- Cardiovascular diseases
- Fatigue
- Poor balance
- Mood disorders

VITAMIN D

FOOD SOURCES



Tuna



Mushrooms



Eggs



Mackerel



Cheese



Milk Products



BENEFITS AS YOU AGE



Lower Risk
of Fractures



Improves
Heart Function



Supports
Immune System



Speeds
Wound Healing

DEFICIENCY CAUSES

- Bone Pain
- Arthritis
- Obesity
- Backache
- Depression
- Diabetes
- Hypertension
- Osteoporosis
- Heart Disease
- Skin Conditions

MITOCHONDRIA

WAYS TO INCREASE MITOCHONDRIAL FUNCTIONING



Build muscle mass
(strength conditioning)



Decrease toxin exposure
(metals, persistent organic pollutants)



Vitamins
Glutathione
CoQ10
a-lipoic acid + acetyl-l-carnitine
Resveratrol
NAC
Vitamin E
PQQ
Ginkgo biloba
Proanthocyanidins



FUNCTIONS (OR BENEFITS AS YOU AGE)

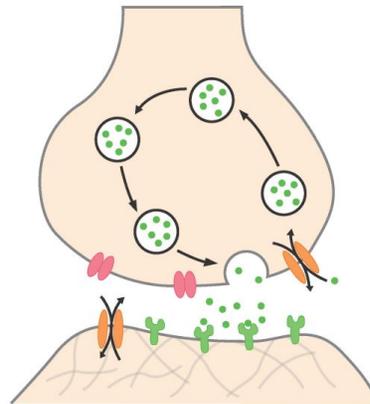
- Energy production for growth, movement & homeostasis
- Programmed cell death for dysfunctional, old cells
- Calcium signaling for neuron excitability, neurotransmission & plasticity (strengthening)

DEFICIENCY CAUSES

- Early Aging
- Neurological Disorders
- Diabetes
- Developmental Issues
- Psychiatric Disorders
- Cardiovascular Diseases
- Fatigue

NEUROTRANSMITTERS & PATHWAY

TRANSMIT INFORMATION FOR ESSENTIAL PROCESSES SUCH AS DIGESTION, BREATHING, HEARTBEAT, MOVEMENT, PAIN REGULATION ETC.



RELEVANT GENES

- **HTR2, TPH2, SLC6A4, MAO-A** genes are important in the synthesis, breakdown, transport and/or functioning of serotonin
- **COMT, MAO-A, MAO-B** genes are important for the breakdown of serotonin, norepinephrine and/or dopamine
- The **DBH** gene is important for norepinephrine synthesis
- The **GAD1** gene is important for GABA synthesis
- Variants in **COMT, MAO-A, MAO-B** and **GAD1** genes have been associated with mood, anxiety and focus issues

WAYS TO INCREASE LEVELS



Aerobic Exercise



Dietary Factors



Mediation/Yoga

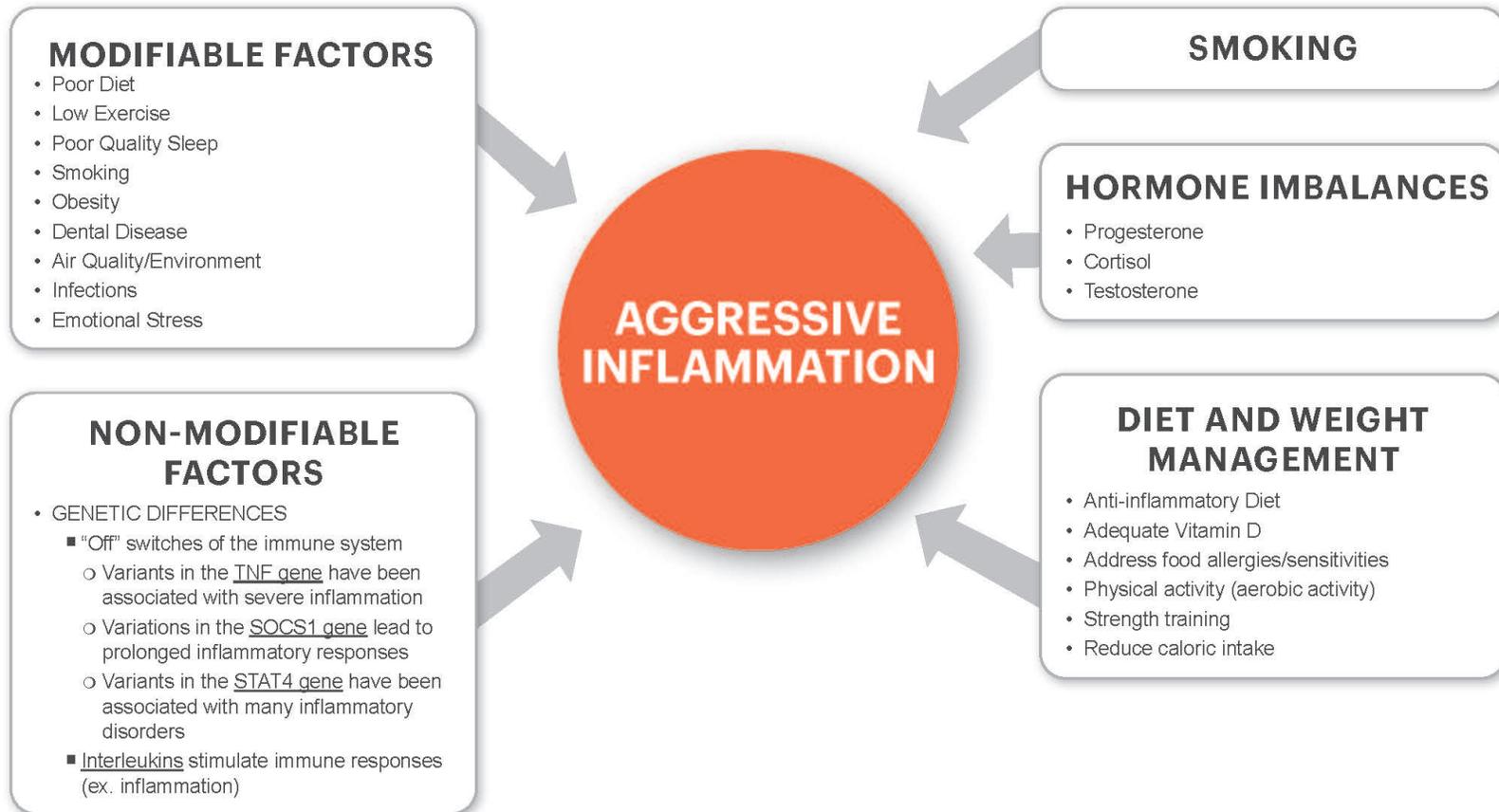


Increase Sun Exposure

ANTI-INFLAMMATORY

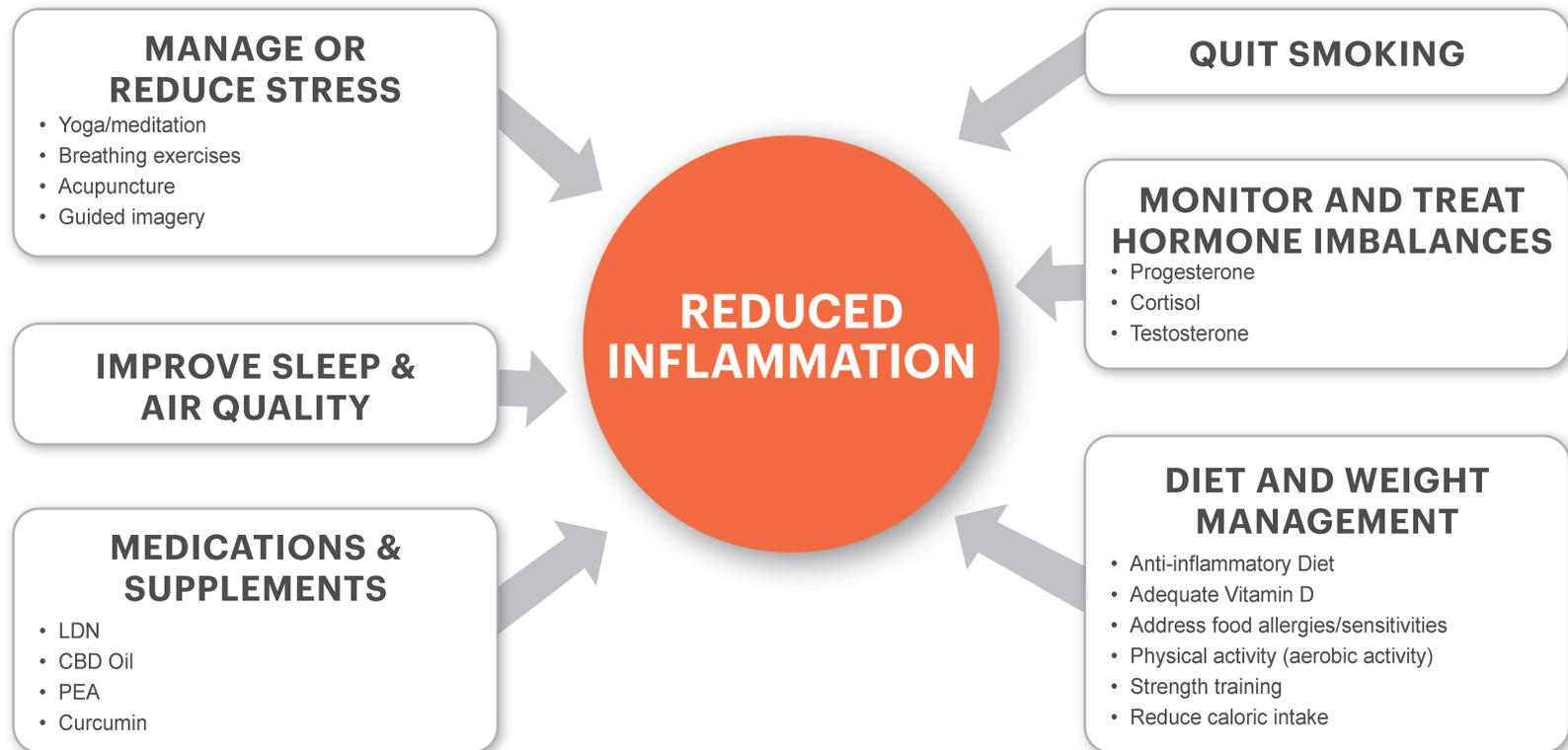
AN IMMUNE SYSTEM RESPONSE TRIGGERED BY HARMFUL STIMULI
(EX. PATHOGENS, DAMAGED CELLS, TOXIC COMPOUNDS, IRRADIATION)

DRIVERS OF INFLAMMATION



ANTI-INFLAMMATORY

WAYS TO REDUCE INFLAMMATION

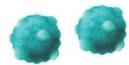


THE IMMUNE SYSTEM & AUTOIMMUNITY

WHAT DOES THE IMMUNE SYSTEM DO?

Prevent or limit infections by distinguishing between healthy and unhealthy cells

KEY PLAYERS & RELEVANT GENES



CYTOKINES

(ex. IL family, TNF-a)

- Helps with immune cell growth, activation, and function
- Interleukins (IL2, IL4, IL5, IL6, IL13, IL23R, IL2RA) stimulate the immune response
- SOCS1 & TNF are involved in cytokine signaling for the inflammatory response



LYMPHOCYTES

(ex. B, T & Natural Killer cells)

- Identify & kill infected cells
- Produces antibodies to fight future infections
- IDO1, CTLA4 & CD14 are involved in the suppression of T-cells
- C3, STAT4 & TRAF1 activate, form and/or differentiate T-cells

IMMUNE AGGRESSION

The immune system begins to attack healthy tissue

COMMON SYMPTOMS



Fatigue



Hair loss



Achy muscles



Inflammation



Skin rashes



Pain



Low-grade fever



Numbness and tingling in hands and feet



Trouble concentrating

MALFUNCTIONS LEAD TO

- Chronic inflammation
- Allergic reactions
- Immune aggressive diseases (Inflammatory bowel disease, skin & neurological disorders)

LOW-INFLAMMATORY

FOODS TO EAT



Fruits: strawberries, blueberries, cherries, oranges



Fatty fish: salmon, mackerel, tuna, sardines



Spices - turmeric, ginger



Green leafy vegetables & tomatoes



Dark chocolate



Olive oil



LOW-INFLAMMATORY DIET

FOODS TO AVOID



Soda & other sugar-sweetened drinks



Dairy products



Fried foods



Red & Processed meats (hotdogs, sausage)



Refined carbohydrates: white bread, pastries



Margarine, shortening, lard

BENEFITS



Reduces inflammation



Reduces risk for cardiovascular disease & Type II diabetes

DETOXIFICATION

GLUTATHIONE IN DETOXIFICATION

Relevant genes for production are AHCY, CTH, GSTP1, GSTM1, GSTM3, GSR, MTRR & MTR

WHY IS IT IMPORTANT?



Maintains health by protecting the body from toxins



Regulates cell production and programmed cell death



Critical role in chemical detoxification



Vital for proper mitochondrial function



WAYS TO INCREASE GLUTATHIONE

- Limit alcohol intake
- N-acetyl-cysteine (NAC)
- Glutathione therapies
- (ie. IV Glutathione, Glutathione suppository, Liposomal Glutathione)
- Include whey in diet, unless allergic or intolerant
- Methylation Support - if necessary

SUPEROXIDES & ANTIOXIDANTS

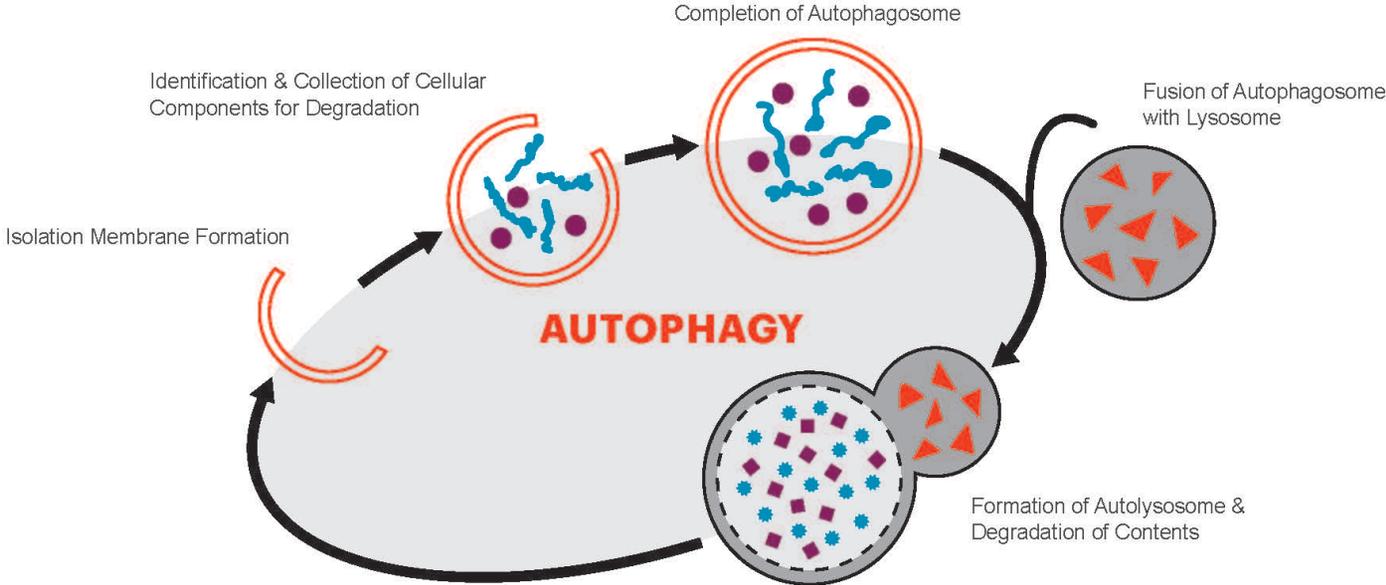
- SOD1, SOD2, SOD3 genes are important to transform superoxides to protect against mitochondrial damage
- Reactive Oxygen Species (ROS) can damage mitochondria and cause cell death.
- Antioxidants such as Vitamin A, Vitamin C and Vitamin E act as a defense against ROS

DEFICIENCY CAUSES

- Auto-immune diseases
- Cardiovascular diseases
- Neurodegenerative diseases
- Cell death
- Poor mitochondrial function

AUTOPHAGY

VARIANTS IN THE ATG GENES HAVE BEEN ASSOCIATED WITH CELLULAR BLOCKAGE



DEFECTS LEAD TO:

- Neurodegenerative Diseases
- Aging
- Heart Disease
- Developmental Disorders
- Type II Diabetes
- Insulin Resistance
- Fatty Liver
- Cancers

WAYS TO INCREASE

-  Intermittent fasting or low-calorie diet
-  Routine Exercise
-  Ketogenic diets (high fat, low carbs)
-  Medications & Supplements
D-Chiro Inositol (B8)
Metformin

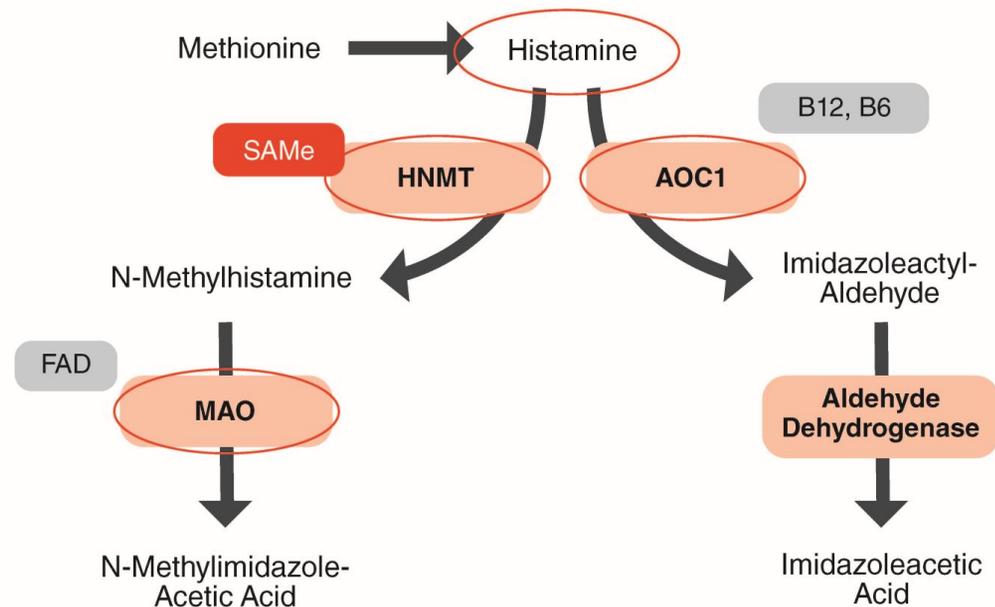
HISTAMINE

HISTAMINE

- Natural substance found in various foods

IMPLICATIONS

- Metabolic Enzymes: amine oxidases (ex. AOC1, MAO, DAO) & HNMT
- High histamine & low amine oxidase activity is associated with:
 - Diarrhea
 - Headaches
 - Nose congestion
 - Asthma
 - Hypotension
 - Arrhythmia
 - Flushing
 - Urticaria (hives)
 - Pruritus (itchy skin)
- Dietary histamine can be rapidly detoxified by amine oxidases, whereas persons with low amine oxidase activity are at risk of histamine toxicity



AOCI & HNMT POLYMORPHISM HISTAMINE

LOW HISTAMINE LEVEL FOODS



Meats & Fish
fresh meat (ex. chicken, turkey, pork and red meat), fresh fish (ex. hake, trout, plaice)



Milk substitutes
(Coconut milk, rice milk)



Cream cheese, butter



Egg yolk



Fresh fruits
(with the exception of strawberries)



Most cooking oils



Most leafy herbs



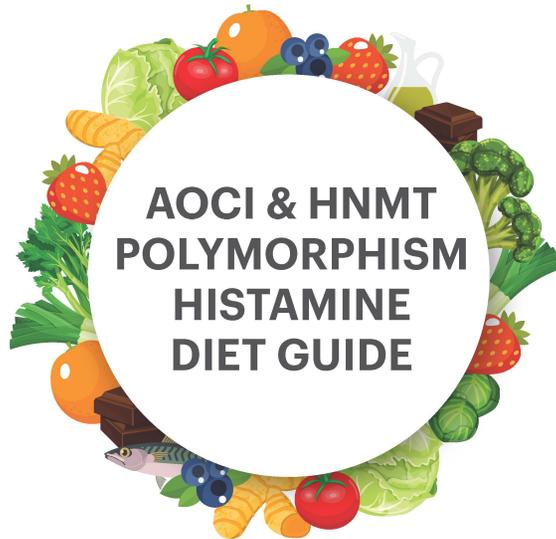
Fresh vegetables



Beverages
(non-citric fruit juices, herbal teas)



Grains



AOCI & HNMT POLYMORPHISM HISTAMINE DIET GUIDE

HIGH HISTAMINE LEVEL FOODS



Egg whites



Processed, cured, smoked and fermented meats/fish (lunch meat, bacon, sausage, pepperoni, canned tuna)



Leftover meat
(After meat is cooked, the histamine levels increase due to microbial action as the meat sits)



Dairy products: All fermented milk products (ex. aged cheeses, yogurt, buttermilk, kefir)



Beverages (Black Tea, alcohol)



Chocolate, cocoa



Fruits (oranges, grapefruit, lemons, lime, berries, dried fruit)



Vegetables (spinach, tomatoes, eggplant)



Artificial food colors and preservatives



Fermented & vinegar-containing foods (sauerkraut, kombucha, pickles, relishes, ketchup, prepared mustard)



Spices (cinnamon, chili powder, cloves, nutmeg, curry powder, cayenne)

SELENIUM

WAYS TO INCREASE LEVELS



Brazil nuts



Low-fat milk products



Meats & seafood – fish (tuna, halibut, sardines), ham, shrimp, beef, liver, chicken, turkey



Boiled eggs



Wheat germ, Brewer's yeast



Whole grains (unless gluten free)



Supplements



ABSORBABLE SELENIUM

FUNCTIONS



Role in proper thyroid function & thyroid hormone metabolism



Role in DNA synthesis



Role in reproduction



Protection from infection & oxidative damage

DEFICIENCY VS HIGH INTAKE

Deficiency

- Cardiovascular disorders
- Developmental issues
- Thyroid disorders
- Joint & bone issues
- Infertility issues
- Cancers

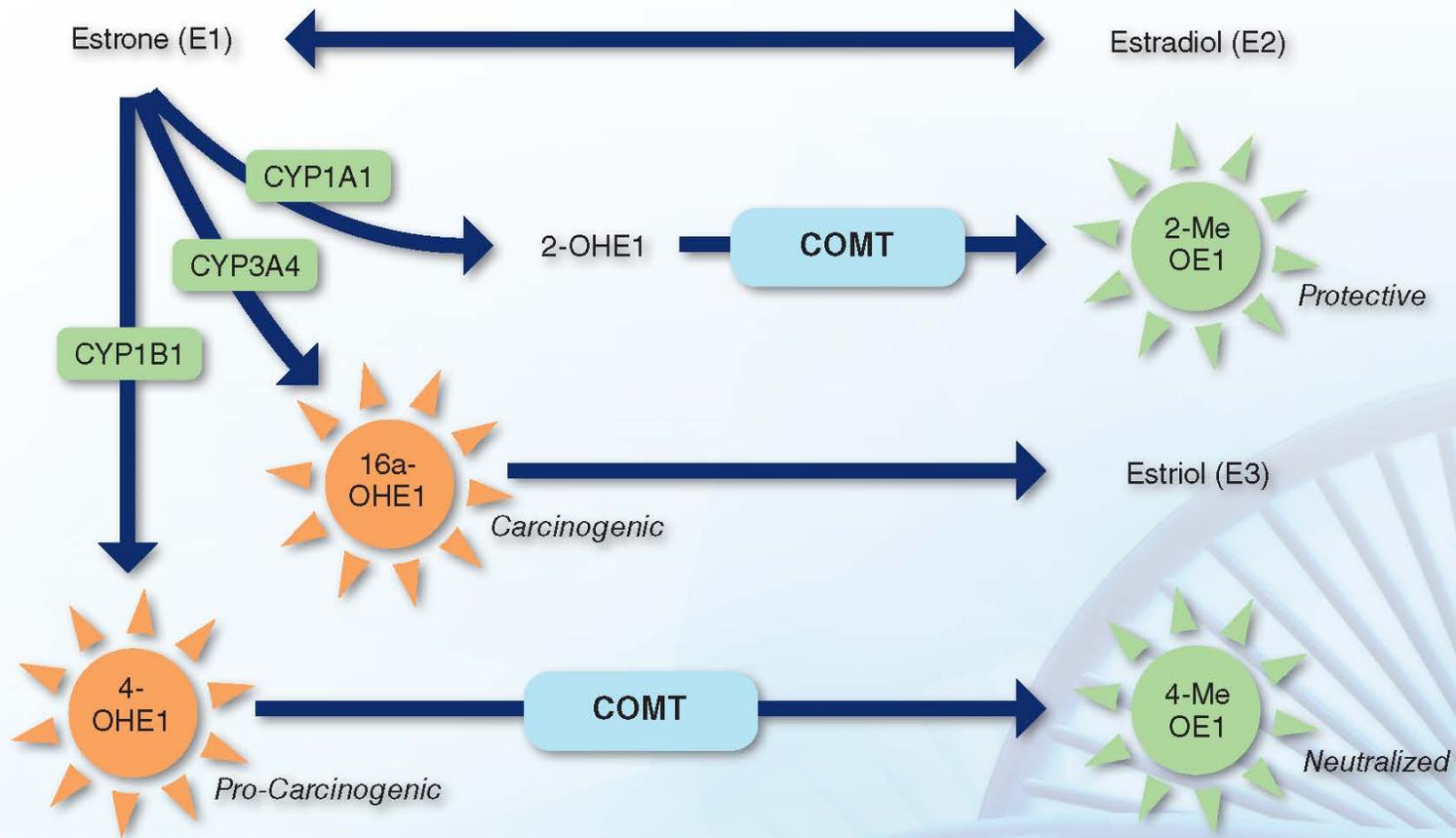
High intake

- Metallic taste in mouth
- Garlic odor of breath
- Hair and nail loss or brittleness
- Nervous system abnormalities
- Nausea
- Diarrhea
- Skin rashes
- Fatigue
- Irritability



**SELENOMETHIONINE & SELENOCYSTEINE
ACTIVE FORM**

Phase I and II - Estrogen Metabolism



HYPERTENSION RISK FACTOR

HIGH BLOOD PRESSURE

- Ranges
 - Normal: 120/80
 - Range of concern: 140/90 or higher
- Risk factors: high salt diet, high alcohol intake, stress, little potassium intake, alcohol & tobacco use, obesity, genetics/family history, age, lack of physical activity
- Uncontrolled high blood pressure has been associated with an increased risk for cardiovascular diseases and stroke

AGT & ACE GENES

Variants have been associated with an increased risk for:



Salt retention



Kidney issues



Preeclampsia



Poor sports performance



Hypertension & other cardiovascular issues

LIFESTYLE CHANGES



Limit salt intake



Angiotensin II Receptor Blockers ("sartans")



Weight management & routine exercise



Mediterranean diet



Quit smoking



Heart-healthy diet/
Low-sodium diet/
DASH diet

DASH DIET

FOOD TO EAT



Fruits & vegetables



Egg whites



Whole grains (unless gluten free)



Nuts & nut butters



Lean, skinless meat & fish (salmon, trout, herring)



Legumes



Olive oils high in polyphenols



Low-fat or fat-free dairy products



**DASH
DIET**

BENEFITS



Improves heart health



Improves and/or reduces risk for hypertension, heart disease and stroke

FOODS TO AVOID AND/OR LIMIT



Red meat



Fried foods



Sweets



Processed meats - deli meat, hotdogs, sausage, bacon



Sugar-sweetened beverages



Fats/oils - Butter, margarine, tropical oils (coconut, palm)



High-salt foods

Gene Information Key

rsID	Gene	"-" variant	"+" variant
rs4343	ACE	A	G
rs10156191	AOC1	C	T
rs26538	ATG12	T	C
rs10210302	ATG16L1	C	T
rs510432	ATG5	T	C
rs3733890	BHMT	G	A
rs2250656	C3	C	T
rs234706	CBS	A	G
rs2569190	CD14	G	A
rs4680	COMT	G	A
rs1021737	CTH	G	T
rs231775	CTLA4	A	G
rs1801222	CUBN	G	A
rs4646	CYP19A1	A	C
rs762551	CYP1A2	A	C
rs70991108	DHFR	DEL	INS
rs3211719	F10	A	G
rs6025	F5	C	T
rs2071010	FOLR1	G	A
rs492602	FUT2	A	G
rs3828275	GAD1	C	T
rs769407	GAD1	G	C
rs17883901	GCLC	G	A
rs8190955	GSR	G	A
rs1695	GSTP1	A	G
rs2187668	HLA-DQA1	C	T
rs7454108	HLA-DQB1	T	C
rs1800925	IL13	C	T
rs2069812	IL5	A	G

rsID	Gene	"-" variant	"+" variant
rs1800795	IL6	C	G
rs6323	MAOA	T	G
rs1799836	MAOB	C	T
rs1076991	MTHFD1	C	T
rs1801131	MTHFR: A1298C	T	G
rs1801133	MTHFR: C677T	G	A
rs1805087	MTR	G	A
rs1801394	MTRR	A	G
rs1801280	NAT2	T	C
rs6721961	NFE2L2	G	T
rs4795067	NOS2	A	G
rs1800566	NQO1	G	A
rs4704397	PDE8B	G	A
rs8192678	PPARGC1A	C	T
rs1467568	SIRT1	A	G
rs1051266	SLC19A1	T	C
rs33972313	SLC23A1	C	T
rs4880	SOD2	G	A
rs10181656	STAT4	C	G
rs526934	TCN1	A	G
rs1801198	TCN2	C	G
rs1937	TFAM	C	G
rs1800629	TNF	G	A
rs2274924	TRPM6	T	C
rs6994076	TTPA	A	T
rs2228570	VDR	G	A

Definitions

AUTOPHAGY	
ATG12	Autophagy-related 12 protein is part of the core autophagy machinery inside the cell. Autophagy, a form of cellular "recycling" is necessary for many cell functions. ATG12 is specifically involved in turning off the innate immune response. Mutations in the ATG12 gene are predicted to lead to increased activity of the innate immune response, and overall inflammation.
ATG16L1 rs10210302	The ATG16L1 (autophagy related 16 like 1) gene encodes a protein that is part of a major protein complex essential for autophagy, a process of digesting cellular components for nutrient sensing and cellular regulation. The polymorphism rs10210302 occurs in the promoter region of the gene, and a comprehensive study has linked the T allele with Crohn's disease, an inflammatory bowel disease.
ATG5	The ATG5 (autophagy-related 5) gene is an important intracellular mediator of the autophagy response, which is essential for maintaining homeostasis. The polymorphism rs510432 occurs in the promoter region of ATG5, and individuals homozygous for the C allele have been shown to have increased mRNA expression of ATG5. Additionally, individuals homozygous for the C allele are at an increased risk for developing childhood asthma, but they have a reduced risk for developing sepsis. Individuals who are heterozygous or homozygous for the T allele have been shown to have reduced levels of C-reactive protein.
DETOXIFICATION	Detoxification enzymes are responsible for clearing environmental chemicals and metabolites from our body. Accumulation of these chemicals and by-products can damage intracellular biochemical functions. Alterations in these systems can have a significant negative effect on the nervous system and immune systems functions. These polymorphisms can result in decreased "quality of life" and even decreased "life-span".
CTH	The CTH (cystathionine gamma-lyase) gene encodes an enzyme that catalyzes the last step in the transsulfuration of L-methionine to L-cysteine. More specifically, it converts cystathionine, derived from methionine, into cysteine, which is utilized in the liver to synthesize glutathione, a ubiquitous antioxidant. As a result, CTH has an important role in glutathione production. The polymorphism rs1021737 results in an isoleucine substitution for a serine residue in the enzyme at position 403. The T allele, which encodes the isoleucine variant, has been associated with an accumulation of homocysteine.
GCLC rs17883901	The GCLC (glutamate-cysteine ligase catalytic subunit) gene encodes the first and rate-limiting enzyme in glutathione biosynthesis. Glutathione is a potent, nonprotein antioxidant that has an important role in protecting cells from oxidative stress and xenobiotics. Therefore, a reduction in biosynthetic capacity through reduction of GCLC activity results in reduced antioxidant capacity and activity of enzyme that use glutathione as a cofactor, such as glutathione transferase and glutathione peroxidase. The polymorphism rs17883901 occurs in the promoter region of the gene, and mechanistic studies have found that the variant encoded by the A allele reduces promoter activity by as much as 60% in cell models. Furthermore, A allele carriers had significantly increased risk for coronary heart disease and heart attack. Lastly, plasma glutathione levels were reduced in individuals with the AA genotype.
GSR rs8190955	The GSR (glutathione-disulfide reductase) gene encodes a riboflavin-dependent enzyme that reduces oxidized glutathione (GSSH) to its antioxidant form (GSH). Therefore, GSR is essential for maintaining adequate glutathione levels and cellular antioxidant capacity. The polymorphism rs8190955 results in a cysteine substitution for an arginine residue at position 153. Mechanistic studies have shown that the A allele, which encodes a cysteine residue, produces an enzyme that is less stable than the version produced by the G allele. Furthermore, the instability was predicted to reduce enzyme function, and the A allele has been associated with obstructive heart defects and hereditary anemia.
GSTP1	The GSTP1 (glutathione S-transferase pi 1) gene encodes a cytosolic enzyme that has a keystone role in cellular detoxification. It conjugates cytotoxic and carcinogenic substances to glutathione for elimination, thereby aiding in antioxidant defense and preserving DNA integrity. The polymorphism rs1695 results in a valine substitution for an isoleucine residue in the enzyme at position 105, which is a region of the protein that is known to undergo several post-translational modifications. Mechanistic studies have shown that the protein produced by the G allele, which encodes a valine residue, has reduced substrate binding capacity and enzymatic activity. Numerous clinical studies have shown that the GG genotype is a risk factor for asthma, especially when individuals are exposed to environmental toxins, such as cigarette smoke or traffic-related air pollution. Additionally, the G allele is associated with increased risk for heart failure, and the frequency of the G allele is decreased in populations of older, living adults, suggesting it does not confer increased longevity.
NAT2 rs1801280	The NAT2 (N-acetyltransferase 2) gene encodes an enzyme that catalyzes the transfer of an acetyl group from acetyl coenzyme A to compounds with aromatic amines, heterocyclic amines, or hydrazine structures. Therefore, NAT2 has an important role in the metabolism and elimination of a large number of pharmaceutical drugs and environmental toxins. The polymorphism rs1801280 results in a threonine substitution for an isoleucine residue at position 114. The C allele, which encodes a threonine residue, defines the NAT2*5B haplotype, which has been found to have slow acetylation activity. Consistent with this status, carriers of the C allele are at increased risk for various types of cancer and adverse drug reactions. Additionally, increased toxin exposure, such as frequent or intense smoking, can further increase these risks.
NFE2L2 rs6721961	The NFE2L2 (NFE2 like bZIP transcription factor 2) gene encodes a transcription factor, known as NRF2, that has a crucial role in the regulation of a network of antioxidant genes. NRF2 activates expression of genes with a conserved promoter sequence called the antioxidant response elements (ARE). Genes with an ARE include superoxide dismutase (SOD), catalase (CAT), and glutathione peroxidase (GPX), etc. Therefore, NRF2 is a master regulator of oxidant and antioxidant balance. The polymorphism rs6721961 occurs in the promoter region of NFE2L2, and mechanistic studies have found that the variant encoded by the T allele has reduced promoter activity and mRNA levels. Consistent with these findings, carriers of the T allele have been shown to have lower total antioxidant capacity. T allele carriers had less SOD, CAT, GPX, and glutathione activity. Furthermore, T allele carriers are at increased risk for insulin resistance and vascular stiffness.

SOD2	The SOD2 (superoxide dismutase 2) gene encodes a mitochondrial matrix enzyme that uses iron and manganese to convert superoxide, a byproduct of the electron transport chain, to hydrogen peroxide and oxygen. Moreover, SOD2 participates in phase 1 detoxification to initiate the transformation of highly reactive oxygen species to a less reactive intermediary metabolite. Hydrogen peroxide, the intermediary metabolite produced by SOD2 activity, requires further detoxification by phase 2 enzymes, such as catalase or glutathione peroxidase. Nevertheless, SOD2 function is crucial to initiate the detoxification process of reactive oxygen species in the mitochondria, aiding in protection against oxidative damage. The polymorphism rs4880 results in a valine substitution for an alanine residue in the enzyme at amino acid position 16, which occurs in the mitochondrial targeting sequence. Mechanistic studies have shown that the A allele, which encodes a valine residue, has incomplete transfer to the mitochondria due to a conformational change in the targeting sequence. Furthermore, studies in animal models have shown that the enzyme encoded by the A allele has decreased formation of active enzyme in the mitochondrial matrix, and clinical studies have found individuals with the AA genotype to have less SOD2 activity. Consistently, individuals with the AA genotype have been shown to have decreased plasma total antioxidant status and increased markers of oxidative stress and lipid peroxidation. However, implementation of a healthy diet and exercise intervention has been shown to reduce markers of lipid peroxidation in those with the AA genotype. Lastly, the AA genotype was associated with increased risk for coronary heart disease.
ESSENTIAL MINERALS	The polymorphisms in this panel will identify any potential weakness of absorption or delivery of your essential minerals..
TRPM6	The TRPM6 (transient receptor potential cation channel subfamily M member 6) gene encodes a membrane transporter that is essential for magnesium homeostasis. TRPM6 is expressed in gut and kidneys, and it is needed for magnesium absorption. The polymorphism rs2274924 results in a nucleotide change that causes translation of the transporter to terminate prematurely, resulting in a truncated version of the transporter that is approximately 500 amino acids shorter than the full-length protein. Carriers of the C allele, which encodes the truncated protein, are at increased risk for hypomagnesemia, with individuals with the CC genotype at greater risk.
ESSENTIAL VITAMINS	The polymorphisms in this panel will identify any potential weakness of absorption, conversion or delivery of your essential vitamins.
SLC23A1	The SLC23A1 (solute carrier family 23 member 1) gene encodes a sodium-dependent vitamin C transporter required for the absorption of vitamin C and its distribution to tissues throughout the body. The polymorphism rs33972313 results in a leucine substitution for a valine residue at amino acid position 264, which occurs in a transmembrane domain of the transporter. The T allele, which encodes a leucine residue, has been associated with decreased levels of vitamin C, and levels were further reduced in individuals with the TT genotype.
TTPA rs6994076	The TTPA (alpha tocopherol transfer protein) gene encodes a carrier protein that binds alpha-tocopherol, the predominate form of vitamin E in the body, and facilitates its export from the liver to circulation. The polymorphism rs6994076 occurs in the promoter region of the gene, and mechanistic studies found that the variant encoded by the T allele reduces transcription of TTPA. In clinical studies, carrier of the T allele had lower levels of circulating vitamin E.
VDR rs2228570	The VDR (vitamin D receptor) gene encodes a receptor for vitamin D3 that is highly expressed in the intestines. VDR is a member of the nuclear hormone receptor superfamily, so when activated by vitamin D, it can impact transcription of many genes involved in mineral metabolism, cell proliferation, and immune activation. The polymorphism rs2228570, sometimes termed FokI for the restriction enzyme that can detect it, results in a threonine substitution for a methionine residue in the first codon of the protein, altering the translation start site. As a result, translation of the receptor produced by the A allele, which does not contain the FokI restriction site (f) and encodes a methionine residue, is 427 amino acids in length, whereas the receptor produced by the G allele, which does contain the FokI restriction site (F) and encodes a threonine residue, is three amino acids shorter. Mechanistic studies indicate that the shorter variant encoded by the G allele has greater capacity to bind vitamin D and more transcriptional activity in response to vitamin D. Consistent with these findings, A allele carriers were less responsive to vitamin D supplementation, and A allele carriers were shown to have reduced calcium absorption and bone mineral density. Furthermore, vitamin D supplementation was less effective at reducing inflammatory markers in carriers of the A allele, and the A allele is associated with risk for celiac disease and type 2 diabetes.
ESTROGEN METABOLISM AND CLEARANCE	The conversion of estrogen and its' metabolites is essential to effective safe estrogen treatment. These SNPs will identify your potential for increased production of possible carcinogenic forms of estrogen
CYP19A1	The CYP19A1 (cytochrome P450 family 19 subfamily A member 1) gene encodes a monooxygenase enzyme termed aromatase. Aromatase catalyzes the last step in the conversion of androgens to estrogen. The polymorphism rs4646 occurs in the 3' untranslated region, suggesting that it might affect gene expression by altering mRNA stability. Furthermore, the C allele has been associated with higher circulating estrogen levels, indicating increased aromatase activity.
GASTROINTESTINAL	
AOC1	The AOC1 (amine oxidase copper-containing 1) gene encodes for the diamine oxidase (DAO) enzyme, which is one of two enzymes that breaks down pro-inflammatory amines such as histamine and putrescine. DAO is active in intestinal mucosal cells, and a deficiency of its activity results in the accumulation of high levels of histamine, which can cause a wide range of neurological, gastrointestinal, and epidermal disorders. The polymorphism rs10156191 results in a methionine substitution for a threonine residue in the enzyme at position 16. The T allele, which encodes the methionine variant, results in an enzyme with lower metabolic capacity than the enzyme encoded by the C allele, possibly resulting in reduced ability to break down histamine.
FUT2	The FUT2 (fucosyltransferase 2) gene encodes an enzyme involved in the synthesis of histoblood group antigens (HBGA), which are found on the intestinal mucosa and various bodily fluids. HBGA are oligosaccharide molecules, and in the intestinal mucosa, they act as an attachment site and nutrient source for intestinal bacteria. The polymorphism rs492602 is in near perfect linkage disequilibrium with rs601338, meaning that the alleles are nonrandomly associated and inherited together. Therefore, the G allele for rs492602 indicates the inheritance of the minor allele for rs601338, which results in a stop gain mutation that produces a truncated version of FUT2 that is unable to secrete the oligosaccharide molecules. As a result, individuals of the GG genotype for rs492602 are considered "non-secretors". Carriers of the G allele were found to have compositional and functional changes to the gut microbiota and reduced microbial diversity. Furthermore, G allele carriers had increased susceptibility for inflammatory bowel disease.

HLA-DQA1	The HLA-DQA1 (major histocompatibility complex, class II, DQ alpha 1) gene encodes a cell surface protein that plays a central role in the function of the immune system and the development of autoimmune disease. HLA-DQA1 is a class II, human leukocyte antigens (HLA), which are expressed on the surface of antigen presenting cells where HLA can bind antigens or substances that induce an immune response for recognition by T cells. HLA-DQA1 encodes a component of HLA-DQ2, a serotype or distinct variation among the HLA structure that determines its antigenic complements. More specifically, the T allele for the polymorphism, rs2187668, can be used to identify HLA-DQ2.5, a high-risk factor for gluten sensitivity and celiac disease. Consistently, genome-wide association studies have found the T allele is associated with celiac disease.
HLA-DQB1 rs7454108	The HLA-DQB1 (major histocompatibility complex, class II, DQ beta 1) gene encodes a cell surface protein that plays a central role in the function of the immune system and the development of autoimmune disease. HLA-DQB1 is a class II, human leukocyte antigens (HLA), which are expressed on the surface of antigen presenting cells where HLA can bind antigens or substances that induce an immune response for recognition by T cells. More specifically, the C allele for the polymorphism, rs7454108, can be used to identify HLA-DQ8, a serotype or distinct variation among the HLA structure that determines its antigenic complements. HLA-DQ8, for example, is a high-risk factor for gluten sensitivity and celiac disease.
HEALTH PRECAUTIONS	
ACE	The ACE (angiotensin-converting enzyme) gene encodes a protein that plays a crucial role in regulating blood pressure and maintaining electrolyte balance. It converts angiotensin I to the active form, angiotensin II, which leads to vasoconstriction and elevated blood pressure. The polymorphism rs4343 confers an insertion/deletion of a small DNA sequence in the gene. Carriers of the G allele display increased ACE activity and elevated plasma levels of angiotensin II. Additionally, carriers of the G allele are more prone to blood pressure spikes when consuming high-salt diets than individuals with the AA genotype. Heterozygous individuals display an intermediate phenotype.
CYP1A2	The CYP1A2 (cytochrome P450 family 1 subfamily A member 2) gene encodes a monooxygenase enzyme that mainly functions in the liver. It catalyzes the metabolism of about 10% of clinically used drugs that are metabolized by CYP enzymes, including caffeine. Additionally, it metabolizes some endogenous compounds, such as melatonin and estradiol. The A allele of rs762551 was found to have higher CYP1A2 enzyme activity with exposure to smoking or heavy coffee consumption. In contrast, the C allele was found to be associated with lower enzyme activity. rs762551 was also associated with caffeine consumption. Specifically, the AA genotype may predispose an individual to have higher coffee intake.
Factor V	The F5 gene encodes for coagulation factor V, an essential component of the blood coagulation cascade. Specifically, it serves as a cofactor for the prothrombinase activity of factor Xa that results in the activation of prothrombin to thrombin. The polymorphism rs6025 is a well-known missense mutation known as the Leiden mutation. The T allele of rs6025 encodes for a variant in which glutamine is substituted for arginine at position 506. Carriers of the T allele are at an elevated risk for venous thromboembolism and related conditions with the risk being even higher in individuals homozygous for the T allele.
Factor X	The F10 (coagulation factor X) gene encodes a vitamin K-dependent factor in the blood coagulation cascade. Factor X (FX) plays an important role in blood clotting, as both the intrinsic and extrinsic coagulation pathways converge on FX activation. FX is translated as a preproprotein, which is processed to a mature and activated version that converts prothrombin to thrombin. The polymorphism rs3211719 occurs in intron 1, and genome-wide association studies have found that the G-allele is associated with decreased prothrombin time and increased levels of factor VII, which initiates the extrinsic coagulation pathway. Additional studies have shown that the G allele is associated with increased factor VII antigen and factor VII coagulant activity, suggesting that clotting propensity is increased.
HOMOCYSTEINE METABOLISM	
BHMT rs3733890	The BHMT (betaine-homocysteine S-methyltransferase) gene encodes an essential enzyme that consume betaine, or trimethylglycine, to convert homocysteine to dimethylglycine and methionine. Therefore, BHMT both detoxifies homocysteine and generates methionine needed to maintain methylation capacity. It is primarily expressed in the liver and kidneys. The polymorphism rs3733890 results in a glutamine substitution for an arginine residue at position 239. The A allele, which encodes a glutamine residue, results in more partitioning of choline, a precursor of betaine, for phosphatidylcholine synthesis via the cytidine diphosphate (CDP)-choline pathway, suggesting that less betaine is available for detoxification of homocysteine and regeneration of methionine. Likewise, carriers of the A allele have been shown to have reduced levels of betaine and dimethylglycine. Furthermore, folate was shown to be a less effective treatment to lower homocysteine levels in carrier of the A allele with hyperhomocysteinemia. Nevertheless, increased intake of choline in A allele carriers can increase flux to betaine synthesis to support BHMT activity. In summary, A allele carriers may benefit from increased choline or betaine intake, especially when managing high levels of homocysteine.
CBS rs234706	The CBS (cystathionine beta-synthase) gene encodes an enzyme that catalyzes the first step in the transsulfuration pathway. More specifically, CBS, a pyridoxal 5'-phosphate-dependent enzyme, consumes serine to convert homocysteine to cystathionine, which is further catabolized to generate substrate for glutathione synthesis. Therefore, homocysteine clearance and glutathione synthesis converge on the function of CBS. The polymorphism rs234706 results in a nucleotide substitution in exon 8. Carriers of the G allele have been found to have higher levels of homocysteine and lower levels of cystathionine and betaine, consistent with reduced CBS activity. Furthermore, individuals with the GG genotype had higher plasma homocysteine following the ingestion of a methionine load, and individuals with the GG genotype were less responsive to folate supplementation to lower homocysteine levels. Lastly, the GG genotype is associated with increased risk for coronary artery disease.

MTR	The MTR (5-methyltetrahydrofolate-homocysteine methyltransferase) gene encodes a metabolic enzyme that catalyzes the remethylation of homocysteine to methionine; therefore, the enzyme is also referred to as methionine synthase. The reaction requires both active vitamin B9 (methyltetrahydrofolate) and vitamin B12 (methylcobalamin), and the enzyme works in close coordination with 5-methyltetrahydrofolate-homocysteine methyltransferase reductase (MTRR), which regenerates MTR to a functional state. The polymorphism rs1805087 results in a glycine substitution for an aspartic acid residue in the enzyme at position 919, which is located in the binding site for accessory proteins needed to regenerate the enzyme to its active form. Numerous studies indicate that the G allele variant, which encodes a glycine residue, results in increased enzyme function. For example, the G allele was associated with reduced levels of homocysteine and increased levels of DNA methylation, suggesting accelerated conversion of homocysteine to methionine. However, increased MTR activity also results in accelerated consumption of methylcobalamin, and predictably, the GG genotype has been associated with vitamin B12 deficiency. Nevertheless, A allele carriers may have decreased methionine synthesis, which is essential for production of S-adenosyl methionine (SAME) and maintenance of methylation capacity.
INFLAMMATORY	This Enzyme category has significant effects on the inflammatory state of a person's body. Polymorphisms in these specific enzymes will significantly increase the levels of inflammation in the body. By supplementing these enzyme deficiencies, the patient will effectively reduce inflammatory damage to the body.
C3 rs2250656	The C3 (complement C3) gene encodes an abundant protein in the complement cascade, a major component of the innate immune system. C3 is secreted into the bloodstream, and activation of C3 is essential for both classical and alternative complement activation. The polymorphism rs2250656 occurs in the fourth intron, and carriers of the T allele were shown to have increased plasma levels of C3 and C-reactive protein, two markers of inflammation. Additionally, increasing consumption of n-6 PUFAs, which are known to have a proinflammatory effect, further increased plasma levels of C3 in individuals with the TT genotype. Lastly, T allele carriers had a 2-fold higher risk for metabolic syndrome compared to individuals with the CC genotype, and the T allele was associated with severe COVID-19, suggesting that the T allele confers increased complement activity and inflammation.
CD14 rs2569190	The CD14 (CD14 molecule) gene encodes a crucial determinate of the innate immune response and protector from atopy. CD14 is an endotoxin receptor that is expressed on the surface of monocytes and macrophages. The polymorphism rs2569190 occurs in the promoter region of the gene, and mechanistic studies have shown that the variant encoded by the A allele is preferentially bound by RNA polymerase, suggesting that there is increased transcription of the A allele variant. Consistent with this mechanism, carriers of the A have been shown to have increased levels of CD14. Clinical studies have found that while A allele carriers have basal IgE levels that are lower than G allele carriers, IgE levels rise to a greater extent in A allele carriers when exposed to various forms of endotoxins. Moreover, A allele carriers are at increased risk for asthma or allergic disease when highly exposed to endotoxins, suggesting that A allele carriers are at increased risk for environmentally instigated inflammation. Lastly, A allele carriers may have increased risk for cardiovascular disease and inflammatory bowel disease, both of which are characterized by increased inflammation.
CTLA4	The CTLA4 (cytotoxic T-lymphocyte associated protein 4) gene encodes a cell-surface receptor that acts as an important inhibitor of T cell activity and T cell-mediated immune responses. Therefore, CTLA4 has a crucial role in T cell homeostasis and self-tolerance, the loss of which can lead to the development of autoimmunity. The polymorphism rs231775 results in an alanine substitution for a threonine residue in the receptor at position 17, and the G allele, which encodes the alanine variant, results in decreased expression and cell surface localization of CTLA4 and increased proliferative response of T cells. Furthermore, clinical studies found that the G allele was associated with a variety of autoimmune diseases, such as autoimmune thyroid disease, type 1 diabetes, and rheumatoid arthritis. Additionally, the GG genotype was associated with positivity for insulinoma associated-2 autoantibodies (IA-2A) in patients with type 1 diabetes.
IL13	The IL13 (interleukin 13) gene encodes a cell signaling molecule that has a central role in the regulation of allergic inflammation. IL13 has a crucial role in the activation of Th2 immune responses, including the stimulation of B cells to synthesize IgE, a type of immunoglobulin that mediates allergic reactions. The polymorphism rs1800925 is located in the promoter region the gene, and functional studies have shown that the variant encoded by the T allele increases transcription of IL13 by increasing the binding of STAT transcription factors to the promoter region. Consistent with these studies, the TT genotype was found to be more prevalent in individuals with asthma and atopic dermatitis, and it has been associated with increased risk of sensitization to food and outdoor allergens. Additionally, the TT genotype was associated with increased risk for appendicitis, and the T allele was associated with increased risk for chronic obstructive pulmonary disease (COPD).
IL5	The IL5 (interleukin 5) gene encodes a cytokine that promotes the growth, differentiation, and activation of eosinophils. Therefore, it has an important role in Th2 immune responses and the development of allergic disease, which includes the promotion of IgE production and eosinophil response. The polymorphism rs2069812 is associated with a Th2-dominant autoimmune thyroid disease known as Graves' disease (GD). Carriers of the G allele have been shown to be at increased risk for GD as well as Graves' ophthalmopathy. Additionally, genome-wide association studies have linked the G allele with asthma, whereas the A allele has been associated with remission of GD and reduced risk of asthma.
IL6	The IL6 (interleukin 6) gene encodes a cytokine with pro- and anti-inflammatory functions depending on the context; however, continuous and dysregulated synthesis of IL6 plays a key role in both acute and chronic inflammation. Additionally, IL6 is a crucial link between the innate and adaptive immune systems, and the gene is known to be mainly regulated at the transcriptional level. The polymorphism rs1800795 occurs in the promoter region of the gene, and mechanistic studies showed that the C allele yielded lower levels of IL6 than the G allele. As a result, the G allele has the potential to increase the potency of any immunological response stimulated by IL6, and the G allele has been associated with conditions of high inflammation. For example, the G allele may increase the risk for rheumatoid arthritis, IBS, and various types of liver disease. Additionally, the GG genotype may increase the risk for psoriasis, poly cystic ovarian syndrome, and pre-term birth, all conditions associated with high levels of circulating IL6.

NOS2	The NOS2 (nitric oxide synthase 2) gene encodes an isoform of an enzyme that can be induced by pro-inflammatory agents like lipopolysaccharide and cytokines to produce nitric oxide (NO), a potent signaling molecule that can influence immune activation, inflammation, and cell survival. NOS2 can be conditionally activated in many cell types, but it is especially important for the function of immune cells, like macrophages. While NO is needed to defend against invading pathogens and unregulated cellular proliferation, excessive NO can damage healthy tissue. Carriers of the G allele for rs4795067 have an increased ratio of nitrite to nitrate in plasma. Because NO is quickly metabolized to nitrite in the body, nitrite is considered to be a measure of NO reserve, suggesting that carriers of the G allele have increased production of NO. Additionally, the G allele is associated with psoriasis, and NOS2 expression has been shown to be increased in psoriatic lesions. In summary, studies suggest that G allele carriers for rs4795067 produce increased amounts of NO, which can lead to inflammation.
STAT4	The STAT4 (signal transducer and activator of transcription 4) gene encodes a transcription factor that responds to extracellular growth factors and cytokines. It is present in the cytosol, and following cytokine signaling at the cell surface, STAT4 is phosphorylated and translocates to the nucleus, initiating the expression of cytokines, receptors, and signaling factors. The polymorphism rs10181656 occurs in the third intron, and cell-based experiments indicate that the G allele results in overexpression of STAT4, suggesting that the variant regulates gene expression. The G allele has also been associated with increased risk for numerous autoimmune diseases, such as rheumatoid arthritis, type 1 diabetes, lupus, and autoimmune thyroid disease. Furthermore, multiple studies support that there is additional disease risk for individuals carrying two copies of the G allele (GG genotype) compared to single allele carriers (GC genotype).
TNF-?	The TNF-? (tumor necrosis factor alpha) gene encodes an important pro-inflammatory cytokine that is mainly secreted by activated macrophages and monocytes. TNF-? functions in a plethora of biological functions from pathogen defense to tissue remodeling as it plays a role in cell survival, growth, and differentiation. Given its pro-inflammatory nature, dysregulation of TNF-? is also associated with numerous pathological conditions. The polymorphism rs1800629 occurs in the promoter region of the gene, and mechanistic studies have shown that the variant encoded by the A allele results in increased transcription and secretion of TNF-?. Therefore, the A allele has the potential to increase the potency of any immunological response stimulated by TNF-?. Consistent with this mechanism, the A allele has been associated with increased susceptibility to autoimmune disease, such as asthma, Graves' disease, psoriatic arthritis, and lupus. Additionally, individuals with the AA genotype were less responsive to TNF-? blockers for the treatment of autoimmune disease, and A allele carriers experienced fewer anti-inflammatory benefits of physical activity compared to individuals with the GG genotype. Similarly, obese individuals with the AA genotype were less responsive to a hypocaloric diet high in polyunsaturated fats as an intervention to improve metabolic markers. In summary, these results suggest that the inflammation generated by increased transcription of TNF-? caused by the A allele variant is markedly resistant to repression.
METABOLIC RISK FACTOR	The polymorphisms in this category relate to increase risk of developing metabolic syndromes including diabetes, fatty liver, hypothyroidism and insulin resistance.
PDE8B rs4704397	The PDE8B (phosphodiesterase 8B) gene encodes a enzyme that catalyzes the hydrolysis of cAMP, a second messenger crucial for cellular energy sensing. The polymorphism rs4704397 occurs in the first intron, and numerous studies have found that the A allele is associated with increased levels of TSH, consistent with hypothyroidism. Additionally, the A allele has been associated with sub-clinical hypothyroidism, hypothyroidism, and infertility.
METHYLATION	Methylation is a primary biochemical process in the body that involves the addition of a "methyl" chemical group to a vitamin or neurotransmitter. The addition of the "methyl" group allows for very specific biochemical interactions. Poor "methylation" function alters the effectiveness, delivery and function of many vitamins and important chemicals in the cell.
CUBN rs1801222	The CUBN (cubilin) gene encodes a receptor for intrinsic factor-cobalamin (Cbl-IF) complexes, and it is essential for intestinal absorption of vitamin B12. The polymorphism rs1801222 results in a phenylalanine substitution for a cysteine residue at position 253 in the protein. The A allele, which encodes a phenylalanine residue, is associated with reduced plasma levels of vitamin B12 and increased levels of homocysteine. Additionally, the A allele has been associated with risk for neural tube defects.
DHFR rs70991108	The DHFR (dihydrofolate reductase) gene encodes an enzyme essential for converting folic acid, a synthetic form of folate that is common in supplements and fortified foods, to tetrahydrofolate, a usable form of folate. The polymorphism rs70991108 results in a 19-bp deletion in the first intron, and mechanistic studies indicate that the deletion reduces translation and stability of the enzyme. Individuals homozygous for the deletion had higher levels of circulating unmetabolized folic acid, compared to carriers of the full length gene. Furthermore, cognitive function was reduced in deletion carriers, suggesting the 19-bp deletion reduces DHFR activity and folate metabolism. Lastly, the deletion allele has been associated with neural tube defects, pre-term delivery, and hepatic toxicity in response to treatment with methotrexate, which competitively inhibits DHFR activity. In summary, individuals with the 19-bp deletion should prioritize natural forms of folate, instead of folic acid, to maintain productive folate metabolism.
FOLR1	The FOLR1 (folate receptor alpha) gene produces a folate receptor that is responsible for transporting folate and its derivatives into cells. Variations in this gene can affect the delivery of folate in the bloodstream to cells. A study found that individuals who were heterozygous for the polymorphism rs2071010 had elevated serum folate levels compared to those with the GG genotype, suggesting that the A allele may reduce FOLR1 function. Additionally, individuals with the AA genotype may be at increased risk for elevated homocysteine levels.
MTHFD1	The MTHFD1 (methylene tetrahydrofolate dehydrogenase, cyclohydrolase, and formyltetrahydrofolate synthetase 1) gene encodes an enzyme that is essential for folate metabolism. The enzyme catalyzes three sequential steps in folate metabolism, utilizing separate catalytic domains in the protein. It converts 1) tetrahydrofolate (THF) to 10-formylTHF 2) 10-formylTHF to 5,10-methenylTHF, and 3) 5,10-methenylTHF to 5,10-methyleneTHF, which can then be converted to the bioactive form of folate, 5-methylTHF (MTHF), by methylenetetrahydrofolate reductase (MTHFR). The polymorphism rs1076991 occurs in the promoter region of the gene, and mechanistic studies found that the variant encoded by the T allele had a 60% reduction in transcription rate, suggesting that T allele carriers produce significantly less enzyme and MTHF. Congruently, the T allele has also been associated with risk for heart attack.

MTHFR rs1801131	The MTHFR (metylenetetrahydrofolate reductase) gene encodes a metabolic enzyme that catalyzes the conversion of 5,10-metylenetetrahydrofolate to 5-methyltetrahydrofolate (MTHF), the bioactive form of folate. Folate is a crucial mediator of one-carbon metabolism, which is necessary for a plethora of biochemical functions, such as nucleotide biosynthesis, amino acid metabolism, epigenetic maintenance, and oxidative defense. The polymorphism rs1801131, sometimes referred to as A1298C, results in an alanine substitution for a glutamate residue in the enzyme at position 429, which occurs near the binding site for an allosteric inhibitor, S-adenosyl-L-methionine (SAME). Cell-based assays have shown that the enzyme produced by the G allele, which encodes an alanine residue, reduces MTHFR activity by about 30% compared to the enzyme produced by the T allele. Consistent with these findings, the GG genotype has been associated with increased risk for ischemic stroke and infertility due to decreased sperm production in men. Furthermore, individuals heterozygous for rs1801131 and rs1801133, another polymorphism in the MTHFR gene, have a more severe clinical phenotype that is similar to the AA genotype for rs1801133. Lastly, despite the prevalence of both minor alleles, the genotype combination rs1801131 GG and rs1801133 AA is nearly nonexistent in the population, suggesting it confers a significant genetic disadvantage.
MTHFR rs1801133	The MTHFR (metylenetetrahydrofolate reductase) gene encodes a metabolic enzyme that catalyzes the conversion of 5,10-metylenetetrahydrofolate to 5-methyltetrahydrofolate (MTHF), the bioactive form of folate. Folate is a crucial mediator of one-carbon metabolism, which is necessary for a plethora of biochemical functions, such as nucleotide biosynthesis, amino acid metabolism, epigenetic maintenance, and oxidative defense. The polymorphism rs1801133, sometimes referred to as C677T, results in a valine substitution for an alanine residue in the enzyme at position 222, which occurs near the binding site for a cofactor and the substrate, FAD and 5,10-metylenetetrahydrofolate respectively. Mechanistic studies have shown that the enzyme produced by the A allele, which encodes a valine residue, has reduced thermal stability and 55% reduced activity compared to the enzyme produced by the G allele. Consistent with these results, carriers of the A allele were found to have decreased levels of folate and increased levels of homocysteine. As a result, carriers of the A allele are at risk for neural tubes defects, vascular disease, stroke, migraine, depression, and infertility. Furthermore, individuals heterozygous for rs1801133 and rs1801131, another polymorphism in the MTHFR gene, have a more severe clinical phenotype that is similar to the AA genotype for rs1801133. Lastly, despite the prevalence of both minor alleles, the genotype combination rs1801131 GG and rs1801133 AA is nearly nonexistent in the population, suggesting it confers a significant genetic disadvantage.
MTRR rs1801394	The MTRR (5-methyltetrahydrofolate-homocysteine methyltransferase reductase) gene encodes an enzyme that regenerates methionine synthase, an enzyme encoded by the MTR gene, to a functional state. As a results, MTRR is also known as methionine synthase reductase, and it has a key role maintaining folate-methionine homeostasis. The polymorphism rs1801394 results in a methionine substitution for an isoleucine residue in the enzyme at position 22, and biochemical studies have found that the enzyme encoded by the G allele has a lower affinity for its target, methionine synthase, than the enzyme encoded by the A allele. These results suggest that carriers of the G allele, which produces a protein containing a methionine residue, may have reduced regeneration of methionine synthase and reduced conversion of homocysteine to methionine. Congruently, numerous studies have shown that G allele carriers have elevated homocysteine levels, which can be mediated by supplementation with folate. The G allele was also found to be a risk factor for neural tube defects and Down syndrome. Lastly, risk of neural tube defects was increased in G allele carriers who were also deficient in vitamin B12.
SLC19A1	The SLC19A1 (solute carrier family 19 member 1) gene encodes a folate transporter known as reduced folate carrier (RFC). RFC mediates cellular uptake of folate and folate derivatives, including antifolate pharmaceuticals. Folate is an essential nutrient that supplies a methyl group to support important biochemical functions, such as DNA synthesis and substrate methylation. For example, folate, with the help of vitamin B12, supplies the methyl group needed to convert homocysteine to methionine. The polymorphism rs1051266 results in an arginine substitution for a histidine residue in the transporter at position 27, which occurs in a transmembrane domain. Individuals with the CC genotype were found to have lower levels of plasma folate compared to individuals with the TT genotype, suggesting that the C allele, which encodes the arginine variant, produces a less efficient transporter. Additionally, individuals with the CC genotype for rs1051266 and the TT genotype for rs1801133, a variant in the MTHFR gene, were found to have higher levels of homocysteine. The C allele has been associated with delayed memory ability and increased susceptibility for neural tube defects. Lastly, carriers of the C allele may be less responsive to treatment with methotrexate, and individuals with the CC genotype may be at increased risk for ischemic stroke.
TCN1	The TCN1 (transcobalamin 1) gene encodes various isoforms of a carrier protein that binds vitamin B12 (cobalamin). The isoforms are differentially glycosylated, and they dimerize to form a vitamin B12-binding protein called haptocorrin. Haptocorrin protects vitamin B12 from the acidic environment of the stomach and transports it to the small intestine, where it can be bound by intrinsic factor. It is also estimated that haptocorrin carries 70-80% of vitamin B12 in circulation. However, unlike transcobalamin encoded by the TCN2 gene, haptocorrin mainly delivers vitamin B12 to the liver. The polymorphism rs526934 occurs in the eighth intron, and carriers of the G allele have been shown to have lower vitamin B12 levels compared to individuals with the AA genotype.
TCN2	The TCN2 (transcobalamin 2) gene encodes a carrier protein that binds vitamin B12 (cobalamin) and delivers it to all tissues. Around 30% of circulating vitamin B12 is bound to TCN2. The polymorphism rs1801198 results in an arginine substitution for a proline residue in the protein at position 259, which occurs in the binding region for vitamin B12. In individuals with adequate vitamin B12 status, carriers of the G allele, which encodes an arginine residue, had less vitamin B12-bound TCN2 than C allele carriers. A large meta-analysis also reported that individuals with the GG genotype had significantly lower concentrations of vitamin B12-bound TCN2 and higher concentrations of homocysteine, a functional indicator of vitamin B12 status, compared to individuals with the CC genotype. Lastly, carriers of the C allele were shown to have lower levels of methylmalonic acid, which is converted to succinyl CoA in a vitamin B12-dependent reaction, suggesting that G allele carriers may have reduced levels of vitamin B12.
MITOCHONDRIA	The mitochondrial enzymes are responsible for energy production from the mitochondria. The mitochondria is known as the "powerhouse" of the cell and produces over 90% of the energy for a cell. The mitochondrial respiratory chain (also known as the electron transport chain) is where these 4 protein complexes are found. Polymorphic alterations in these enzymes reduce the energy output of the mitochondria and leads to symptoms of chronic fatigue, cognitive deficiency, exercise intolerance, low metabolic rate, muscle weakness, poor healing and higher rates of sleep disorders and mood abnormalities.

NQO1 rs1800566	The NQO1 (NAD(P)H quinone dehydrogenase 1) gene encodes a riboflavin-dependent enzyme that protects against oxidative stress. Moreover, it regenerates the antioxidant capacity of CoQ10 by reducing it. The polymorphism rs1800566 results in a serine substitution for a proline residue at position 187, which occurs in the FAD-binding site. The A allele, which encodes a serine residue, produces a variant with reduced stability due to reduced ability to bind FAD, a necessary co-factor. Because CoQ10 is sensitive to oxidative stress, molecular studies suggest that NQO1 has a role in maintaining CoQ10 status, and a clinical study found an association with NQO1 and CoQ10 status and response to supplementation. Lastly, molecular studies have found that NQO1 function is also dependent on adequate levels of riboflavin.
PPARGC1A rs8192678	The PPARGC1A (PPARG coactivator 1 alpha) gene encodes a transcriptional coactivator, termed PGC-1?, that enhances mitochondrial biogenesis and function. PGC-1? activity leads to the transcription of TFAM, which translocates to the mitochondrial matrix where it stimulates mitochondrial DNA replication and expression of other mitochondrial gene needed for replication. The polymorphism rs8192678 results in a serine substitution for a glycine at position 487, and the T allele encodes a serine residue. Individuals with the TT genotype were found to have reduced mitochondrial DNA copy number, less endurance exercise capacity, and increased risk of metabolic dysfunction, suggesting that mitochondrial content is decreased. Additionally, carriers of the T allele may have increased risk of polycystic ovary syndrome (PCOS), which often coincides with metabolic dysfunction.
SIRT1 rs1467568	The SIRT1 (sirtuin 1) gene encodes a nicotinamide adenine dinucleotide (NAD+) and zinc-dependent histone deacetylase. SIRT1 is an important epigenetic regulator that responds to metabolic and oxidative stress to activate genes related to mitochondrial biogenesis and ATP production. SIRT1 activates PGC-1? through the SIRT1/PGC-1? axis, which responds to the cytosolic ratio of NAD+ to NADH. The polymorphism rs1467568 occurs in the eighth intron, and studies have found that G allele carriers may be at risk for increased BMI and obesity. Studies have also shown that SIRT1 expression was lower in overweight or obese individuals than it was in lean individuals. Together, these results suggest that G allele carriers may have reduced SIRT1 expression, restricting the SIRT1/PGC-1? axis and mitochondrial biogenesis. Furthermore, studies have found that physical activity and calorie restriction, known activators of SIRT1, can effectively address excess weight in G allele carriers. Lastly, BMI was noticeably higher in G allele carriers with low vitamin E, indicating that antioxidants may support SIRT1 and mitochondrial activity in G allele carriers as well.
TFAM rs1937	The TFAM (transcription factor A, mitochondrial) gene codes a transcription factor that promotes the expression of genes essential for mitochondrial DNA replication and repair. The polymorphism rs1937 results in a threonine substitution for a serine residue at position 12, which occurs in the mitochondrial signaling sequence. Individuals with the GG genotype were found to have reduced endurance exercise capacity and decreased longevity, suggesting that ability of the mitochondria to produce sufficient energy may be decreased. Furthermore, the GG genotype was associated with increased risk for Alzheimer disease.
NEUROTRANSMITTERS	Neurotransmitters are chemicals that are used to produce specific effects in the nervous system. These specific neurotransmitter genomics assess a person's risk for anxiety, depression and dysphoria.
COMT rs4680	The COMT (catechol-O-methyltransferase) gene encodes an enzyme that deactivates catecholamines, including neurotransmitters (adrenaline, noradrenaline and dopamine), by catalyzing the transfer of a methyl groups from S-adenosyl-methionine to a hydroxyl group on a catechol. Therefore, COMT has a crucial role in catecholamine neurotransmission and the metabolism of catechol hormones and xenobiotics. The polymorphism rs4860 results in a methionine substitution for a valine residue at position 108 for soluble COMT, which is prevalent in peripheral tissues, or position 158 for membrane-bound COMT, which is prevalent in the brain. The enzyme produced by the A allele, which encodes a methionine residue, reduces COMT activity due to thermal instability. Moreover, the A allele variant can have a three-to-fourfold reduction in enzyme activity compared to the G allele variant, and the A allele has been associated with a disadvantage processing aversive stimuli, reduced appetite, OCD, and anxiety. Furthermore, COMT metabolizes estrogen, and a study found that girls with the AA genotype had higher levels of free estradiol and earlier pubertal development than girls with the GG genotype, suggesting that the A allele may be associated with less efficient estrogen clearance. However, individuals with the GG genotype may have increased homocysteine levels when combined with a MTHFR variant.
GAD1 rs3828275	The GAD1 (Glutamic Acid Decarboxylase 1) gene encodes the rate-limiting enzyme responsible for conversion of glutamate, a stimulating neurotransmitter, to GABA, a calming neurotransmitter. A deficiency of GABA is associated with a variety of neuropsychological disorders, including anxiety, depression, and sleep disorders. The polymorphism, rs3828275, occurs in the third intron. Carriers of the minor allele have an increased risk for post-traumatic epilepsy, whereas carriers of the wild-type allele are potentially more responsive to treatment with SSRIs.
GAD1 rs769407	The GAD1 (Glutamic Acid Decarboxylase 1) gene encodes the rate-limiting enzyme responsible for conversion of glutamate, a stimulating neurotransmitter, to GABA, a calming neurotransmitter. A deficiency of GABA is associated with a variety of neuropsychological disorders, including anxiety, depression, and sleep disorders. The polymorphism, rs769407, occurs in the sixth intron and has a possible association with an increased risk of neuroticism and mood disorders. Additionally, it has been shown to associate with sleep disturbances in depressed patients.
MAOA	The MAOA (monoamine oxidase A) gene encodes for a riboflavin-dependent enzyme that degrades monoamine neurotransmitters, such as serotonin, dopamine, and norepinephrine. Thus, MAOA ends neuronal signaling induced by those neurotransmitters. MAOA is bound to the mitochondrial membrane by a transmembrane segment, and it has overlapping function with MAOB. However, MAOA has a higher affinity for serotonin and norepinephrine than MAOB. The G allele for the polymorphism rs6323 results in an enzyme with increased activity, and the G allele may be associated with attention deficit hyperactivity disorder (ADHD). Furthermore, women with the GG genotype had a sustained reaction to stressful stimuli, suggesting reduced stress resiliency.
MAOB	The MAOB (monoamine oxidase B) gene encodes for a riboflavin-dependent enzyme that degrades monoamine neurotransmitters, such as serotonin, dopamine, and norepinephrine. Thus, MAOB ends neuronal signaling induced by those neurotransmitters. MAOB is bound to the mitochondrial membrane by a transmembrane segment, and it has overlapping function with MAOA. However, MAOB has a higher affinity for dopamine, phenylethylamine, and benzylamine than MAOA. The polymorphism rs1799836 occurs in intron 13, and the T allele results in an enzyme with increased activity leading to a higher rate of dopamine turnover. As a result, the T allele has been associated with motor complications in Parkinson's disease, and the TT genotype may be a risk factor for Alzheimer's disease, which is known to present with decreased dopamine signaling. Lastly, survey data suggests that the T allele might be associated with feelings of stress, loneliness, sporadic attention, and anxiety.

Disclaimers

TESTING:

Testing Performed By: AC

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Testing for genetic variation/mutation on listed genes was performed using ProFlex PCR and Real-Time PCR with TaqMan® allele-specific probes on the QuantStudio 12K Flex. All genetic testing is performed by GX Sciences, LLC d/b/a Fagron Genomics US ("Fagron Genomics US") (807 Las Cimas Pkwy, Suite 145, Austin, TX. 78746). This test will not detect all the known alleles that result in altered or inactive tested genes. This test does not account for all individual variations in the individual tested. Test results do not rule out the possibility that this individual could be a carrier of other mutations/variations not detected by this gene mutation/variation panel. Rare mutations surrounding these alleles may also affect our detection of genetic variations. Thus, the interpretation is given as a probability. Therefore, this genetic information shall be interpreted in conjunction with other clinical findings and familial history for the administration of specific nutrients. Patients should receive appropriate genetic counseling to explain the implications of these test results. Details of assay performance and algorithms leading to clinical recommendations are available upon request. The analytical and performance characteristics of this laboratory developed test (LDT) were determined by Fagron Genomics US's laboratory (Laboratory Director: James Jacobson, PhD) pursuant to Clinical Laboratory Improvement Amendments (CLIA) requirements (CLIA #: 45D2144988).

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Fagron Genomics US SNP References

DETOXIFICATION SNP References

CTH

• Wenstrom, K. D., Johanning, G. L., Owen, J., Johnston, K. E., Acton, S., & Tamura, T. (2000). Role of amniotic fluid homocysteine level and of fetal 5, 10-methylenetetrahydrofolate reductase genotype in the etiology of neural tube defects. *American Journal of Medical Genetics*, 90(1), 12–16. [https://doi.org/10.1002/\(sici\)1096-8628\(20000103\)90:1<12::aid-ajmg3>3.0.co;2-h](https://doi.org/10.1002/(sici)1096-8628(20000103)90:1<12::aid-ajmg3>3.0.co;2-h)

GCLC rs17883901

• Koide, S., Kugiyama, K., Sugiyama, S., Nakamura, S., Fukushima, H., Honda, O., Yoshimura, M., & Ogawa, H. (2003). Association of polymorphism in glutamate-cysteine ligase catalytic subunit gene with coronary vasomotor dysfunction and myocardial infarction. *Journal of the American College of Cardiology*, 41(4), 539–545. [https://doi.org/10.1016/s0735-1097\(02\)02866-8](https://doi.org/10.1016/s0735-1097(02)02866-8)

• Zuo, H., Xu, W., Luo, M., Zhu, Z., & Zhu, G. (2007). [The glutamate-cysteine ligase catalytic subunit gene C-129T and modifier subunit gene G-23T polymorphisms and risk for coronary diseases]. *Zhonghua Xin Xue Guan Bing Za Zhi*, 35(7), 637–640.

• Azarova, I., Klysova, E., Lazarenko, V., Konoplya, A., & Polonikov, A. (2020). Genetic variants in glutamate cysteine ligase confer protection against type 2 diabetes. *Molecular Biology Reports*, 47(8), 5793–5805. <https://doi.org/10.1007/s11033-020-05647-5>

GSR rs8190955

• Krishnamurthy, H. K., Rajavelu, I., Pereira, M., Jayaraman, V., Krishna, K., Wang, T., Bei, K., & Rajasekaran, J. J. (2024). Inside the genome: Understanding genetic influences on oxidative stress. *Frontiers in Genetics*, 15, 1397352. <https://doi.org/10.3389/fgene.2024.1397352>

• Vyas, B., Bhowmik, R., Akhter, M., & Ahmad, F. J. (2022). Identification, analysis of deleterious SNPs of the human GSR gene and their effects on the structure and functions of associated proteins and other diseases. *Scientific Reports*, 12(1), 5474. <https://doi.org/10.1038/s41598-022-09295-6>

GSTP1

• Simeunovic, D., Odanovic, N., Pljesa-Erecgovac, M., Radic, T., Radovanovic, S., Coric, V., Milinkovic, I., Matic, M., Djukic, T., Ristic, A., Risimic, D., Seferovic, P., Simic, T., Simic, D., & Savic-Radojevic, A. (2019). Glutathione Transferase P1 Polymorphism Might Be a Risk Determinant in Heart Failure. *Disease Markers*, 2019, 6984845. <https://doi.org/10.1155/2019/6984845>

• Scarfó, M., Sciandra, C., Ruberto, S., & Santovito, A. (2021). GSTT1, GSTP1 and XPC genes are associated with longevity in an Italian cohort. *Annals of Human Biology*, 48(5), 443–447. <https://doi.org/10.1080/03014460.2021.1985170>

• Palmer, C. N. A., Doney, A. S. F., Lee, S. P., Murrie, I., Ismail, T., Macgregor, D. F., & Mukhopadhyay, S. (2006). Glutathione S-transferase M1 and P1 genotype, passive smoking, and peak expiratory flow in asthma. *Pediatrics*, 118(2), 710–716. <https://doi.org/10.1542/peds.2005-3030>

• Mukhammadjiyeva, G. F., Bakirov, A. B., Karimov, D. O., Ziatdinova, M. M., Valova, Y. V., Borisova, A. I., & Distanova, A. A. (2022). Analysis of the GSTP1 rs1695 polymorphism association with the development of asthma and phenotypic manifestations. *The Journal of Asthma: Official Journal of the Association for the Care of Asthma*, 59(6), 1065–1069. <https://doi.org/10.1080/02770903.2021.1910295>

• Moyer, A. M., Salavaggione, O. E., Wu, T.-Y., Moon, I., Eckloff, B. W., Hildebrandt, M. A. T., Schaid, D. J., Wieben, E. D., & Weinsilboum, R. M. (2008). Glutathione S-Transferase P1: Gene Sequence Variation and Functional Genomic Studies. *Cancer Research*, 68(12), 4791–4801. <https://doi.org/10.1158/0008-5472.CAN-07-6724>

• Melén, E., Nyberg, F., Lindgren, C. M., Berglind, N., Zucchelli, M., Nordling, E., Hallberg, J., Svartengren, M., Morgenstern, R., Kere, J., Bellander, T., Wickman, M., & Pershagen, G. (2008). Interactions between glutathione S-transferase P1, tumor necrosis factor, and traffic-related air pollution for development of childhood allergic disease. *Environmental Health Perspectives*, 116(8), 1077–1084. <https://doi.org/10.1289/ehp.11117>

• Katsarou, M.-S., Giakoumaki, M., Papadimitriou, A., Demertzis, N., Androusoopoulos, V., & Drakoulis, N. (2018). Genetically driven antioxidant capacity in a Caucasian Southeastern European population. *Mechanisms of Ageing and Development*, 172, 1–5. <https://doi.org/10.1016/j.mad.2017.08.010>

• Dai, X., Bui, D. S., & Lodge, C. (2021). Glutathione S-Transferase Gene Associations and Gene-Environment Interactions for Asthma. *Current Allergy and Asthma Reports*, 21(5), 31. <https://doi.org/10.1007/s11882-021-01005-y>

• Hollman, A. L., Tchounwou, P. B., & Huang, H.-C. (2016). The Association between Gene-Environment Interactions and Diseases Involving the Human GST Superfamily with SNP Variants. *International Journal of Environmental Research and Public Health*, 13(4), 379. <https://doi.org/10.3390/ijerph13040379>

• do Nascimento, M. R., Silva de Souza, R. O., Silva, A. L., Lima, E. S., Gonçalves, M. S., & de Moura Neto, J. P. (2021). GSTP1 rs1695 and rs1871042, and SOD2 rs4880 as molecular markers of lipid peroxidation in blood storage. *Blood Transfusion = Trasfusione Del Sangue*, 19(4), 309–316. <https://doi.org/10.2450/2020.0062-20>

• Tamer, L., Calikoğlu, M., Ates, N. A., Yildirim, H., Ercan, B., Saritas, E., Unlü, A., & Atik, U. (2004). Glutathione-S-transferase gene polymorphisms (GSTT1, GSTM1, GSTP1) as increased risk factors for asthma. *Respirology (Carlton, Vic)*, 9(4), 493–498. <https://doi.org/10.1111/j.1440-1843.2004.00657.x>

NAT2 rs1801280

• Patin, E., Barreiro, L. B., Sabeti, P. C., Austerlitz, F., Luca, F., Sajantila, A., Behar, D. M., Semino, O., Sakuntabhai, A., Guiso, N., Gicquel, B., McElreavey, K., Harding, R. M., Heyer, E., & Quintana-Murci, L. (2006). Deciphering the ancient and complex evolutionary history of human arylamine N-acetyltransferase genes. *American Journal of Human Genetics*, 78(3), 423–436. <https://doi.org/10.1086/500614>

• Martínez-González, L. J., Antúnez-Rodríguez, A., Vázquez-Alonso, F., Hernández, A. F., & Álvarez-Cubero, M. J. (2020). Genetic variants in xenobiotic detoxification enzymes, antioxidant defenses and hormonal pathways as biomarkers of susceptibility to prostate cancer. *The Science of the Total Environment*, 730, 138314. <https://doi.org/10.1016/j.scitotenv.2020.138314>

• Magalon, H., Patin, E., Austerlitz, F., Hegay, T., Aldashev, A., Quintana-Murci, L., & Heyer, E. (2008). Population genetic diversity of the NAT2 gene supports a role of acetylation in human adaptation to farming in Central Asia. *European Journal of Human Genetics: EJHG*, 16(2), 243–251. <https://doi.org/10.1038/sj.ejhg.5201963>

• Moore, L. E., Baris, D. R., Figueroa, J. D., García-Closas, M., Karagas, M. R., Schwenn, M. R., Johnson, A. T., Lubin, J. H., Hein, D. W., Dagnall, C. L., Colt, J. S., Kida, M., Jones, M. A., Schned, A. R., Cherala, S. S., Chanock, S. J., Cantor, K. P., Silverman, D. T., & Rothman, N. (2011). GSTM1 null and NAT2 slow acetylation genotypes, smoking intensity and bladder cancer risk: Results from the New England bladder cancer study and NAT2 meta-analysis. *Carcinogenesis*, 32(2), 182–189. <https://doi.org/10.1093/carcin/bgg223>

• Hein, D. W. (2002). Molecular genetics and function of NAT1 and NAT2: Role in aromatic amine metabolism and carcinogenesis. *Mutation Research*, 506–507, 65–77. [https://doi.org/10.1016/s0027-5107\(02\)00153-7](https://doi.org/10.1016/s0027-5107(02)00153-7)

• García-Closas, M., Malats, N., Silverman, D., Dosemeci, M., Kogevinas, M., Hein, D. W., Tardón, A., Serra, C., Carrato, A., García-Closas, R., Lloreta, J., Castaño-Vinyals, G., Yeager, M., Welch, R., Chanock, S., Chatterjee, N., Wacholder, S., Samanic, C., Torá, M., ... Rothman, N. (2005). NAT2 slow acetylation, GSTM1 null genotype, and risk of bladder cancer: Results from the Spanish Bladder Cancer Study and meta-analyses. *Lancet (London, England)*, 366(9486), 649–659. [https://doi.org/10.1016/S0140-6736\(05\)67137-1](https://doi.org/10.1016/S0140-6736(05)67137-1)

NFE2L2 rs6721961

• Marzec, J. M., Christie, J. D., Reddy, S. P., Jedlicka, A. E., Vuong, H., Lanken, P. N., Aplenc, R., Yamamoto, T., Yamamoto, M., Cho, H.-Y., & Kleeberger, S. R. (2007). Functional polymorphisms in the transcription factor NRF2 in humans increase the risk of acute lung injury. *FASEB Journal: Official Publication of the Federation of American Societies for Experimental Biology*, 21(9), 2237–2246. <https://doi.org/10.1096/fj.06-7759com>

• Reuland, D. J., McCord, J. M., & Hamilton, K. L. (2013). The role of Nrf2 in the attenuation of cardiovascular disease. *Exercise and Sport Sciences Reviews*, 41(3), 162–168. <https://doi.org/10.1097/JES.0b013e3182948a1e>

• Shimizu, S., Mimura, J., Hasegawa, T., Shimizu, E., Imoto, S., Tsushima, M., Kasai, S., Yamazaki, H., Ushida, Y., Suganuma, H., Tomita, H., Yamamoto, M., Nakaji, S., & Itoh, K. (2020). Association of single nucleotide polymorphisms in the NRF2 promoter with vascular stiffness with aging. *PLoS One*, 15(8), e0236834. <https://doi.org/10.1371/journal.pone.0236834>

• Wang, X., Chen, H., Liu, J., Ouyang, Y., Wang, D., Bao, W., & Liu, L. (2015). Association between the NF-E2 Related Factor 2 Gene Polymorphism and Oxidative Stress, Anti-Oxidative Status, and Newly-Diagnosed Type 2 Diabetes Mellitus in a Chinese Population. *International Journal of Molecular Sciences*, 16(7), 16483–16496. <https://doi.org/10.3390/ijms160716483>

• Satta, S., Mahmoud, A. M., Wilkinson, F. L., Yvonne Alexander, M., & White, S. J. (2017). The Role of Nrf2 in Cardiovascular Function and Disease. *Oxidative Medicine and Cellular Longevity*, 2017, 9237263. <https://doi.org/10.1155/2017/9237263>

SOD2

• Paludo, F. J. O., Bristol, I. J., Alho, C. S., Gelain, D. P., & Moreira, J. C. F. (2014). Effects of 47C allele (rs4880) of the SOD2 gene in the production of intracellular reactive species in peripheral blood mononuclear cells with and without lipopolysaccharides induction. *Free Radical Research*, 48(2), 190–199. <https://doi.org/10.3109/10715762.2013.859385>

• Rosenblum, J. S., Gilula, N. B., & Lerner, R. A. (1996). On signal sequence polymorphisms and diseases of distribution. *Proceedings of the National Academy of Sciences of the United States of America*, 93(9), 4471–4473.

• Sutton, A., Khoury, H., Prip-Buus, C., Cepanec, C., Pessayre, D., & Degoul, F. (2003). The Ala16Val genetic dimorphism modulates the import of human manganese superoxide dismutase into rat liver mitochondria. *Pharmacogenetics*, 13(3), 145–157. <https://doi.org/10.1097/01.fpc.0000054067.64000.8f>

• Hernández-Guerrero, C., Parra-Carriedo, A., Ruiz-de-Santiago, D., Galicia-Castillo, O., Buenostro-Jáuregui, M., & Díaz-Gutiérrez, C. (2018). Genetic polymorphisms of antioxidant enzymes CAT and SOD affect the outcome of clinical, biochemical, and anthropometric variables in people with obesity under a dietary intervention. *Genes & Nutrition*, 13, 1. <https://doi.org/10.1186/s12263-017-0590-2>

• Fj, P., A. S.-P., Cs, A., Dp, G., & Jc, M. (2013). Participation of 47C>T SNP (Ala-9Val polymorphism) of the SOD2 gene in the intracellular environment of human peripheral blood mononuclear cells with and without lipopolysaccharides. *Molecular and Cellular Biochemistry*, 372(1–2). <https://doi.org/10.1007/s11010-012-1453-1>

• Broz, M., Furlan, V., Lešnik, S., Jukič, M., & Bren, U. (2022). The Effect of the Ala16Val Mutation on the Secondary Structure of the Manganese Superoxide Dismutase Mitochondrial Targeting Sequence. *Antioxidants*, 11(12), 2348. <https://doi.org/10.3390/antiox1122348>

• Bresciani, G., González-Gallego, J., da Cruz, I. B., de Paz, J. A., & Cuevas, M. J. (2013). The Ala16Val MnSOD gene polymorphism modulates oxidative response to exercise. *Clinical Biochemistry*, 46(4–5), 335–340. <https://doi.org/10.1016/j.clinbiochem.2012.11.020>

• Becer, E., & Ç?rakoğlu, A. (2015). Association of the Ala16Val MnSOD gene polymorphism with plasma leptin levels and oxidative stress biomarkers in

obese patients. *Gene*, 568(1), 35–39. <https://doi.org/10.1016/j.gene.2015.05.009> • Jones, D. A., Prior, S. L., Tang, T. S., Bain, S. C., Hurel, S. J., Humphries, S. E., & Stephens, J. W. (2010). Association between the rs4880 superoxide dismutase 2 (C>T) gene variant and coronary heart disease in diabetes mellitus. *Diabetes Research and Clinical Practice*, 90(2), 196–201. <https://doi.org/10.1016/j.diabres.2010.07.009>

DEVELOPMENTAL SNP References

ATG12

• Anton, R. F. et al. Pharmacogenomics. *Nat. Genet.* 16, 268–278 (2008). • Antunes, F. et al. Autophagy and intermittent fasting: the connection for cancer therapy? *Clinics* (Sao Paulo, Brazil) (2018). doi:10.6061/clinics/2018/e814s • Lindberg, S. Autophagy: Definition, Diet, Fasting, Cancer, Benefits, and More. *Healthline* (2014). Available at: <https://www.healthline.com/health/autophagy#bottom-line>. • Takagi, A., Kume, S., Maegawa, H. & Uzu, T. Emerging role of mammalian autophagy in ketogenesis to overcome starvation. *Autophagy* (2016). doi:10.1080/15548627.2016.1151597 • Smith, G. S., Walter, G. L. & Walker, R. M. Clinical Pathology in Non-Clinical Toxicology Testing. in Haschek and Rousseaux's Handbook of Toxicologic Pathology (2013). doi:10.1016/B978-0-12-415759-0.00018-2 • Levine, B. & Kroemer, G. Autophagy in the Pathogenesis of Disease. *Cell* (2008). doi:10.1016/j.cell.2007.12.018 • Mizushima, N. Autophagy: Process and function. *Genes and Development* (2007). doi:10.1101/gad.1599207 • Yuan, J. et al. Polymorphisms in autophagy related genes and the coal workers' pneumoconiosis in a Chinese population. *Gene* 632, 36–42 (2017).

ESSENTIAL MINERALS SNP References

TRPM6 rs7087728

• Kieboom, B. C. T., Ligthart, S., Dehghan, A., Kustjens, S., de Baaij, J. H. F., Franco, O. H., Hofman, A., Zietse, R., Stricker, B. H., & Hoom, E. J. (2017). Serum magnesium and the risk of prediabetes: A population-based cohort study. *Diabetologia*, 60(5), 843–853. <https://doi.org/10.1007/s00125-017-4224-4> • Hess, M. W., de Baaij, J. H. F., Broekman, M. M. T. J., Bisseling, T. M., Haahrhuis, B. J. T., Tan, A. C. I. T. L., Te Morsche, R. H. M., Hoenderop, J. G. J., Bindels, R. J. M., & Drenth, J. P. H. (2017). Common single nucleotide polymorphisms in transient receptor potential melastatin type 6 increase the risk for proton pump inhibitor-induced hypomagnesemia: A case-control study. *Pharmacogenetics and Genomics*, 27(3), 83–88. <https://doi.org/10.1097/FPC.0000000000000259> • Huang, S., Ge, Y., Li, Y., Cui, N., Tan, L., Guo, S., Hao, L., Lei, G., & Yang, X. (2023). Magnesium Status, Genetic Variants of Magnesium-Related Ion Channel Transient Receptor Potential Membrane Melastatin 6 (TRPM6) and the Risk of Gestational Diabetes Mellitus in Chinese Pregnant Women: A Nested Case-Control Study. *Molecular Nutrition & Food Research*, 67(22), e2200835. <https://doi.org/10.1002/mnfr.202200835> • Song, Y., Hsu, Y.-H., Niu, T., Manson, J. E., Buring, J. E., & Liu, S. (2009). Common genetic variants of the ion channel transient receptor potential membrane melastatin 6 and 7 (TRPM6 and TRPM7), magnesium intake, and risk of type 2 diabetes in women. *BMC Medical Genetics*, 10, 4. <https://doi.org/10.1186/1471-2350-10-4> • Wu, J., Xun, P., Tang, Q., Cai, W., & He, K. (2017). Circulating magnesium levels and incidence of coronary heart diseases, hypertension, and type 2 diabetes mellitus: A meta-analysis of prospective cohort studies. *Nutrition Journal*, 16(1), 60. <https://doi.org/10.1186/s12937-017-0280-3>

ESSENTIAL VITAMINS SNP References

SLC23A1

• Kobylecki, C. J., Afzal, S., Davey Smith, G., & Nordestgaard, B. G. (2015). Genetically high plasma vitamin C, intake of fruit and vegetables, and risk of ischemic heart disease and all-cause mortality: A Mendelian randomization study. *The American Journal of Clinical Nutrition*, 101(6), 1135–1143. <https://doi.org/10.3945/ajcn.114.104497> • Yin, X., Chan, L. S., Bose, D., Jackson, A. U., VandeHaar, P., Locke, A. E., Fuchsberger, C., Stringham, H. M., Welch, R., Yu, K., Fernandes Silva, L., Service, S. K., Zhang, D., Hector, E. C., Young, E., Ganel, L., Das, I., Abel, H., Erdos, M. R., ... Boehnke, M. (2022). Genome-wide association studies of metabolites in Finnish men identify disease-relevant loci. *Nature Communications*, 13(1), 1644. <https://doi.org/10.1038/s41467-022-29143-5> • Timpson, N. J., Forouhi, N. G., Brion, M.-J., Harbord, R. M., Cook, D. G., Johnson, P., McConachie, A., Morris, R. W., Rodriguez, S., Luan, J., Ebrahim, S., Padmanabhan, S., Watt, G., Bruckdorfer, K. R., Wareham, N. J., Whincup, P. H., Chanock, S., Sattar, N., Lawlor, D. A., & Davey Smith, G. (2010). Genetic variation at the SLC23A1 locus is associated with circulating concentrations of L-ascorbic acid (vitamin C): Evidence from 5 independent studies with >15,000 participants. *The American Journal of Clinical Nutrition*, 92(2), 375–382. <https://doi.org/10.3945/ajcn.2010.29438> • Ravindran, R. D., Sundaresan, P., Krishnan, T., Vashist, P., Marathi, G., Saravanan, V., Chakravarthy, U., Smeeth, L., Nitsch, D., Young, I. S., & Fletcher, A. E. (2019). Genetic variants in a sodium-dependent vitamin C transporter gene and age-related cataract. *The British Journal of Ophthalmology*, 103(9), 1223–1227. <https://doi.org/10.1136/bjophthalmol-2018-312257>

TPPA rs6994076

• Wright, M. E., Peters, U., Gunter, M. J., Moore, S. C., Lawson, K. A., Yeager, M., Weinstein, S. J., Snyder, K., Virtamo, J., & Albanes, D. (2009). Association of variants in two vitamin e transport genes with circulating vitamin e concentrations and prostate cancer risk. *Cancer Research*, 69(4), 1429–1438. <https://doi.org/10.1158/0008-5472.CAN-08-2343> • Schmölz, L., Birringer, M., Lorkowski, S., & Wallert, M. (2016). Complexity of vitamin E metabolism. *World Journal of Biological Chemistry*, 7(1), 14–43. <https://doi.org/10.4331/wjbc.v7.i1.14> • Ulatowski, L., Dreussi, C., Noy, N., Barnholtz-Sloan, J., Klein, E., & Manor, D. (2012). Expression of the γ -tocopherol transfer protein gene is regulated by oxidative stress and common single-nucleotide polymorphisms. *Free Radical Biology & Medicine*, 53(12), 2318–2326. <https://doi.org/10.1016/j.freeradbiomed.2012.10.528>

VDR rs2228570

• Ebrahimof, S., Angoorani, P., Shab-Bidar, S., Abedini, S., Jahangir, F., & Hedayati, M. (2022). The interactive effect of vitamin D3 supplementation and vitamin D receptor polymorphisms on weight and body composition in overweight women with hypovitaminosis D: A randomized, double-blind, placebo-controlled clinical trial. *Canadian Journal of Physiology and Pharmacology*. <https://doi.org/10.1139/cjpp-2022-0192> • Whitfield, G. K., Remus, L. S., Jurutka, P. W., Zitzer, H., Oza, A. K., Dang, H. T., Hausler, C. A., Galligan, M. A., Thatcher, M. L., Encinas Dominguez, C., & Hausler, M. R. (2001). Functionally relevant polymorphisms in the human nuclear vitamin D receptor gene. *Molecular and Cellular Endocrinology*, 177(1–2), 145–159. [https://doi.org/10.1016/s0303-7207\(01\)00406-3](https://doi.org/10.1016/s0303-7207(01)00406-3) • Uitterlinden, A. G., Fang, Y., Van Meurs, J. B. J., Pols, H. A. P., & Van Leeuwen, J. P. T. M. (2004). Genetics and biology of vitamin D receptor polymorphisms. *Gene*, 338(2), 143–156. <https://doi.org/10.1016/j.gene.2004.05.014> • T, S., P, B., & S, S. (2023). A meta-analysis suggests the association of reduced serum level of vitamin D and T-allele of Fok1 (rs2228570) polymorphism in the vitamin D receptor gene with celiac disease. *Frontiers in Nutrition*, 9. <https://doi.org/10.3389/fnut.2022.996450> • Neyestani, T. R., Djazayeri, A., Shab-Bidar, S., Eshraghian, M. R., Kalayi, A., Shariatzadeh, N., Khalaji, N., Zahedirad, M., Gharavi, A., Houshiarad, A., Chamari, M., & Asadzadeh, S. (2013). Vitamin D Receptor Fok-I polymorphism modulates diabetic host response to vitamin D intake: Need for a nutrigenetic approach. *Diabetes Care*, 36(3), 550–556. <https://doi.org/10.2337/dc12-0919> • Li, L., Wu, B., Liu, J.-Y., & Yang, L.-B. (2013). Vitamin D receptor gene polymorphisms and type 2 diabetes: A meta-analysis. *Archives of Medical Research*, 44(3), 235–241. <https://doi.org/10.1016/j.arcmed.2013.02.002> • Arai, H., Miyamoto, K., Taketani, Y., Yamamoto, H., Iemori, Y., Morita, K., Tonai, T., Nishisho, T., Mori, S., & Takeda, E. (1997). A vitamin D receptor gene polymorphism in the translation initiation codon: Effect on protein activity and relation to bone mineral density in Japanese women. *Journal of Bone and Mineral Research: The Official Journal of the American Society for Bone and Mineral Research*, 12(6), 915–921. <https://doi.org/10.1359/jbmr.1997.12.6.915> • Abrams, S. A., Griffin, I. J., Hawthorne, K. M., Chen, Z., Gunn, S. K., Wilde, M., Darlington, G., Shypailo, R. J., & Ellis, K. J. (2005). Vitamin D receptor Fok1 polymorphisms affect calcium absorption, kinetics, and bone mineralization rates during puberty. *Journal of Bone and Mineral Research: The Official Journal of the American Society for Bone and Mineral Research*, 20(6), 945–953. <https://doi.org/10.1359/JBMR.050114> • Ames, S. K., Ellis, K. J., Gunn, S. K., Copeland, K. C., & Abrams, S. A. (1999). Vitamin D receptor gene Fok1 polymorphism predicts calcium absorption and bone mineral density in children. *Journal of Bone and Mineral Research: The Official Journal of the American Society for Bone and Mineral Research*, 14(5), 740–746. <https://doi.org/10.1359/jbmr.1999.14.5.740>

General SNP References

CYP1A2

• Tornio, A., & Backman, J. T. (2018). Cytochrome P450 in Pharmacogenetics: An Update. *Advances in Pharmacology* (San Diego, Calif.), 83, 3–32. <https://doi.org/10.1016/bs.apha.2018.04.007> • Djordjevic, N., Ghotbi, R., Jankovic, S., & Akillu, E. (2010). Induction of CYP1A2 by heavy coffee consumption is associated with the CYP1A2 -163C>A polymorphism. *European Journal of Clinical Pharmacology*, 66(7), 697–703. <https://doi.org/10.1007/s00228-010-0823-4> • Djordjevic, N., Ghotbi, R., Bertilsson, L., Jankovic, S., & Akillu, E. (2008). Induction of CYP1A2 by heavy coffee consumption in Serbs and Swedes. *European Journal of Clinical Pharmacology*, 64(4), 381–385. <https://doi.org/10.1007/s00228-007-0436-6> • Sachse, C., Brockmöller, J., Bauer, S., & Roots, I. (1999). Functional significance of a C>A polymorphism in intron 1 of the cytochrome P450 CYP1A2 gene tested with caffeine. *British Journal of Clinical Pharmacology*, 47(4), 445–449. <https://doi.org/10.1046/j.1365-2125.1999.00898.x>

HEALTH PRECAUTIONS SNP References

ACE

• Chung, C.-M., Wang, R.-Y., Chen, J.-W., Fann, C. S. J., Leu, H.-B., Ho, H.-Y., Ting, C.-T., Lin, T.-H., Sheu, S.-H., Tsai, W.-C., Chen, J.-H., Jong, Y.-S., Lin, S.-J., Chen, Y.-T., & Pan, W.-H. (2010). A genome-wide association study identifies new loci for ACE activity: Potential implications for response to ACE inhibitor. *The Pharmacogenomics Journal*, 10(6), 537–544. <https://doi.org/10.1038/tj.2009.70> • Rigat, B., Hubert, C., Alhenc-Gelas, F., Cambien, F., Corvol, P., & Soubrier, F. (1990). An insertion/deletion polymorphism in the angiotensin I-converting enzyme gene accounting for half the variance of serum enzyme levels. *The Journal of Clinical Investigation*, 86(4), 1343–1346. <https://doi.org/10.1172/JCI114844> • Dengel, D. R., Brown, M. D., Ferrell, R. E., & Supiano, M. A. (2001). Role of angiotensin converting enzyme genotype in sodium sensitivity in older hypertensives. *American Journal of Hypertension*, 14(12), 1178–1184. [https://doi.org/10.1016/s0895-7061\(01\)02204-x](https://doi.org/10.1016/s0895-7061(01)02204-x)

• Abdollahi, M. R., Huang, S., Rodriguez, S., Guthrie, P. A. I., Smith, G. D., Ebrahimi, S., Lawlor, D. A., Day, I. N. M., & Gaunt, T. R. (2008). Homogeneous assay of rs4343, an ACE I/D proxy, and an analysis in the British Women's Heart and Health Study (BWHHS). *Disease Markers*, 24(1), 11–17. <https://doi.org/10.1155/2008/813679>

F10

• Thareja, G., Belkadi, A., Arnold, M., Albagha, O. M. E., Graumann, J., Schmidt, F., Grallert, H., Peters, A., Gieger, C., Consortium, T. Q. G. P. R., & Suhre, K. (2023). Differences and commonalities in the genetic architecture of protein quantitative trait loci in European and Arab populations. *Human Molecular Genetics*, 32(6), 907–916. <https://doi.org/10.1093/hmg/ddac243> • Kanai, M., Akiyama, M., Takahashi, A., Matoba, N., Momozawa, Y., Ikeda, M., Iwata, N., Ikegawa, S., Hirata, M., Matsuda, K., Kubo, M., Okada, Y., & Kamatani, Y. (2018). Genetic analysis of quantitative traits in the Japanese population links cell types to complex human diseases. *Nature Genetics*, 50(3), 390–400. <https://doi.org/10.1038/s41588-018-0047-6> • Ken-Dror, G., Drenos, F., Humphries, S. E., Talmud, P. J., Hingorani, A. D., Kivimäki, M., Kumari, M., Bauer, K. A., Morrissey, J. H., & Ireland, H. A. (2010). Haplotype and genotype effects of the *F7* gene on circulating factor VII, coagulation activation markers and incident coronary heart disease in UK men. *Journal of Thrombosis and Haemostasis*: JTH, 8(11), 2394–2403. <https://doi.org/10.1111/j.1538-7836.2010.04035.x>

F5

• Klarin, D., Emdin, C. A., Natarajan, P., Conrad, M. F., INVENT Consortium, & Kathiresan, S. (2017). Genetic Analysis of Venous Thromboembolism in UK Biobank Identifies the ZFPM2 Locus and Implicates Obesity as a Causal Risk Factor. *Circulation. Cardiovascular Genetics*, 10(2), e001643. <https://doi.org/10.1161/CIRCGENETICS.116.001643> • Juul, K., Tybjaerg-Hansen, A., Schnohr, P., & Nordestgaard, B. G. (2004). Factor V Leiden and the risk for venous thromboembolism in the adult Danish population. *Annals of Internal Medicine*, 140(5), 330–337. <https://doi.org/10.7326/0003-4819-140-5-200403020-00008> • Simone, B., De Stefano, V., Leoncini, E., Zacho, J., Martinelli, I., Emmerich, J., Rossi, E., Folsom, A. R., Almawi, W. Y., Scarabin, P. Y., den Heijer, M., Cushman, M., Penco, S., Vaya, A., Angchaisuksiri, P., Okumus, G., Gemmati, D., Cima, S., Akar, N., ... Boccia, S. (2013). Risk of venous thromboembolism associated with single and combined effects of Factor V Leiden, Prothrombin 20210A and Methylenelethraydrofolate reductase C677T: A meta-analysis involving over 11,000 cases and 21,000 controls. *European Journal of Epidemiology*, 28(8), 621–647. <https://doi.org/10.1007/s10654-013-9825-8>

PDE8B rs4704397

• Taylor, P. N., Porcu, E., Chew, S., Campbell, P. J., Traglia, M., Brown, S. J., Mullin, B. H., Shihab, H. A., Min, J., Walter, K., Memari, Y., Huang, J., Barnes, M. R., Beilby, J. P., Charoen, P., Danecek, P., Dudbridge, F., Forgetta, V., Greenwood, C., ... UKOK Consortium. (2015). Whole-genome sequence-based analysis of thyroid function. *Nature Communications*, 6, 5681. <https://doi.org/10.1038/ncomms6681> • Agretti, P., De Marco, G., Di Cosmo, C., Bagattini, B., Ferrarini, E., Montanelli, L., Vitti, P., & Tonacchera, M. (2014). Frequency and effect on serum TSH of phosphodiesterase 8B (PDE8B) gene polymorphisms in patients with sporadic nonautoimmune subclinical hypothyroidism. *Journal of Endocrinological Investigation*, 37(2), 189–194. <https://doi.org/10.1007/s40618-013-0036-7> • Arnaud-Lopez, L., Usala, G., Ceresini, G., Mitchell, B. D., Pilia, M. G., Piras, M. G., Sestu, N., Maschio, A., Busonero, F., Albai, G., Dei, M., Lai, S., Mulas, A., Crisponi, L., Tanaka, T., Bandinelli, S., Guralnik, J. M., Loi, A., Balaci, L., ... Naitza, S. (2008). Phosphodiesterase 8B Gene Variants Are Associated with Serum TSH Levels and Thyroid Function. *American Journal of Human Genetics*, 82(6), 1270–1280. <https://doi.org/10.1016/j.ajhg.2008.04.019> • Mansuri, T., Jadeja, S. H. D., Singh, M., Begum, R., & Robin, P. (2020). Phosphodiesterase 8B Polymorphism rs4704397 Is Associated with Infertility in Subclinical Hypothyroid Females: A Case-Control Study. *International Journal of Fertility & Sterility*, 14(2), 122–128. <https://doi.org/10.22074/ijfs.2020.6015>

HORMONE METABOLISM SNP References

CYP19A1

• Haiman, C. A., Dossus, L., Setiawan, V. W., Stram, D. O., Dunning, A. M., Thomas, G., Thun, M. J., Albanes, D., Altshuler, D., Ardanaz, E., Boeing, H., Buring, J., Burt, N., Calle, E. E., Chanock, S., Clavel-Chapelon, F., Colditz, G. A., Cox, D. G., Feigelson, H. S., ... Ziegler, R. G. (2007). Genetic variation at the CYP19A1 locus predicts circulating estrogen levels but not breast cancer risk in postmenopausal women. *Cancer Research*, 67(5), 1893–1897. <https://doi.org/10.1158/0008-5472.CAN-06-4123> • Yip, L., Zaloumis, S., Irwin, J., Severi, G., Hopper, J., Giles, G., Harrap, S., Sinclair, R., & Ellis, J. (2009). Gene-wide association study between the aromatase gene (CYP19A1) and female pattern hair loss. *The British Journal of Dermatology*, 161(2), 289–294. <https://doi.org/10.1111/j.1365-2133.2009.09186.x>

INFLAMMATORY SNP References

AOC1

• Maintz, L., & Novak, N. (2007). Histamine and histamine intolerance. *The American Journal of Clinical Nutrition*, 85(5), 1185–1196. <https://doi.org/10.1093/ajcn/85.5.1185> • Maintz, L., Yu, C.-F., Rodríguez, E., Baurecht, H., Bieber, T., Illig, T., Weidinger, S., & Novak, N. (2011). Association of single nucleotide polymorphisms in the diamine oxidase gene with diamine oxidase serum activities. *Allergy*, 66(7), 893–902. <https://doi.org/10.1111/j.1398-9995.2011.02548.x>

ATG16L1

• Wellcome Trust Case Control Consortium. (2007). Genome-wide association study of 14,000 cases of seven common diseases and 3,000 shared controls. *Nature*, 447(7145), 661–678. <https://doi.org/10.1038/nature05911> • Levine, B., & Kroemer, G. (2008). Autophagy in the Pathogenesis of Disease. *Cell*, 132(1), 27–42. <https://doi.org/10.1016/j.cell.2007.12.018>

ATG5

• Shao, Y., Chen, F., Chen, Y., Zhang, W., Lin, Y., Cai, Y., Yin, Z., Tao, S., Liao, Q., Zhao, J., Mai, H., He, Y., He, J., & Cui, L. (2017). Association between genetic polymorphisms in the autophagy-related 5 gene promoter and the risk of sepsis. *Scientific Reports*, 7, 9399. <https://doi.org/10.1038/s41598-017-09978-5> • Grosjean, I., Roméo, B., Domdrom, M.-A., Belaid, A., D'Andréa, G., Guillot, N., Gherardi, R. K., Gal, J., Milano, G., Marquette, C. H., Hung, R. J., Landi, M. T., Han, Y., Brest, P., Von Bergen, M., Klionsky, D. J., Amos, C. I., Hofman, P., & Mograbi, B. (n.d.). Autophagopathies: From autophagy gene polymorphisms to precision medicine for human diseases. *Autophagy*, 18(11), 2519–2536. <https://doi.org/10.1080/15548627.2022.2039994> • Martin, L. J., Gupta, J., Jyothula, S. S. S. K., Butsch Kovacic, M., Biagini Myers, J. M., Patterson, T. L., Erickson, M. B., He, H., Gibson, A. M., Baye, T. M., Amirsetty, S., Tsoras, A. M., Sha, Y., Eissa, N. T., & Hershey, G. K. K. (2012). Functional variant in the autophagy-related 5 gene promoter is associated with childhood asthma. *PLoS One*, 7(4), e33454. <https://doi.org/10.1371/journal.pone.0033454> • Tamargo-Gómez, I., Fernández, Á. F., & Mariño, G. (2020). Pathogenic Single Nucleotide Polymorphisms on Autophagy-Related Genes. *International Journal of Molecular Sciences*, 21(21), 8196. <https://doi.org/10.3390/ijms21218196>

C3 rs2250656

• Phillips, C. M., Goumidi, L., Bertrais, S., Ferguson, J. F., Field, M. R., Kelly, E. D., Peloso, G. M., Cupples, L. A., Shen, J., Ordovas, J. M., McManus, R., Hercberg, S., Portugal, H., Lairon, D., Planells, R., & Roche, H. M. (2009). Complement component 3 polymorphisms interact with polyunsaturated fatty acids to modulate risk of metabolic syndrome. *The American Journal of Clinical Nutrition*, 90(6), 1665–1673. <https://doi.org/10.3945/ajcn.2009.28101> • Asteris, P. G., Gavriilaki, E., Touloumenidou, T., Koravou, E.-E., Koutra, M., Papayanni, P. G., Pouleres, A., Karali, V., Lemonis, M. E., Mamou, A., Skentou, A. D., Papalexandri, A., Varelas, C., Chatzopoulou, F., Chatzidimitriou, M., Chatzidimitriou, D., Veleni, A., Rapti, E., Kioumis, I., ... Anagnostopoulos, A. (2022). Genetic prediction of ICU hospitalization and mortality in COVID-19 patients using artificial neural networks. *Journal of Cellular and Molecular Medicine*, 26(5), 1445–1455. <https://doi.org/10.1111/jcmm.17098>

CD14 rs2569190

• Xu, J.-J., Liu, K.-Q., Ying, Z.-M., Zhu, X.-W., Xu, X.-J., Zhao, P.-P., Bai, W.-Y., Qiu, M.-C., Zhang, X.-W., & Zheng, H.-F. (2019). Effect of CD14 polymorphisms on the risk of cardiovascular disease: Evidence from a meta-analysis. *Lipids in Health and Disease*, 18(1), 74. <https://doi.org/10.1186/s12944-019-1018-3> • Williams, L. K., McPhee, R. A., Ownby, D. R., Peterson, E. L., James, M., Zoratti, E. M., & Johnson, C. C. (2006). Gene-environment interactions with CD14 C-260T and their relationship to total serum IgE levels in adults. *The Journal of Allergy and Clinical Immunology*, 118(4), 851–857. <https://doi.org/10.1016/j.jaci.2006.07.007> • Wang, Z., Hu, J., Fan, R., Zhou, J., & Zhong, J. (2012). Association between CD14 gene C-260T polymorphism and inflammatory bowel disease: A meta-analysis. *PLoS One*, 7(9), e45144. <https://doi.org/10.1371/journal.pone.0045144> • Mertens, J., Bregadze, R., Mansur, A., Askar, E., Bickeböller, H., Ramadori, G., & Milhm, S. (2009). Functional impact of endotoxin receptor CD14 polymorphisms on transcriptional activity. *Journal of Molecular Medicine (Berlin, Germany)*, 87(8), 815–824. <https://doi.org/10.1007/s00109-009-0479-7> • Kamel, M. A., Selim, E. S., Tantawy, E. A., Elgendy, A., Abdulmageed, A., & Anis, R. H. (2023). Association of serum CD14 level and functional polymorphism C-159T in the promoter region of CD14 gene with allergic rhinitis. *Clinical and Experimental Medicine*, 23(8), 4861–4869. <https://doi.org/10.1007/s10238-023-01097-y>

CTLA4

• Chen, Y., Chen, S., Gu, Y., Feng, Y., Shi, Y., Fu, Q., Wang, Z., Cai, Y., Dai, H., Zheng, S., Sun, M., Zhang, M., Xu, X., Chen, H., Xu, K., & Yang, T. (2018). CTLA-4 +49 G/A, a functional T1D risk SNP, affects CTLA-4 level in Treg subsets and IA-2a positivity, but not beta-cell function. *Scientific Reports*, 8(1), 10074. <https://doi.org/10.1038/s41598-018-28423-9>

• Mousavi, M. J., Shayesteh, M. R. H., Jamalzehi, S., Alimohammadi, R., Rahimi, A., Aslani, S., & Rezaei, N. (2021). Association of the genetic polymorphisms in inhibiting and activating molecules of immune system with rheumatoid arthritis: A systematic review and meta-analysis. *Journal of Research in Medical Sciences: The Official Journal of Isfahan University of Medical Sciences*, 26, 22. https://doi.org/10.4103/jrms.JRMS_567_20

• Patel, H., Mansuri, M. S., Singh, M., Begum, R., Shastri, M., & Misra, A. (2016). Association of Cytotoxic T-Lymphocyte Antigen 4 (CTLA4) and Thyroglobulin (TG) Genetic Variants with Autoimmune Hypothyroidism. *PLoS One*, 11(3), e0149441. <https://doi.org/10.1371/journal.pone.0149441>

• Mäurer, M., Loserth, S., Kolb-Mäurer, A., Ponath, A., Wiese, S., Kruse, N., & Rieckmann, P. (2002). A polymorphism in the human cytotoxic T-lymphocyte antigen 4 (CTLA4) gene (exon 1 +49) alters T-cell activation. *Immunogenetics*, 54(1), 1–8. <https://doi.org/10.1007/s00251-002-0429-9>

• Fathima, N., Name, P., & Ishaq, M. (2019). Association and gene-gene interaction analyses for polymorphic variants in CTLA-4 and FOXP3 genes: Role in susceptibility to autoimmune thyroid disease. *Endocrine*, 64(3), 591–604. <https://doi.org/10.1007/s12020-019-01859-3>

• Chen, M., & Li, S. (2019). Associations between cytotoxic T-lymphocyte-associated antigen 4 gene polymorphisms and diabetes mellitus: A meta-analysis of 76 case-control studies. *Bioscience Reports*, 39(5), BSR20190309. <https://doi.org/10.1042/BSR20190309>

FUT2

• Rausch, P., Rehman, A., Künzel, S., Häslér, R., Ott, S. J., Schreiber, S., Rosenstiel, P., Franke, A., & Baines, J. F. (2011). Colonic mucosa-associated microbiota is influenced by an interaction of Crohn disease and FUT2 (Secretor) genotype. *Proceedings of the National Academy of Sciences of the United States of America*, 108(47), 19030–19035. <https://doi.org/10.1073/pnas.1106408108>

• Hu, M., Zhang, X., Li, J., Chen, L., He, X., & Sui, T. (2022). Fucosyltransferase 2: A Genetic Risk Factor for Intestinal Diseases. *Frontiers in Microbiology*, 13, 940196. <https://doi.org/10.3389/fmicb.2022.940196>

• Tong, M., McHardy, I., Ruegger, P., Goudarzi, M., Kashyap, P. C., Haritunians, T., Li, X., Graeber, T. G., Schwager, E., Huttenhower, C., Fornace, A. J., Sonnenburg, J. L., McGovern, D. P. B., Borneman, J., & Braun, J. (2014). Reprograming of gut microbiome energy metabolism by the FUT2 Crohn's disease risk polymorphism. *The ISME Journal*, 8(11), 2193–2206. <https://doi.org/10.1038/ismej.2014.64>

HLA-DQA1

• van Heel, D. A., Franke, L., Hunt, K. A., Gwilliam, R., Zernakova, A., Inouye, M., Wapenaar, M. C., Barnardo, M. C. N. M., Bethel, G., Holmes, G. K. T., Feighery, C., Jewell, D., Kelleher, D., Kumar, P., Travis, S., Walters, J. R. F., Sanders, D. S., Howdle, P., Swift, J., ... Wijmenga, C. (2007). A genome-wide association study for celiac disease identifies risk variants in the region harboring IL2 and IL21. *Nature Genetics*, 39(7), 827–829. <https://doi.org/10.1038/ng2058>

• Salles, M., Lopetuso, L. R., Eftymakis, K., & Neri, M. (2020). Beyond the HLA Genes in Gluten-Related Disorders. *Frontiers in Nutrition*, 7. <https://www.frontiersin.org/articles/10.3389/fnut.2020.575844>

• Monsuur, A. J., de Bakker, P. I. W., Zernakova, A., Pinto, D., Verduijn, W., Romanos, J., Auricchio, R., Lopez, A., van Heel, D. A., Crusius, J. B. A., & Wijmenga, C. (2008). Effective detection of human leukocyte antigen risk alleles in celiac disease using tag single nucleotide polymorphisms. *PLoS One*, 3(5), e2270. <https://doi.org/10.1371/journal.pone.0002270>

• Dubois, P. C. A., Trynka, G., Franke, L., Hunt, K. A., Romanos, J., Curtotti, A., Zernakova, A., Heap, G. A. R., Adány, R., Aromaa, A., Bardella, M. T., van den Berg, L. H., Bockett, N. A., de la Concha, E. G., Dema, B., Fehrmann, R. S. N., Fernández-Arquero, M., Fiatal, S., Grandone, E., ... van Heel, D. A. (2010). Multiple common variants for celiac disease influencing immune gene expression. *Nature Genetics*, 42(4), 295–302. <https://doi.org/10.1038/ng.543>

• Senapati, S., Sood, A., Mishra, V., Sood, N., Sharma, S., Kumar, L., & Thelma, B. K. (2016). Shared and unique common genetic determinants between pediatric and adult celiac disease. *BMC Medical Genomics*, 9(1), 44. <https://doi.org/10.1186/s12920-016-0211-8>

• de Bakker, P. I. W., McVean, G., Sabeti, P. C., Miretti, M. M., Green, T., Marchini, J., Ke, X., Monstuur, A. J., Whittaker, P., Delgado, M., Morrison, J., Richardson, A., Walsh, E. C., Gao, X., Galver, L., Hart, J., Hafler, D. A., Pericak-Vance, M., Todd, J. A., ... Rioux, J. D. (2006). A high-resolution HLA and SNP haplotype map for disease association studies in the extended human MHC. *Nature Genetics*, 38(10), 1166–1172. <https://doi.org/10.1038/ng1885>

HLA-DQB1 rs7454108

• Monsuur, A. J., de Bakker, P. I. W., Zernakova, A., Pinto, D., Verduijn, W., Romanos, J., Auricchio, R., Lopez, A., van Heel, D. A., Crusius, J. B. A., & Wijmenga, C. (2008). Effective detection of human leukocyte antigen risk alleles in celiac disease using tag single nucleotide polymorphisms. *PLoS One*, 3(5), e2270. <https://doi.org/10.1371/journal.pone.0002270>

IL-13

• Cameron, L., Webster, R. B., Stempel, J. M., Kiesler, P., Kabesch, M., Ramachandran, H., Yu, L., Stern, D. A., Graves, P. E., Lohman, I. C., Wright, A. L., Halonen, M., Klimecki, W. T., & Vercelli, D. (2006). Th2 cell-selective enhancement of human IL13 transcription by IL13-1112C>T, a polymorphism associated with allergic inflammation. *Journal of Immunology* (Baltimore, Md. : 1950), 177(12), 8633–8642. <https://doi.org/10.4049/jimmunol.177.12.8633>

• Omranianava, M., Eslami, M. M., Aslani, S., Razi, B., Imani, D., & Feyzinia, S. (2022). Interleukin 13 gene polymorphism and susceptibility to asthma: A meta-regression and meta-analysis. *European Annals of Allergy and Clinical Immunology*, 54(4), 150–167. <https://doi.org/10.23822/EurAnnACI.1764-1489.180>

• Liao, N., Zhao, H., Chen, M.-L., & Xie, Z.-F. (2017). Association of the IL-13 polymorphisms rs1800925 and rs20541 with chronic obstructive pulmonary disease risk: An updated meta-analysis. *Medicine*, 96(47), e8556. <https://doi.org/10.1097/MD.00000000000008566>

• Dimberg, J., Rubér, M., Skarstedt, M., Andersson, M., & Andersson, R. E. (2020). Genetic polymorphism patterns suggest a genetic driven inflammatory response as pathogenesis in appendicitis. *International Journal of Colorectal Disease*, 35(2), 277–284. <https://doi.org/10.1007/s00384-019-03473-1>

IL5

• Ishigaki, K., Akiyama, M., Kanai, M., Takahashi, A., Kawakami, E., Sugishita, H., Sakaue, S., Matoba, N., Low, S.-K., Okada, Y., Terao, C., Amariuta, T., Gazal, S., Kochi, Y., Horikoshi, M., Suzuki, K., Ito, K., Koyama, S., Ozaki, K., ... Kamatani, Y. (2020). Large-scale genome-wide association study in a Japanese population identifies novel susceptibility loci across different diseases. *Nature Genetics*, 52(7), 669–679. <https://doi.org/10.1038/s41588-020-0640-3>

• Principe, S., Porsbjerg, C., Bolm Ditlev, S., Kjærsgaard Klein, D., Golebski, K., Dyhre?Petersen, N., van Dijk, Y. E., van Bragt, J. J. M. H., Dankelman, L. L. H., Dahlen, S., Brightling, C. E., Vijverberg, S. J. H., & Maitland?van der Zee, A. H. (2021). Treating severe asthma: Targeting the IL75 pathway. *Clinical and Experimental Allergy*, 51(8), 992–1005. <https://doi.org/10.1111/cea.13885>

• Zhu, W., Liu, N., Zhao, Y., Jia, H., Cui, B., & Ning, G. (2010). Association analysis of polymorphisms in IL-3, IL-4, IL-5, IL-9, and IL-13 with Graves' disease. *Journal of Endocrinological Investigation*, 33(10), 751–755. <https://doi.org/10.1007/BF03346682>

• Ganesh, B. B., Bhattacharya, P., Gopisetty, A., & Prabhakar, B. S. (2011). Role of Cytokines in the Pathogenesis and Suppression of Thyroid Autoimmunity. *Journal of Interferon & Cytokine Research*, 31(10), 721–731. <https://doi.org/10.1089/jir.2011.0049>

• Mestiri, S., Zaafer, I., Inoubli, O., Abid, N., Omrani, A., Nejehi, H., & Marmouch, H. (2020). Association of cytokine Th2 gene polymorphisms with autoimmune thyroid diseases in Tunisian population. *International Journal of Immunogenetics*, 47(3), 294–308. <https://doi.org/10.1111/iji.12472>

• Kabesch, M., Depner, M., Dahmen, I., Weiland, S. K., Vogelberg, C., Niggemann, B., Lau, S., Illig, T., Klopp, N., Wahn, U., Reinhardt, D., von Mutius, E., & Nickel, R. (2007). Polymorphisms in eosinophil pathway genes, asthma and atopy. *Allergy*, 62(4), 423–428. <https://doi.org/10.1111/j.1398-9995.2006.01300.x>

IL6

• Carini, M., Fredi, M., Cavazzana, I., Bresciani, R., Ferrari, F., Monti, E., Franceschini, F., & Biasiotto, G. (2023). Frequency Evaluation of the Interleukin-6 ?174G>C Polymorphism and Homeostatic Iron Regulator (HFE) Mutations as Disease Modifiers in Patients Affected by Systemic Lupus Erythematosus and Rheumatoid Arthritis. *International Journal of Molecular Sciences*, 24(22), 16300. <https://doi.org/10.3390/ijms242216300>

• Zhu, S., Wang, B., Jia, Q., & Duan, L. (2019). Candidate single nucleotide polymorphisms of irritable bowel syndrome: A systemic review and meta-analysis. *BMC Gastroenterology*, 19(1), 165. <https://doi.org/10.1186/s12876-019-1084-z>

• Wang, X., Yan, Z., & Ye, Q. (2019). Interleukin-6 gene polymorphisms and susceptibility to liver diseases: A meta-analysis. *Medicine*, 98(50), e18408. <https://doi.org/10.1097/MD.00000000000018408>

• Wu, W., Clark, E. A. S., Stoddard, G. J., Watkins, W. S., Espin, M. S., Manuck, T. A., Xing, J., Varner, M. W., & Jorde, L. B. (2013). Effect of interleukin-6 polymorphism on risk of preterm birth within population strata: A meta-analysis. *BMC Genetics*, 14, 30. <https://doi.org/10.1186/1471-2156-14-30>

• Tanaka, T., Narazaki, M., & Kishimoto, T. (2014). IL-6 in Inflammation, Immunity, and Disease. *Cold Spring Harbor Perspectives in Biology*, 6(10), a016295. <https://doi.org/10.1101/cshperspect.a016295>

• Nie, G., Xie, C. L., Cao, Y. J., Xu, M. M., Shi, X., Zou, A. L., & Qi, J. H. (2016). Meta-analysis of IL-6 -174G/C polymorphism and psoriasis risk. *Genetics and Molecular Research: GMR*, 15(2). <https://doi.org/10.4238/gmr.15028255>

• Fishman, D., Faulds, G., Jeffery, R., Mohamed-Ali, V., Yudkin, J. S., Humphries, S., & Woo, P. (1998). The effect of novel polymorphisms in the interleukin-6 (IL-6) gene on IL-6 transcription and plasma IL-6 levels, and an association with systemic-onset juvenile chronic arthritis. *Journal of Clinical Investigation*, 102(7), 1369–1376. • Cheng, H., Zhu, W., Zhu, M., Sun, Y., Sun, X., Jia, D., Yang, C., Yu, H., & Zhang, C. (2021). Meta-analysis: Interleukin 6 gene -174G/C polymorphism associated with type 2 diabetes mellitus and interleukin 6 changes. *Journal of Cellular and Molecular Medicine*, 25(12), 5628–5639. <https://doi.org/10.1111/jcmm.16575>

• Chen, L., Zhang, Z., Huang, J., & Jin, M. (2018). Association between rs1800795 polymorphism in the interleukin-6 gene and the risk of polycystic ovary syndrome: A meta-analysis. *Medicine*, 97(29), e11558. <https://doi.org/10.1097/MD.00000000000011558>

NOS2

• Bouzigon, E., Monier, F., Boussaha, M., Le Moul, N., Huyvaert, H., Matran, R., Letort, S., Bousquet, J., Pin, I., Lathrop, M., Kauffmann, F., Demenais, F., & Nadif, R. (2012). Associations between Nitric Oxide Synthase Genes and Exhaled NO-Related Phenotypes according to Asthma Status. *PLoS ONE*, 7(5), e36672. <https://doi.org/10.1371/journal.pone.0036672>

• Stuart, P. E., Nair, R. P., Ellinghaus, E., Ding, J., Tejasvi, T., Gudjonsson, J. E., Li, Y., Weidinger, S., Eberlein, B., Gieger, C., Wichmann, H. E., Kunz, M., Ike, R., Krueger, G. G., Bowcock, A. M., Mrowietz, U., Lim, H. W., Voorhees, J. J., Abecasis, G. R., ... Elder, J. T. (2010). Genome-wide association analysis identifies three psoriasis susceptibility loci. *Nature Genetics*, 42(11), 1000–1004. <https://doi.org/10.1038/ng.693>

STAT4

• Lee, H.-S., Park, H., Yang, S., Kim, D., & Park, Y. (2008). STAT4 polymorphism is associated with early-onset type 1 diabetes, but not with late-onset type 1 diabetes. *Annals of the New York Academy of Sciences*, 1150, 93–98. <https://doi.org/10.1196/annals.1447.013>

• Jiang, Y., Zhang, R., Zheng, J., Liu, P., Tang, G., Lv, H., Zhang, L., Shang, Z., Zhan, Y., Lv, W., Shi, M., & Zhang, R. (2012). Meta-analysis of 125 rheumatoid arthritis-related single nucleotide polymorphisms studied in the past two decades. *PLoS One*, 7(12), e51571. <https://doi.org/10.1371/journal.pone.0051571>

• Yan, N., Meng, S., Zhou, J., Xu, J., Muhali, F. S., Jiang, W., Shi, L., Shi, X., & Zhang, J. (2014). Association between STAT4 gene polymorphisms and autoimmune thyroid diseases in a Chinese population. *International Journal of Molecular Sciences*, 15(7), 12280–12293. <https://doi.org/10.3390/ijms150712280>

• Lee, H.-S., Remmers, E. F., Le, J. M., Kastner, D. L., Bae, S.-C., & Gregersen, P. K. (2007). Association of STAT4 with rheumatoid arthritis in the Korean population. *Molecular Medicine (Cambridge, Mass.)*, 13(9–10), 455–460. <https://doi.org/10.2119/2007-00072.Lee>

• Namjou, B., Sestak, A. L., Armstrong, D. L., Zidovetzki, R., Kelly, J. A., Jacob, N., Ciobanu, V., Kaufman, K. M., Ojwang, J. O., Ziegler, J. Q., Quismorio, F., Reiff, A., Myones, B.

L., Guthridge, J. M., Nath, S. K., Bruner, G. R., Mehriani-Shai, R., Silverman, E., Klein-Gitelman, M., ... Jacob, C. O. (2009). High density genotyping of STAT4 gene reveals multiple haplotypic associations with Systemic Lupus Erythematosus in different racial groups. *Arthritis and Rheumatism*, 60(4), 1085–1095. <https://doi.org/10.1002/art.24387> • Sigurdsson, S., Nordmark, G., Garnier, S., Grundberg, E., Kwan, T., Nilsson, O., Eioranta, M.-L., Gunnarsson, I., Svenungsson, E., Sturfelt, G., Bengtsson, A. A., Jönsson, A., Truedsson, L., Rantapää-Dahlqvist, S., Eriksson, C., Alm, G., Göring, H. H. H., Pastinen, T., Syyänen, A.-C., & Rönnblom, L. (2008). A risk haplotype of STAT4 for systemic lupus erythematosus is over-expressed, correlates with anti-dsDNA and shows additive effects with two risk alleles of IRF5. *Human Molecular Genetics*, 17(18), 2868–2876. <https://doi.org/10.1093/hmg/ddn184>

TNF-?

• Kilpeläinen, T. O., Laaksonen, D. E., Lakka, T. A., Herder, C., Koenig, W., Lindström, J., Eriksson, J. G., Uusitupa, M., Kolb, H., Laakso, M., Tuomilehto, J., & Finnish Diabetes Prevention Study. (2010). The rs1800629 polymorphism in the TNF gene interacts with physical activity on the changes in C-reactive protein levels in the Finnish Diabetes Prevention Study. *Experimental and Clinical Endocrinology & Diabetes: Official Journal, German Society of Endocrinology [and] German Diabetes Association*, 118(10), 757–759. <https://doi.org/10.1055/s-0030-1249686> • Yang, G., Chen, J., Xu, F., Bao, Z., Yao, Y., & Zhou, J. (2014). Association between Tumor Necrosis Factor-? rs1800629 Polymorphism and Risk of Asthma: A Meta-Analysis. *PLoS ONE*, 9(6), e99962. <https://doi.org/10.1371/journal.pone.0099962> • Song, G. G., Seo, Y. H., Kim, J.-H., Choi, S. J., Ji, J. D., & Lee, Y. H. (2015). Association between TNF-? (-308 A/G, -238 A/G, -857 C/T) polymorphisms and responsiveness to TNF-? blockers in spondyloarthritis, psoriasis and Crohn's disease: A meta-analysis. *Pharmacogenomics*, 16(12), 1427–1437. <https://doi.org/10.2217/pgs.15.90> • Kroeger, K. M., Carville, K. S., & Abraham, L. J. (1997). The -308 tumor necrosis factor-alpha promoter polymorphism effects transcription. *Molecular Immunology*, 34(5), 391–399. [https://doi.org/10.1016/s0161-5890\(97\)00052-7](https://doi.org/10.1016/s0161-5890(97)00052-7) • de Luis, D. A., Aller, R., Izaola, O., Gonzalez Sagrado, M., & Conde, R. (2013). Role of G308 promoter variant of tumor necrosis factor alpha gene on weight loss and metabolic parameters after a high monounsaturated versus a high polyunsaturated fat hypocaloric diets. *Medicina Clinica*, 141(5), 189–193. <https://doi.org/10.1016/j.medcli.2012.12.021> • Loures, M. A. R., Alves, H. V., de Moraes, A. G., Santos, T. da S., Lara, F. F., Neves, J. S. F., Macedo, L. C., Teixeira, J. J. V., Sell, A. M., & Visentainer, J. E. L. (2019). Association of TNF, IL12, and IL23 gene polymorphisms and psoriatic arthritis: Meta-analysis. *Expert Review of Clinical Immunology*, 15(3), 303–313. <https://doi.org/10.1080/1744666X.2019.1564039> • Tu, Y., Fan, G., Zeng, T., Cai, X., & Kong, W. (2018). Association of TNF-? promoter polymorphism and Graves' disease: An updated systematic review and meta-analysis. *Bioscience Reports*, 38(2), BSR20180143. <https://doi.org/10.1042/BSR20180143> • Chen, L., Huang, Z., Liao, Y., Yang, B., & Zhang, J. (2019). Association between tumor necrosis factor polymorphisms and rheumatoid arthritis as well as systemic lupus erythematosus: A meta-analysis. *Brazilian Journal of Medical and Biological Research = Revista Brasileira De Pesquisas Medicas E Biologicas*, 52(3), e7927. <https://doi.org/10.1590/1414-431X20187927>

METHYLATION SNP References

BHMT rs3733890

• Ren, B., Tian, D., Wang, L., Han, H., Wang, W., Tian, H., Yue, L., & Zhang, W. (2019). Association between the BHMT gene rs3733890 polymorphism and the efficacy of oral folate therapy in patients with hyperhomocysteinemia. *Annals of Human Genetics*, 83(6), 434–444. <https://doi.org/10.1111/ahg.12326> • Zhang, W.-H., Zhao, S.-M., Guo, J.-F., Liu, Y.-P., & Jiang, Y.-Q. (2023). BHMT polymorphism and susceptibility to PTE in Chinese patients. *European Review for Medical and Pharmacological Sciences*, 27(9), 4098–4102. https://doi.org/10.26355/eurrev_202305_32317 • Li, F., Feng, Q., Lee, C., Wang, S., Palleymounter, L. L., Moon, I., Eckloff, B. W., Wieben, E. D., Schaid, D. J., Yee, V., & Weinshilboum, R. M. (2008). Human betaine-homocysteine methyltransferase (BHMT) and BHMT2: Common gene sequence variation and functional characterization. *Molecular Genetics and Metabolism*, 94(3), 326–335. <https://doi.org/10.1016/j.ymgme.2008.03.013> • Li, D., Yang, J., Zhao, Q., Zhang, C., Ren, B., Yue, L., Du, B., Godfrey, O., Huang, X., & Zhang, W. (2019). Genetic and epigenetic regulation of BHMT is associated with folate therapy efficacy in hyperhomocysteinemia. *Asia Pacific Journal of Clinical Nutrition*, 28(4), 879–887. [https://doi.org/10.6133/apjcn.201912_28\(4\).0025](https://doi.org/10.6133/apjcn.201912_28(4).0025) • Ilizumbia, M. N., Cheng, T.-Y. D., Neuhouser, M. L., Miller, J. W., Beresford, S. A. A., Duggan, A. T., Toriola, A. T., Song, X., Zheng, Y., Bailey, L. B., Shadyab, A. H., Liu, S., Malysheva, O., Caudill, M. A., & Ulrich, C. M. (2020). Associations between Plasma Choline Metabolites and Genetic Polymorphisms in One-Carbon Metabolism in Postmenopausal Women: The Women's Health Initiative Observational Study. *The Journal of Nutrition*, 150(11), 2874–2881. <https://doi.org/10.1093/njn/nxaa266> • Ganz, A. B., Klatt, K. C., & Caudill, M. A. (2017). Common Genetic Variants Alter Metabolism and Influence Dietary Choline Requirements. *Nutrients*, 9(6), 637. <https://doi.org/10.3390/nu9080637> • Chen, X., Huang, X., Zheng, C., Wang, X., & Zhang, W. (2022). Using the optimal method-explained variance weighted genetic risk score to predict the efficacy of folic acid therapy to hyperhomocysteinemia. *European Journal of Clinical Nutrition*, 76(7), 943–949. <https://doi.org/10.1038/s41430-021-01055-5>

CBS rs234706

• Aras, O., Hanson, N. Q., Yang, F., & Tsai, M. Y. (2000). Influence of 699C-->T and 1080C-->T polymorphisms of the cystathionine beta-synthase gene on plasma homocysteine levels. *Clinical Genetics*, 58(6), 455–459. <https://doi.org/10.1034/j.1399-0004.2000.580605.x> • De Stefano, V., Dekou, V., Nicaud, V., Chasse, J. F., London, J., Stansbie, D., Humphries, S. E., & Gudnason, V. (1998). Linkage disequilibrium at the cystathionine beta synthase (CBS) locus and the association between genetic variation at the CBS locus and plasma levels of homocysteine. *The Ears II Group. European Atherosclerosis Research Study. Annals of Human Genetics*, 62(Pt 6), 481–490. <https://doi.org/10.1046/j.1469-1809.1998.6260481.x> • Fredriksen, A., Meyer, K., Ueland, P. M., Vollset, S. E., Grotmol, T., & Schneede, J. (2007). Large-scale population-based metabolic phenotyping of thirteen genetic polymorphisms related to one-carbon metabolism. *Human Mutation*, 28(9), 856–865. <https://doi.org/10.1002/humu.20522> • Kruger, W. D., Evans, A. A., Wang, L., Malinow, M. R., Duell, P. B., Anderson, P. H., Block, P. C., Hess, D. L., Graf, E. E., & Upson, B. (2000). Polymorphisms in the CBS gene associated with decreased risk of coronary artery disease and increased responsiveness to total homocysteine lowering by folic acid. *Molecular Genetics and Metabolism*, 70(1), 53–60. <https://doi.org/10.1006/mgme.2000.2993>

CUBN rs1801222

• Grarup, N., Sulem, P., Sandholt, C. H., Thorleifsson, G., Ahluwalia, T. S., Steinthorsdottir, V., Bjarnason, H., Gudbjartsson, D. F., Magnusson, O. T., Sparsø, T., Albrechtsen, A., Kong, A., Masson, G., Tian, G., Cao, H., Nie, C., Kristiansen, K., Husemoen, L. L., Thuesen, B., ... Pedersen, O. (2013). Genetic architecture of vitamin B12 and folate levels uncovered applying deeply sequenced large datasets. *PLoS Genetics*, 9(6), e1003530. <https://doi.org/10.1371/journal.pgen.1003530> • van Meurs, J. B. J., Pare, G., Schwartz, S. M., Hazra, A., Tanaka, T., Vermeulen, S. H., Cotlarciuc, I., Yuan, X., Mälarstig, A., Bandinelli, S., Bis, J. C., Blom, H., Brown, M. J., Chen, C., Chen, Y.-D., Clarke, R. J., Dehghan, A., Erdmann, J., Ferrucci, L., ... Ahmadi, K. R. (2013). Common genetic loci influencing plasma homocysteine concentrations and their effect on risk of coronary artery disease. *The American Journal of Clinical Nutrition*, 98(3), 668–676. <https://doi.org/10.3945/ajcn.112.044545> • Panglilan, F., Molloy, A. M., Mills, J. L., Troendle, J. F., Parle-McDermott, A., Signore, C., O'Leary, V. B., Chines, P., Seay, J. M., Geiler-Samerotte, K., Mitchell, A., VanderMeer, J. E., Krebs, K. M., Sanchez, A., Corman-Homonoff, J., Stone, N., Conley, M., Kirke, P. N., Shane, B., ... Brody, L. C. (2012). Evaluation of common genetic variants in 82 candidate genes as risk factors for neural tube defects. *BMC Medical Genetics*, 13, 62. <https://doi.org/10.1186/1471-2350-13-62> • Keene, K. L., Chen, W.-M., Chen, F., Williams, S. R., Elkhatib, S. D., Hsu, F.-C., Mychaleckyj, J. C., Doherty, K. F., Pugh, E. W., Ling, H., Laurie, C. C., Gogarten, S. M., Madden, E. B., Worrall, B. B., & Sale, M. M. (2014). Genetic Associations with Plasma B12, B6, and Folate Levels in an Ischemic Stroke Population from the Vitamin Intervention for Stroke Prevention (VISP) Trial. *Frontiers in Public Health*, 2, 112. <https://doi.org/10.3389/fpubh.2014.00112> • Hazra, A., Kraft, P., Lazarus, R., Chen, C., Chanock, S. J., Jacques, P., Selhub, J., & Hunter, D. J. (2009). Genome-wide significant predictors of metabolites in the one-carbon metabolism pathway. *Human Molecular Genetics*, 18(23), 4677–4687. <https://doi.org/10.1093/hmg/ddp428>

DHFR rs70991108

• Kalmbach, R. D., Choumenkovitch, S. F., Troen, A. P., Jacques, P. F., D'Agostino, R., & Selhub, J. (2008). A 19-base pair deletion polymorphism in dihydrofolate reductase is associated with increased unmetabolized folic acid in plasma and decreased red blood cell folate. *The Journal of Nutrition*, 138(12), 2323–2327. <https://doi.org/10.3945/jn.108.096404> • Johnson, W. G., Stenroos, E. S., Spychala, J. R., Chatkupt, S., Ming, S. X., & Buyske, S. (2004). New 19 bp deletion polymorphism in intron-1 of dihydrofolate reductase (DHFR): A risk factor for spina bifida acting in mothers during pregnancy? *American Journal of Medical Genetics. Part A*, 124A(4), 339–345. <https://doi.org/10.1002/ajmg.a.20505> • Johnson, W. G., Scholl, T. O., Spychala, J. R., Buyske, S., Stenroos, E. S., & Chen, X. (2005). Common dihydrofolate reductase 19-base pair deletion allele: A novel risk factor for preterm delivery. *The American Journal of Clinical Nutrition*, 81(3), 664–668. <https://doi.org/10.1093/ajcn/81.3.664> • Philip, D., Buch, A., Moorthy, D., Scott, T. M., Parnell, L. D., Lai, C.-Q., Ordovás, J. M., Selhub, J., Rosenberg, I. H., Tucker, K. L., & Troen, A. M. (2015). Dihydrofolate reductase 19-bp deletion polymorphism modifies the association of folate status with memory in a cross-sectional multi-ethnic study of adults. *The American Journal of Clinical Nutrition*, 102(5), 1279–1288. <https://doi.org/10.3945/ajcn.115.111054> • Noé, V., MacKenzie, S., & Ciudad, C. J. (2003). An intron is required for dihydrofolate reductase protein stability. *The Journal of Biological Chemistry*, 278(40), 38292–38300. <https://doi.org/10.1074/jbc.M212746200>

FOLR1

• Zhu, S., Ni, G., Sui, L., Zhao, Y., Zhang, X., Dai, Q., Chen, A., Lin, W., Li, Y., Huang, M., & Zhou, L. (2021). Genetic Polymorphisms in Enzymes Involved in One-Carbon Metabolism and Anti-epileptic Drug Monotherapy on Homocysteine Metabolism in Patients With Epilepsy. *Frontiers in Neurology*, 12, 683275. <https://doi.org/10.3389/fneur.2021.683275> • Song, X., Wei, J., Shu, J., Liu, Y., Sun, M., Zhu, P., & Qin, J. (2022). Association of polymorphisms of FOLR1 gene and maternal folic acid supplementation with risk of ventricular septal defect: A case-control study. *European Journal of Clinical Nutrition*, 76(9), 1273–1280. <https://doi.org/10.1038/s41430-022-01110-9> • Laanpere, M., Altmäe, S., Kaart, T., Stavreus-Evers, A., Nilsson, T. K., & Salumets, A. (2011). Folate-metabolizing gene variants and pregnancy outcome of IVF. *Reproductive Biomedicine Online*, 22(6), 603–614. <https://doi.org/10.1016/j.rbmo.2011.03.002>

MTHFD1

• Ding, Y. P., Pedersen, E. K. R., Johansson, S., Gregory, J. F., Ueland, P. M., Svingen, G. F. T., Helgeland, Ø., Meyer, K., Fredriksen, Å., & Nygård, O. K. (2016). B vitamin treatments modify the risk of myocardial infarction associated with a MTHFD1 polymorphism in patients with stable angina pectoris. *Nutrition, Metabolism and Cardiovascular Diseases*, 26(6), 495–501. <https://doi.org/10.1016/j.numecd.2015.12.009> • Carroll, N., Panglilan, F., Molloy, A. M., Troendle, J., Mills, J. L., Kirke, P. N., Brody, L. C., Scott, J. M., & Parle-McDermott, A. (2009). Analysis of the MTHFD1 promoter and risk of neural tube defects. *Human Genetics*, 125(3), 247–256. <https://doi.org/10.1007/s00439-008-0616-3>

MTHFR rs1801131

• Ducker, G. S., & Rabinowitz, J. D. (2017). One-Carbon Metabolism in Health and Disease. *Cell Metabolism*, 25(1), 27–42. <https://doi.org/10.1016/j.cmet.2016.08.009> • Weisberg, I. S., Jacques, P. F., Selhub, J., Bostom, A. G., Chen, Z., Curtis Ellison, R., Eckfeldt, J. H., & Rozen, R. (2001). The 1298A-->C polymorphism in methyltetrahydrofolate reductase (MTHFR): In vivo expression and association with homocysteine. *Atherosclerosis*, 156(2), 409–415. [https://doi.org/10.1016/s0021-9150\(00\)00671-7](https://doi.org/10.1016/s0021-9150(00)00671-7) • Wei, L. K., Au, A., Menon, S., Griffiths, L. R., Kooi, C. W., Irene, L., Zhao, J., Lee, C., Alekseeva, A. M., Hassan, M. R. A., & Aziz, Z. A. (2017). Polymorphisms of MTHFR, eNOS, ACE, AGT, ApoE, PON1, PDE4D, and Ischemic Stroke: Meta-Analysis. *Journal of Stroke and Cerebrovascular Diseases: The Official Journal of National Stroke Association*, 26(11), 2482–2493. <https://doi.org/10.1016/j.jstrokecerebrovasdis.2017.05.048> • van der Put, N. M., Gabreëls, F., Stevens, E.

M., Smeitink, J. A., Trijbels, F. J., Eskes, T. K., van den Heuvel, L. P., & Blom, H. J. (1998). A second common mutation in the methylenetetrahydrofolate reductase gene: An additional risk factor for neural-tube defects? *American Journal of Human Genetics*, 62(5), 1044–1051. <https://doi.org/10.1086/301825> • Shen, O., Liu, R., Wu, W., Yu, L., & Wang, X. (2012). Association of the methylenetetrahydrofolate reductase gene A1298C polymorphism with male infertility: A meta-analysis. *Annals of Human Genetics*, 76(1), 25–32. <https://doi.org/10.1111/j.1469-1809.2011.00691.x> • Ogino, S., & Wilson, R. B. (2003). Genotype and haplotype distributions of MTHFR677C>T and 1298A>C single nucleotide polymorphisms: A meta-analysis. *Journal of Human Genetics*, 48(1), 1–7. <https://doi.org/10.1007/s100380300000> • Donnelly, J. G. (2000). The 1298(A-->C) mutation of methylenetetrahydrofolate reductase should be designated to the 1289 position of the gene. *American Journal of Human Genetics*, 66(2), 744–745. <https://doi.org/10.1086/302784> • Frosst, P., Blom, H. J., Milos, R., Goyette, P., Sheppard, C. A., Matthews, R. G., Boers, G. J., den Heijer, M., Kluijtmans, L. A., & van den Heuvel, L. P. (1995). A candidate genetic risk factor for vascular disease: A common mutation in methylenetetrahydrofolate reductase. *Nature Genetics*, 10(1), 111–113. <https://doi.org/10.1038/ng0595-111>

MTHFR rs1801133

• van der Put, N. M., Steegers-Theunissen, R. P., Frosst, P., Trijbels, F. J., Eskes, T. K., van den Heuvel, L. P., Mariman, E. C., den Heyer, M., Rozen, R., & Blom, H. J. (1995). Mutated methylenetetrahydrofolate reductase as a risk factor for spina bifida. *Lancet (London, England)*, 346(8982), 1070–1071. [https://doi.org/10.1016/s0140-6736\(95\)91743-8](https://doi.org/10.1016/s0140-6736(95)91743-8) • Christensen, B., Arbour, L., Tran, P., Leclerc, D., Sabbaghian, N., Platt, R., Gilfix, B. M., Rosenblatt, D. S., Gravel, R. A., Forbes, P., & Rozen, R. (1999). Genetic polymorphisms in methylenetetrahydrofolate reductase and methionine synthase, folate levels in red blood cells, and risk of neural tube defects. *American Journal of Medical Genetics*, 84(2), 151–157. [https://doi.org/10.1002/\(sici\)1096-8628\(19990521\)84:2<151::aid-ajmg12-3.0.co;2-t](https://doi.org/10.1002/(sici)1096-8628(19990521)84:2<151::aid-ajmg12-3.0.co;2-t) • Frosst, P., Blom, H. J., Milos, R., Goyette, P., Sheppard, C. A., Matthews, R. G., Boers, G. J., den Heijer, M., Kluijtmans, L. A., & van den Heuvel, L. P. (1995). A candidate genetic risk factor for vascular disease: A common mutation in methylenetetrahydrofolate reductase. *Nature Genetics*, 10(1), 111–113. <https://doi.org/10.1038/ng0595-111> • Hazra, A., Kraft, P., Lazarus, R., Chen, C., Chanock, S. J., Jacques, P., Selhub, J., & Hunter, D. J. (2009). Genome-wide significant predictors of metabolites in the one-carbon metabolism pathway. *Human Molecular Genetics*, 18(23), 4677–4687. <https://doi.org/10.1093/hmg/ddp428> • Hong, H.-H., Hu, Y., Yu, X.-Q., Zhou, L., Lv, M.-Q., Sun, Y., Ren, W.-J., & Zhou, D.-X. (2017). Associations of C677T polymorphism in methylenetetrahydrofolate reductase (MTHFR) gene with male infertility risk: A meta-analysis. *European Journal of Obstetrics, Gynecology, and Reproductive Biology*, 212, 101–109. <https://doi.org/10.1016/j.ejogrb.2017.03.004> • Kowa, H., Yasui, K., Takeshima, T., Urakami, K., Sakai, F., & Nakashima, K. (2000). The homozygous C677T mutation in the methylenetetrahydrofolate reductase gene is a genetic risk factor for migraine. *American Journal of Medical Genetics*, 96(6), 762–764. [https://doi.org/10.1002/1096-8628\(20001204\)96:6<762::aid-ajmg12-3.0.co;2-x](https://doi.org/10.1002/1096-8628(20001204)96:6<762::aid-ajmg12-3.0.co;2-x) • Ogino, S., & Wilson, R. B. (2003). Genotype and haplotype distributions of MTHFR677C>T and 1298A>C single nucleotide polymorphisms: A meta-analysis. *Journal of Human Genetics*, 48(1), 1–7. <https://doi.org/10.1007/s100380300000> • Shadrina, M., Bondarenko, E. A., & Slominsky, P. A. (2018). Genetics Factors in Major Depression Disease. *Frontiers in Psychiatry*, 9, 334. <https://doi.org/10.3389/fpsy.2018.00334> • Wei, L. K., Au, A., Menon, S., Griffiths, L. R., Kooi, C. W., Irene, L., Zhao, J., Lee, C., Alekseevna, A. M., Hassan, M. R. A., & Aziz, Z. A. (2017). Polymorphisms of MTHFR, eNOS, ACE, AGT, ApoE, PON1, PDE4D, and Ischemic Stroke: Meta-Analysis. *Journal of Stroke and Cerebrovascular Diseases: The Official Journal of National Stroke Association*, 26(11), 2482–2493. <https://doi.org/10.1016/j.jstrokecerebrovasdis.2017.05.048> • Weisberg, I. S., Jacques, P. F., Selhub, J., Bostom, A. G., Chen, Z., Curtis Ellison, R., Eckfeldt, J. H., & Rozen, R. (2001). The 1298A-->C polymorphism in methylenetetrahydrofolate reductase (MTHFR): In vitro expression and association with homocysteine. *Atherosclerosis*, 156(2), 409–415. [https://doi.org/10.1016/s0021-9150\(00\)00671-7](https://doi.org/10.1016/s0021-9150(00)00671-7)

MTR

• Weiner, A. S., Boyarskikh, U. A., Voronina, E. N., Mishukova, O. V., & Filipenko, M. L. (2014). Methylenetetrahydrofolate reductase C677T and methionine synthase A2756G polymorphisms influence on leukocyte genomic DNA methylation level. *Gene*, 533(1), 168–172. <https://doi.org/10.1016/j.gene.2013.09.098> • Saha, T., Chatterjee, M., Sinha, S., Rajamma, A., & Mukhopadhyay, K. (2017). Components of the folate metabolic pathway and ADHD core traits: An exploration in eastern Indian probands. *Journal of Human Genetics*, 62(7), 687–695. <https://doi.org/10.1038/jhg.2017.23> • Guedes, T., Santos, A. A., Vieira-Neto, F. H., Bianco, B., Barbosa, C. P., & Christofolini, D. M. (2022). Folate metabolism abnormalities in infertile patients with endometriosis. *Biomarkers in Medicine*, 16(7), 549–557. <https://doi.org/10.2217/bmm-2021-0076> • Dekou, V., Gudnason, V., Hawn, E., Miller, G. J., Stansbie, D., & Humphries, S. E. (2001). Gene-environment and gene-gene interaction in the determination of plasma homocysteine levels in healthy middle-aged men. *Thrombosis and Haemostasis*, 85(1), 67–74. • Chen, J., Stamper, M. J., Ma, J., Selhub, J., Malinow, M. R., Hennekens, C. H., & Hunter, D. J. (2001). Influence of a methionine synthase (D919G) polymorphism on plasma homocysteine and folate levels and relation to risk of myocardial infarction. *Atherosclerosis*, 154(3), 667–672. [https://doi.org/10.1016/s0021-9150\(00\)0469-x](https://doi.org/10.1016/s0021-9150(00)0469-x) • Summers, C. M., Mitchell, L. E., Stanislawski-Sachadyn, A., Baido, S. F., Blair, I. A., Von Feldt, J. M., & Whitehead, A. S. (2010). Genetic and Lifestyle Variables Associated with Homocysteine Concentrations and the Distribution of Folate Derivatives in Healthy Premenopausal Women. *Birth Defects Research. Part A, Clinical and Molecular Teratology*, 88(8), 679–688. <https://doi.org/10.1002/bdra.20683>

MTRR rs1801394

• Wilson, A., Platt, R., Wu, Q., Leclerc, D., Christensen, B., Yang, H., Gravel, R. A., & Rozen, R. (1999). A common variant in methionine synthase reductase combined with low cobalamin (vitamin B12) increases risk for spina bifida. *Molecular Genetics and Metabolism*, 67(4), 317–323. <https://doi.org/10.1006/mgme.1999.2879> • Leclerc, D., Wilson, A., Dumas, R., Gafuik, C., Song, D., Watkins, D., Heng, H. H., Rommens, J. M., Scherer, S. W., Rosenblatt, D. S., & Gravel, R. A. (1998). Cloning and mapping of a cDNA for methionine synthase reductase, a flavoprotein defective in patients with homocystinuria. *Proceedings of the National Academy of Sciences of the United States of America*, 95(6), 3059–3064. <https://doi.org/10.1073/pnas.95.6.3059> • Kwon, B. N., Lee, N. R., Kim, H. J., Kang, Y. D., Kim, J. S., Park, J. W., & Jin, H. J. (2021). Folate metabolizing gene polymorphisms and genetic vulnerability to preterm birth in Korean women. *Genes & Genomics*, 43(8), 937–945. <https://doi.org/10.1007/s13258-021-01082-3> • Hobbs, C. A., Sherman, S. L., Yi, P., Hopkins, S. E., Torfs, C. P., Hine, R. J., Pogribna, M., Rozen, R., & James, S. J. (2000). Polymorphisms in genes involved in folate metabolism as maternal risk factors for Down syndrome. *American Journal of Human Genetics*, 67(3), 623–630. <https://doi.org/10.1086/303055> • Du, B., Tian, H., Tian, D., Zhang, C., Wang, W., Wang, L., Ge, M., Hou, Q., & Zhang, W. (2018). Genetic polymorphisms of key enzymes in folate metabolism affect the efficacy of folate therapy in patients with hyperhomocysteinaemia. *The British Journal of Nutrition*, 119(8), 887–895. <https://doi.org/10.1017/S0007114518000508> • Cho, S. H., Kim, J. H., An, H. J., Kim, J. O., Kim, Y. R., Lee, W. S., & Kim, N. K. (2021). Association of methionine synthase (rs1801394), methionine synthase reductase (rs1801394), and methylenetetrahydrofolate dehydrogenase 1 (rs2236225) genetic polymorphisms with recurrent implantation failure. *Human Fertility (Cambridge, England)*, 24(3), 161–168. <https://doi.org/10.1080/14647273.2019.1613679> • Chatterjee, M., Saha, T., Maitra, S., Sinha, S., & Mukhopadhyay, K. (2020). Folate System Gene Variant rs1801394 66A>G may have a Causal Role in Down Syndrome in the Eastern Indian Population. *International Journal of Molecular and Cellular Medicine*, 9(3), 215–224. <https://doi.org/10.22088/IJMCMB.BUMS.9.3.215> • Li, D., Zhao, Q., Huang, X., Zhang, C., Godfrey, O., & Zhang, W. (2020). Association of genetic and epigenetic variants in one-carbon metabolism gene with folate treatment response in hyperhomocysteinaemia. *European Journal of Clinical Nutrition*, 74(7), 1073–1083. <https://doi.org/10.1038/s41430-020-0611-x> • Oltman, H., Munson, T., & Banerjee, R. (2002). Differences in the efficiency of reductive activation of methionine synthase and exogenous electron acceptors between the common polymorphic variants of human methionine synthase reductase. *Biochemistry*, 41(45), 13378–13385. <https://doi.org/10.1021/bi020536s>

TCN1

• Hall, C. A. (1975). Transcobalamins I and II as natural transport proteins of vitamin B12. *Journal of Clinical Investigation*, 56(5), 1125–1131. • Tanaka, T., Scheet, P., Giusti, B., Bandinelli, S., Piras, M. G., Usala, G., Lai, S., Mulas, A., Corsi, A. M., Vestriani, A., Sofi, F., Gori, A. M., Abbate, R., Guralnik, J., Singleton, A., Abecasis, G. R., Schlessinger, D., Uda, M., & Ferrucci, L. (2009). Genome-wide association study of vitamin B6, vitamin B12, folate, and homocysteine blood concentrations. *American Journal of Human Genetics*, 84(4), 477–482. <https://doi.org/10.1016/j.ajhg.2009.02.011> • Velkova, A., Diaz, J. E. L., Pangilinan, F., Molloy, A. M., Mills, J. L., Shane, B., Sanchez, E., Cunningham, C., McNulty, H., Cropp, C. D., Bailey-Wilson, J. E., Wilson, A. F., & Brody, L. C. (2017). The FUT2 secretor variant p.Trp154Ter influences serum vitamin B12 concentration via holo-haptocorrin, but not holo-transcobalamin, and is associated with haptocorrin glycosylation. *Human Molecular Genetics*, 26(24), 4975–4988. <https://doi.org/10.1093/hmg/ddx369>

TCN2

• Miller, J. W., Ramos, M. I., Garrod, M. G., Flynn, M. A., & Green, R. (2002). Transcobalamin II 775G>C polymorphism and indices of vitamin B12 status in healthy older adults. *Blood*, 100(2), 718–720. <https://doi.org/10.1182/blood-2002-01-0209> • Oussalah, A., Levy, J., Filhine-Trésarrieu, P., Namour, F., & Guéant, J.-L. (2017). Association of TCN2 rs1801198 c.776G>C polymorphism with markers of one-carbon metabolism and related diseases: A systematic review and meta-analysis of genetic association studies. *The American Journal of Clinical Nutrition*, 106(4), 1142–1156. <https://doi.org/10.3945/ajcn.117.156349> • Afman, L. A., Lievers, K. J. A., van der Put, N. M. J., Trijbels, F. J. M., & Blom, H. J. (2002). Single nucleotide polymorphisms in the transcobalamin gene: Relationship with transcobalamin concentrations and risk for neural tube defects. *European Journal of Human Genetics*, 10(7), 433–438. <https://doi.org/10.1038/sj.ejhg.5200830> • Castro, R., Barroso, M., Rocha, M., Esse, R., Ramos, R., Ravasco, P., Rivera, I., & de Almeida, I. T. (2010). The TCN2 776CNG polymorphism correlates with vitamin B12 cellular delivery in healthy adult populations. *Clinical Biochemistry*, 43(7–8), 645–649. <https://doi.org/10.1016/j.clinbiochem.2010.01.015> • Guéant, J.-L., Chabi, N. W., Guéant-Rodriguez, R.-M., Mutchinick, O. M., Debard, R., Payet, C., Lu, X., Villalume, C., Bronowicki, J.-P., Quadros, E. V., Sanni, A., Amouzou, E., Xia, B., Chen, M., Anello, G., Bosco, P., Romano, C., Arrieta, H. R., Sánchez, B. E., ... Namour, F. (2007). Environmental influence on the worldwide prevalence of a 776C->G variant in the transcobalamin gene (TCN2). *Journal of Medical Genetics*, 44(6), 363–367. <https://doi.org/10.1136/jmg.2006.048041> • von Castel-Dunwoody, K. M., Kauwell, G. P. A., Shelnutt, K. P., Vaughn, J. D., Griffin, E. R., Maneval, D. R., Theriaque, D. W., & Bailey, L. B. (2005). Transcobalamin 776C->G polymorphism negatively affects vitamin B-12 metabolism. *The American Journal of Clinical Nutrition*, 81(6), 1436–1441. <https://doi.org/10.1093/ajcn/81.6.1436> • Lievers, K. J. A., Afman, L. A., Kluijtmans, L. A. J., Boers, G. H. J., Verhoef, P., den Heijer, M., Trijbels, F. J. M., & Blom, H. J. (2002). Polymorphisms in the transcobalamin gene: Association with plasma homocysteine in healthy individuals and vascular disease patients. *Clinical Chemistry*, 48(9), 1383–1389.

MITOCHONDRIA SNP References

NQO1 rs1800566

• Beyer, R. E., Segura-Aguilar, J., Di Bernardo, S., Cavazzoni, M., Fato, R., Fiorentini, D., Galli, M. C., Setti, M., Landi, L., & Lenaz, G. (1996). The role of DT-diaphorase in the maintenance of the reduced antioxidant form of coenzyme Q in membrane systems. *Proceedings of the National Academy of Sciences of the United States of America*, 93(6), 2528–2532. <https://doi.org/10.1073/pnas.93.6.2528> • Surapaneni, A., Schlosser, P., Zhou, L., Liu, C., Chatterjee, N., Arking, D. E., Dutta, D., Coresh, J., Rhee, E. P., & Grams, M. E. (2022). Identification of 969 protein quantitative trait loci in an African American population with kidney disease attributed to hypertension. *Kidney International*, 102(5), 1167–1177. <https://doi.org/10.1016/j.kint.2022.07.005> • Ross, D., & Siegel, D. (2021). The diverse functionality of NQO1 and its roles in redox control. *Redox Biology*, 41, 101950. <https://doi.org/10.1016/j.redox.2021.101950> • Ross, D., & Siegel, D. (2017). Functions of NQO1 in Cellular Protection and CoQ10 Metabolism and its Potential Role as a Redox Sensitive Molecular Switch. *Frontiers in Physiology*, 8, 595. <https://doi.org/10.3389/fphys.2017.00595> • Pey, A. L., Megarity, C. F., & Timson, D. J. (2014). FAD binding overcomes defects in activity and stability displayed by cancer-associated variants of human NQO1. *Biochimica Et Biophysica Acta*, 1842(11), 2163–2173. <https://doi.org/10.1016/j.bbdis.2014.08.011> • Martínez-Limón, A., Aliquet, M., Lang, W.-H., Calloni, G., Wittig, I., & Vabulas, R. M. (2016). Recognition of enzymes lacking bound cofactor by protein quality control. *Proceedings of the National Academy of Sciences of the United States of America*, 113(43), 12156–12161. <https://doi.org/10.1073/pnas.1611994113> • Fischer, A., Schmelzer, C., Rimbach, G., Niklowitz, P., Menke, T., & Döring, F. (2011). Association between genetic variants in the Coenzyme Q10 metabolism and Coenzyme Q10 status in humans. *BMC Research Notes*, 4, 245. <https://doi.org/10.1186/1756-0500-4-245>

PPARGC1A rs8192678

• Reddy, T. V., Govatati, S., Deenadayal, M., Shivaji, S., & Bhanoori, M. (2018). Polymorphisms in the TFAM and PGC1-? genes and their association with polycystic ovary syndrome among South Indian women. *Gene*, 641, 129–136. <https://doi.org/10.1016/j.gene.2017.10.010> • Qian, L., Zhu, Y., Deng, C., Liang, Z., Chen, J., Chen, Y., Wang, X., Liu, Y., Tian, Y., & Yang, Y. (2024). Peroxisome proliferator-activated receptor gamma coactivator-1 (PGC-1) family in physiological and pathophysiological process and diseases. *Signal Transduction and Targeted Therapy*, 9(1), 50. <https://doi.org/10.1038/s41392-024-01756-w> • Petr, M., Stastny, P., Zajac, A., Tufano, J. J., & Maciejewska-Skrendo, A. (2018). The Role of Peroxisome Proliferator-Activated Receptors and Their Transcriptional Coactivators Gene Variations in Human Trainability: A Systematic Review. *International Journal of Molecular Sciences*, 19(5), 1472. <https://doi.org/10.3390/ijms19051472> • Montes-de-Oca-García, A., Corral-Pérez, J., Velázquez-Díaz, D., Perez-Bey, A., Rebollo-Ramos, M., Marín-Galindo, A., Gómez-Gallego, F., Calderon-Dominguez, M., Casals, C., & Ponce-González, J. G. (2022). Influence of Peroxisome Proliferator-Activated Receptor (PPAR)-gamma Coactivator (PGC)-1 alpha gene rs8192678 polymorphism by gender on different health-related parameters in healthy young adults. *Frontiers in Physiology*, 13, 885185. <https://doi.org/10.3389/fphys.2022.885185> • Konopka, M. J., van den Bundler, J. C. M. L., Rietjens, G., Sperlrich, B., & Zeegers, M. P. (2022). Genetics of long-distance runners and road cyclists-A systematic review with meta-analysis. *Scandinavian Journal of Medicine & Science in Sports*, 32(10), 1414–1429. <https://doi.org/10.1111/sms.14212> • Jannas-Vela, S., & Castro-Sepulveda, M. (2023). Chapter 8—Dietary eicosapentaenoic acid and docosahexaenoic acid for mitochondrial biogenesis and dynamics. In S. M. Ostojic (Ed.), *Molecular Nutrition and Mitochondria* (pp. 213–224). Academic Press. <https://doi.org/10.1016/B978-0-323-90256-4.00028-x>

213–224). Academic Press. <https://doi.org/10.1016/B978-0-323-90256-4.00028-x>

SIRT1 rs1467568

• Zillikens, M. C., van Meurs, J. B. J., Sijbrands, E. J. G., Rivadeneira, F., Dehghan, A., van Leeuwen, J. P. T. M., Hofman, A., van Duijn, C. M., Witterman, J. C. M., Uitterlinden, A. G., & Pols, H. A. P. (2009). SIRT1 genetic variation and mortality in type 2 diabetes: Interaction with smoking and dietary niacin. *Free Radical Biology & Medicine*, 46(6), 836–841. <https://doi.org/10.1016/j.freeradbiomed.2008.12.022> • Zillikens, M. C., van Meurs, J. B. J., Rivadeneira, F., Amin, N., Hofman, A., Oostra, B. A., Sijbrands, E. J. G., Witterman, J. C. M., Pols, H. A. P., van Duijn, C. M., & Uitterlinden, A. G. (2009). SIRT1 genetic variation is related to BMI and risk of obesity. *Diabetes*, 58(12), 2828–2834. <https://doi.org/10.2337/db09-0536> • Pedersen, S. B., Øholm, J., Paulsen, S. K., Bennetzen, M. F., & Richelsen, B. (2008). Low Sirt1 expression, which is upregulated by fasting, in human adipose tissue from obese women. *International Journal of Obesity* (2005), 32(8), 1250–1255. <https://doi.org/10.1038/ijo.2008.78> • Lozano, O., Solís-Castañol, D., Cantú-Casas, S., Mendoza Muraira, P. I., & García-Rivas, G. (2023). Chapter 14 - A review of quercetin delivery through nanovectors: Cellular and mitochondrial effects on noncommunicable diseases. In S. M. Ostojic (Ed.), *Molecular Nutrition and Mitochondria* (pp. 363–382). Academic Press. <https://doi.org/10.1016/B978-0-323-90256-4.00006-0> • Higashibata, T., Wakai, K., Naito, M., Morita, E., Hishida, A., Hamajima, N., Hara, M., Suzuki, S., Hosono, S., Takashima, N., Ohnaka, K., Takada, A., Mikami, H., Watanabe, Y., Uemura, H., Kubo, M., & Tanaka, H. (2016). Effects of self-reported calorie restriction on correlations between SIRT1 polymorphisms and body mass index and long-term weight change. *Gene*, 594(1), 16–22. <https://doi.org/10.1016/j.gene.2016.08.051> • Clark, S. J., Falchi, M., Olsson, B., Jacobson, P., Cauchi, S., Balkau, B., Marre, M., Lantieri, O., Andersson, J. C., Jernås, M., Altman, T. J., Richardson, S., Sjöström, L., Wong, H. Y., Carlsson, L. M. S., Froguel, P., & Walley, A. J. (2012). Association of sirtuin 1 (SIRT1) gene SNPs and transcript expression levels with severe obesity. *Obesity* (Silver Spring, Md.), 20(1), 178–185. <https://doi.org/10.1038/oby.2011.200>

SLC19A1

• Stanis?awska-Sachadyn, A., Mitchell, L. E., Woodside, J. V., Buckley, P. T., Kealey, C., Young, I. S., Scott, J. M., Murray, L., Boreham, C. A., McNulty, H., Strain, J. J., & Whitehead, A. S. (2009). The reduced folate carrier (SLC19A1) c.80G>A polymorphism is associated with red cell folate concentrations among women. *Annals of Human Genetics*, 73(Pt 5), 484–491. <https://doi.org/10.1111/j.1469-1809.2009.00529.x> • Devlin, A. M., Clarke, R., Birks, J., Evans, J. G., & Halsted, C. H. (2006). Interactions among polymorphisms in folate-metabolizing genes and serum total homocysteine concentrations in a healthy elderly population. *The American Journal of Clinical Nutrition*, 83(3), 708–713. <https://doi.org/10.1093/ajcn.83.3.708> • Clinical Annotation for rs1051266 (SLC19A1): methotrexate; Arthritis, Rheumatoid (level 2A Efficacy). (n.d.). PharmGKB. Retrieved January 15, 2024, from <https://www.pharmgkb.org/clinicalAnnotation/1451245360> • Cho, Y., Kim, J. O., Lee, J. H., Park, H. M., Jeon, Y. J., Oh, S. H., Bae, J., Park, Y. S., Kim, O. J., & Kim, N. K. (2015). Association of reduced folate carrier-1 (RFC-1) polymorphisms with ischemic stroke and silent brain infarction. *PLoS One*, 10(2), e0115295. <https://doi.org/10.1371/journal.pone.0115295> • Chang, A., Emery-Fillon, N., de Courcy, G. P., Lambert, D., Pfister, M., Rosenblatt, D. S., & Nicolas, J. P. (2000). A polymorphism (80G>A) in the reduced folate carrier gene and its associations with folate status and homocysteinemia. *Molecular Genetics and Metabolism*, 70(4), 310–315. <https://doi.org/10.1006/mgme.2000.3034> • Cao, L., Wang, Y., Zhang, R., Dong, L., Cui, H., Fang, Y., Zhao, L., Shi, O., & Cai, C. (2018). Association of neural tube defects with gene polymorphisms in one-carbon metabolic pathway. *Child's Nervous System: ChNS: Official Journal of the International Society for Pediatric Neurosurgery*, 34(2), 277–284. <https://doi.org/10.1007/s00381-017-3558-z> • Cai, C., Xiao, R., Van Halm-Lutterodt, N., Zhen, J., Huang, X., Xu, Y., Chen, S., & Yuan, L. (2016). Association of MTHFR, SLC19A1 Genetic Polymorphism, Serum Folate, Vitamin B12 and Hcy Status with Cognitive Functions in Chinese Adults. *Nutrients*, 8(10), 665. <https://doi.org/10.3390/nu8100665> • Yee, S. W., Gong, L., Badagnani, I., Giacomini, K. M., Klein, T. E., & Altman, R. B. (2010). SLC19A1 Pharmacogenomics Summary. *Pharmacogenetics and Genomics*, 20(11), 708–715. <https://doi.org/10.1097/FPC.0b013e32833ca92a>

TFAM rs1937

• Akhmetov, I. I., Popov, D. V., Missina, S. S., Vinogradova, O. L., & Rogozkin, V. A. (2010). [Association of the mitochondrial transcription factor (TFAM) gene polymorphism with physical performance of athletes]. *Fiziologija Cheloveka*, 36(2), 121–125. • Alvarez, V., Corao, A. I., Alonso-Montes, C., Sánchez-Ferrero, E., De Mena, L., Morales, B., García-Castro, M., & Coto, E. (2008). Mitochondrial transcription factor A (TFAM) gene variation and risk of late-onset Alzheimer's disease. *Journal of Alzheimer's Disease: JAD*, 13(3), 275–280. <https://doi.org/10.3233/jad-2008-13305> • Chen, Q., Li, Z.-H., Song, W.-Q., Yao, Y., Zhang, Y.-J., Zhong, W.-F., Zhang, P.-D., Liu, D., Zhang, X.-R., Huang, Q.-M., Zhao, X.-Y., Shi, X.-M., & Mao, C. (2022). Association between single nucleotide polymorphism of rs1937 in TFAM gene and longevity among the elderly Chinese population: Based on the CLHLS study. *BMC Geriatrics*, 22(1), 16. <https://doi.org/10.1186/s12877-021-02655-3> • Günther, C., von Hadeln, K., Müller-Thomsen, T., Alberici, A., Binetti, G., Hock, C., Nitsch, R. M., Stoppe, G., Reiss, J., Gal, A., & Finckh, U. (2004). Possible association of mitochondrial transcription factor A (TFAM) genotype with sporadic Alzheimer disease. *Neuroscience Letters*, 369(3), 219–223. <https://doi.org/10.1016/j.neulet.2004.07.070>

NEUROTRANSMITTERS SNP References

COMT rs4680

• Sardahaee, F. S., Holmen, T. L., Micali, N., & Kvaløy, K. (2017). Effects of single genetic variants and polygenic obesity risk scores on disordered eating in adolescents—The HUNT study. *Appetite*, 118, 8–16. <https://doi.org/10.1016/j.appet.2017.07.003> • Lachman, H. M., Papolos, D. F., Saito, T., Yu, Y. M., Szumlanski, C. L., & Weinshilboum, R. M. (1996). Human catechol-O-methyltransferase pharmacogenetics: Description of a functional polymorphism and its potential application to neuropsychiatric disorders. *Pharmacogenetics*, 6(3), 243–250. <https://doi.org/10.1097/00008571-199606000-00007> • Kumar, P., & Rai, V. (2020). Catechol-O-methyltransferase gene Val158Met polymorphism and obsessive-compulsive disorder susceptibility: A meta-analysis. *Metabolic Brain Disease*, 35(2), 241–251. <https://doi.org/10.1007/s11011-019-00495-0> • Eriksson, A.-L., Suuriniemi, M., Mahonen, A., Cheng, S., & Ohlsson, C. (2005). The COMT val158met polymorphism is associated with early pubertal development, height and cortical bone mass in girls. *Pediatric Research*, 58(1), 71–77. <https://doi.org/10.1203/01.PDR.0000163383.49747.B5> • Dawling, S., Roodi, N., Mernaugh, R. L., Wang, X., & Parf, F. F. (2001). Catechol-O-methyltransferase (COMT)-mediated metabolism of catechol estrogens: Comparison of wild-type and variant COMT isoforms. *Cancer Research*, 61(18), 6716–6722. • Wichers, M., Aguilera, M., Kenis, G., Krabbendam, L., Myin-Germeys, I., Jacobs, N., Peeters, F., Derom, C., Vlietinck, R., Mengelers, R., Delespaul, P., & van Os, J. (2008). The catechol-O-methyl transferase Val158Met polymorphism and experience of reward in the flow of daily life. *Neuropsychopharmacology: Official Publication of the American College of Neuropsychopharmacology*, 33(13), 3030–3036. <https://doi.org/10.1038/sj.npp.1301520> • Tunbridge, E. M., Harrison, P. J., Warden, D. R., Johnston, C., Refsum, H., & Smith, A. D. (2008). Polymorphisms in the catechol-O-methyltransferase (COMT) gene influence plasma total homocysteine levels. *American Journal of Medical Genetics*. Part B, *Neuropsychiatric Genetics: The Official Publication of the International Society of Psychiatric Genetics*, 147B(6), 996–999. <https://doi.org/10.1002/ajmg.b.30700> • Stein, M. B., Fallin, M. D., Schork, N. J., & Gelernter, J. (2005). COMT polymorphisms and anxiety-related personality traits. *Neuropsychopharmacology: Official Publication of the American College of Neuropsychopharmacology*, 30(11), 2092–2102. <https://doi.org/10.1038/sj.npp.1300787> • Stein, D. J., Newman, T. K., Savitz, J., & Ramesar, R. (2006). Warriors versus worriers: The role of COMT gene variants. *CNS Spectrums*, 11(10), 745–748. <https://doi.org/10.1017/s1092852900014863> • Scanlon, P. D., Raymond, F. A., & Weinshilboum, R. M. (1979). Catechol-O-methyltransferase: Thermolabile enzyme in erythrocytes of subjects homozygous for allele for low activity. *Science* (New York, N.Y.), 203(4375), 63–65. <https://doi.org/10.1126/science.758679> • Tunbridge, E. M., Narajos, M., Harrison, C. H., Beresford, C., Cipriani, A., & Harrison, P. J. (2019). Which Dopamine Polymorphisms Are Functional? Systematic Review and Meta-analysis of COMT, DAT, DBH, DDC, DRD1-5, MAOA, MAOB, TH, VMAT1, and VMAT2. *Biological Psychiatry*, 86(6), 608–620. <https://doi.org/10.1016/j.biopsych.2019.05.014>

GAD1 rs3828275

• Darrah, S. D. et al. Genetic Variability in Glutamic Acid Decarboxylase Genes: Associations with Post-traumatic Seizures after Severe TBI. *Epilepsy Res* 103, 180–194 (2013). • Lim, S. W. et al. Genetic Prediction of Antidepressant Drug Response and Nonresponse in Korean Patients. *PLoS One* 9, e107098 (2014).

GAD1 rs769407

• Barakat, A. K. et al. Citalopram-induced pathways regulation and tentative treatment-outcome-predicting biomarkers in lymphoblastoid cell lines from depression patients. *Transl Psychiatry* 10, 210 (2020). • Hettema, J. M. et al. Association between glutamic acid decarboxylase genes and anxiety disorders, major depression, and neuroticism. *Mol Psychiatry* 11, 752–762 (2006). • Utge, S. et al. A population-based association study of candidate genes for depression and sleep disturbance. *Am J Med Genet B Neuropsychiatr Genet* 153B, 468–476 (2010). • Weber, H. et al. Gender Differences in Associations of Glutamate Decarboxylase 1 Gene (GAD1) Variants with Panic Disorder. *PLoS One* 7, e37651 (2012).

MAO-A

• Kolla, N. J., & Bortolotto, M. (2020). The role of monoamine oxidase A in the neurobiology of aggressive, antisocial, and violent behavior: A tale of mice and men. *Progress in Neurobiology*, 194, 101875. <https://doi.org/10.1016/j.pneurobio.2020.101875> • M. B., & Jc, S. (2011). Behavioral outcomes of monoamine oxidase deficiency: Preclinical and clinical evidence. *International Review of Neurobiology*, 100. <https://doi.org/10.1016/B978-0-12-386467-3.00002-9> • Nordquist, N., & Orelund, L. (2006). Monoallelic expression of MAOA in skin fibroblasts. *Biochemical and Biophysical Research Communications*, 348(2), 763–767. <https://doi.org/10.1016/j.bbrc.2006.07.131> • Leuchter, A. F., McCracken, J. T., Hunter, A. M., Cook, I. A., & Alpert, J. E. (2009). Monoamine oxidase A and catechol-O-methyltransferase functional polymorphisms and the placebo response in major depressive disorder. *Journal of Clinical Psychopharmacology*, 29(4), 372–377. <https://doi.org/10.1097/jcp.0b013e3181ac4aa1> • Hwang, I. W., Lim, M. H., Kwon, H. J., & Jin, H. J. (2018). Association of Monoamine Oxidase A (MAOA) Gene uVNTR and rs6323 Polymorphisms with Attention Deficit and Hyperactivity Disorder in Korean Children. *Medicina* (Kaunas, Lithuania), 54(3), 32. <https://doi.org/10.3390/medicina54030032> • Hotamisligil, G. S., & Breakfield, X. O. (1991). Human monoamine oxidase A gene determines levels of enzyme activity. *American Journal of Human Genetics*, 49(2), 383–392. • Wang, M., Li, H., Deater-Deckard, K., & Zhang, W. (2018). Interacting Effect of Catechol-O-Methyltransferase (COMT) and Monoamine Oxidase A (MAOA) Gene Polymorphisms, and Stressful Life Events on Aggressive Behavior in Chinese Male Adolescents. *Frontiers in Psychology*, 9, 1079. <https://doi.org/10.3389/fpsyg.2018.01079> • Larson, C. L., Taubitz, L. E., & Robinson, J. S. (2010). MAOA 941G polymorphism and the time course of emotional recovery following unpleasant pictures. *Psychophysiology*, 47(5), 857–862. <https://doi.org/10.1111/j.1469-8986.2010.01005.x> • Larson, C. L., Taubitz, L. E., & Robinson, J. S. (2010). MAOA 941G polymorphism and the time course of emotional recovery following unpleasant pictures. *Psychophysiology*, 47(5), 857–862. <https://doi.org/10.1111/j.1469-8986.2010.01005.x>

MAO-B

- Löhle, M., Mangone, G., Wolz, M., Beuthien-Baumann, B., Oehme, L., van den Hoff, J., Kotzerke, J., Reichmann, H., Corvol, J.-C., & Storch, A. (2018). Functional monoamine oxidase B gene intron 13 polymorphism predicts putaminal dopamine turnover in de novo Parkinson's disease. *Movement Disorders: Official Journal of the Movement Disorder Society*, 33(9), 1496–1501. <https://doi.org/10.1002/mds.27466>
- Dlugos, A. M., Palmer, A. A., & de Wit, H. (2009). Negative emotionality: Monoamine oxidase B gene variants modulate personality traits in healthy humans. *Journal of Neural Transmission (Vienna, Austria?)*, 116(10), 1323–1334. <https://doi.org/10.1007/s00702-009-0281-2>
- Bortolato, M., & Shih, J. C. (2011). Behavioral outcomes of monoamine oxidase deficiency: Preclinical and clinical evidence. *International Review of Neurobiology*, 100, 13–42. <https://doi.org/10.1016/B978-0-12-386467-3.00002-9>
- Babi? Leko, M., Nikolac Perkovi?, M., Nedi? Erjavec, G., Klepac, N., Švob Štrac, D., Borove?ki, F., Pivac, N., Hof, P. R., & Šimi?, G. (2021). Association of the MAOB rs1799836 Single Nucleotide Polymorphism and APOE ?4 Allele in Alzheimer's Disease. *Current Alzheimer Research*, 18(7), 585–594. <https://doi.org/10.2174/1567205018666210917162843>
- Löhle, M., Mangone, G., Hermann, W., Hausbrand, D., Wolz, M., Mende, J., Reichmann, H., Hermann, A., Corvol, J.-C., & Storch, A. (2022). Functional MAOB Gene Intron 13 Polymorphism Predicts Dyskinesia in Parkinson's Disease. *Parkinson's Disease*, 2022, 5597503. <https://doi.org/10.1155/2022/5597503>